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U.S. Patent & Trademark
Administration

**NON-PEPTIDE GnRH AGENTS, METHODS AND
INTERMEDIATES FOR THEIR PREPARATION**

This application claims the benefit of U.S. Provisional Application No.
5 60/097,520, filed on August 20, 1998.

**TECHNICAL FIELD AND INDUSTRIAL APPLICABILITY OF THE
INVENTION**

This invention relates generally to compounds that affect the action of human
10 gonadotropin-releasing hormone (GnRH). More particularly, it relates to non-peptide
GnRH antagonists or agonists and to their preparation. These non-peptide GnRH
agents have advantageous physical, chemical and biological properties, and are useful
medicaments for diseases or conditions mediated by modulation of the pituitary-
gonadal axis. The compounds of the invention avoid the degradation and
15 biodistribution problems of peptide agents.

BACKGROUND OF THE INVENTION

Gonadotropin-Releasing Hormone (GnRH), also known as luteinizing
hormone-releasing hormone (LH-RH), plays a central role in the biology of
reproduction. A large variety of analogs have been used for an increasing number of
20 clinical indications. The GnRH decapeptide (pyro-Glu-His-Trp-Ser-Tyr-Gly-Leu-Arg-
Pro-Gly-NH₂ or p-EHWSYGLRPG-NH₂) is produced in neurons of the medial basal
hypothalamus from a larger precursor by enzymatic processing. The decapeptide is
released in a pulsatile manner into the pituitary portal circulation system where GnRH
interacts with high-affinity receptors (7-Transmembrane G-Protein Coupled Receptors)
25 in the anterior pituitary gland located at the base of the brain. In the pituitary, GnRH
triggers the release of two gonadotropic hormones (gonadotropins): luteinizing
hormone (LH) and follicle-stimulating hormone (FSH). In testes and ovaries, LH
stimulates the production of testosterone and estradiol, respectively. FSH stimulates
follicle growth in women and sperm formation in men. When correctly functioning,
30 the pulse-timed release and concentration levels of GnRH are critical for the

maintenance of gonadal steroidogenesis and for normal functions of reproduction related to growth and sexual development.

The pituitary response to GnRH varies greatly throughout life. GnRH and the gonadotropins first appear in the fetus at about ten weeks of gestation. The sensitivity to GnRH declines, after a brief rise during the first three months after birth, until the onset of puberty. Before puberty, the FSH response to GnRH is greater than that of LH. Once puberty begins, sensitivity to GnRH increases, and pulsatile LH secretion ensues. Later in puberty and throughout the reproductive years, pulsatile release of GnRH occurs throughout the day, with LH responsiveness being greater than that of FSH. Pulsatile GnRH release results in pulsatile LH and FSH release and hence testosterone and estradiol release from the gonads. After menopause, FSH and LH concentrations rise, and post-menopausal FSH levels are higher than those of LH.

Chronic administration of GnRH agonists and antagonists to animals or to man results in decreased circulating levels of both LH and FSH. GnRH agonists are compounds that mimic endogenous GnRH to stimulate receptors on the pituitary gland, resulting in release of LH and FSH. After a transient rise in gonadal hormone production or “flare” response, chronic administration of GnRH agonists results in a down-regulation of GnRH receptors. GnRH receptor down-regulation and desensitization of the pituitary results in a decrease of circulating levels of LH and FSH. In spite of the symptom-exacerbating hormonal flare experienced, GnRH agonists have been the treatment of choice for sex-steroid-dependent pathophysiologies. For example, GnRH agonists have been used to reduce testosterone production, thereby reducing prostate volume in benign prostatic hyperplasia (BPH) and slowing tumor growth in prostate cancer. These compounds have also been used to treat breast and ovarian cancers.

Recently, GnRH antagonists have become available for clinical evaluation. GnRH antagonists have an immediate effect on the pituitary without the observed flare associated with agonists. Use of GnRH antagonists (usually decapeptides) has been reported in the literature for treatment of breast, ovarian, and prostatic cancers. Other uses of antagonists, like agonists, include endometriosis (including endometriosis with

pain), uterine myoma, ovarian and mammary cystic diseases (including polycystic ovarian disease), prostatic hypertrophy, amenorrhea (e.g., secondary amenorrhea), and precocious puberty. These compounds may also be useful in the symptomatic relief of premenstrual syndrome (PMS). Furthermore, antagonists may be useful to regulate the secretion of gonadotropins in male mammals to arrest spermatogenesis (e.g., as male contraceptives), and for treatment of male sex offenders. Importantly, GnRH antagonists (and agonists) have found utility in treatments where a reversible suppression of the pituitary-gonadal axis is desired.

The presence of GnRH receptors on anterior pituitary cells and several tumor cell types offers the opportunity to develop drugs that act upon these receptors to treat both hormone-dependent and hormone-independent cancers.

For over 50 years, androgen deprivation has been the most effective systematic therapy for the treatment of metastatic carcinoma of the prostate. The rationale is simple--the prostate gland requires androgens for proper growth, maintenance, and function. Yet, prostate cancer and benign prostate hyperplasia are common in men and develop in an environment of continuous androgen exposure. Thus, utilizing a GnRH antagonist to interrupt the pituitary-gonadal axis reduces androgen production and results in tumor growth modulation. Furthermore, GnRH antagonists may have a direct effect on tumor growth by blocking receptors on the tumor cells. For those cancer types that respond both to sex hormones and to GnRH directly, antagonists should be effective in slowing tumor growth by two mechanisms. Since GnRH receptors are present on many prostate and breast cancer cells, it has recently been speculated that GnRH antagonists may also be effective in treating non-hormone-dependent tumors. Recent literature examples indicate that GnRH receptors are present on a number of cancer cell lines, including:

- Prostate Cancer: GnRH agonists exert both *in vitro*, and *in vivo*, a direct inhibitory action on the growth of both androgen-dependent (LNCaP) and androgen-independent (DU 145) human prostatic cancer cell lines. Montagnani et al, *Arch. Ital. Urol. Androl.* 1997, 69(4), 257-263. GnRH antagonist inhibit the growth of

androgen-independent PC-3 prostate cancer in nude mice. Jungwirth et al.,
Prostate **1997**, 32(3), 164-172.

- Ovarian Cancer: The demonstration of GnRH receptors in human ovarian cancers provides a rationale for the use of therapeutic approaches based on GnRH analogues in this malignancy. Srkalovic et al., *Int. J. Oncol.* **1998**, 12(3), 489-498.
- Breast Cancer: Breast cancer is the most common type of cancer in women over the age of 40 and is the leading cause of cancer-related death in women. Systematic endocrine intervention represents a major treatment option for the management of advanced breast cancer, especially with estrogen-dependent cancers. The genes for gonadotropin-releasing hormone and its receptor are expressed in human breast with fibrocystic disease and cancer. Kottler et al., *Int. J. Cancer* **1997**, 71(4), 595-599.

Heretofore, available GnRH antagonists have primarily been peptide analogs of GnRH. See, e.g., International Publication No. WO 93/03058. Peptide antagonists of peptide hormones are often quite potent; however, the use of peptide antagonists is typically associated with problems because peptides are degraded by physiological enzymes and often poorly distributed within the organism being treated. Thus, they have limited effectiveness as drugs. Consequently, there presently exists a need for non-peptide antagonists of the peptide hormone GnRH.

SUMMARY OF THE INVENTION

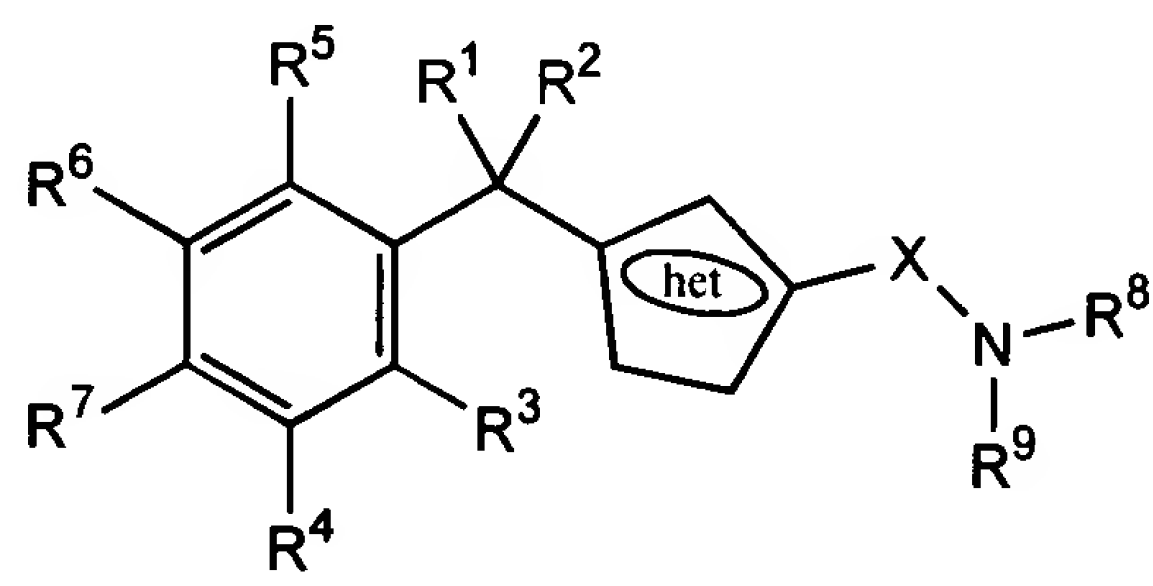
An object of the invention is to develop small-molecule non-peptide GnRH antagonists that exploit both of the above-described mechanisms of action. Non-peptide GnRH agents have advantageous physical, chemical and biological properties compared to peptides, and will be useful medicaments for diseases mediated via the pituitary-gonadal axis and by directly targeting the receptor on tumor cells. There is a need to develop drugs that act upon these receptors to treat both hormone-dependent and hormone-independent cancers.

Another object of the invention is to provide non-peptide compounds that are GnRH agents (agonists or antagonists) that bind to GnRH receptors and thus modulate activity, especially those that are potent GnRH antagonists. Another object of the

invention is to provide effective therapies for individuals needing therapeutic regulation of GnRH and to provide methods for treating diseases and conditions mediated by GnRH regulation.

Such objects have been achieved by the non-peptide GnRH compounds of the invention, which are useful as pharmaceuticals for indications mediated by GnRH regulation. The inventive compounds are pharmaceutically advantageous over peptide compounds since they provide better biodistribution and tolerance to degradation by physiological enzymes. The invention further provides methods of synthesizing the compounds as well as intermediate compounds useful for making the compounds.

The invention is directed to compounds of the general Formula I:



where:

X is selected from C=O, C=S, S=O, and S(O)₂;



is a 5-membered heterocyclic ring containing from 1 to 4, preferably 2 or 3, heteroatoms selected from N, O, and S, wherein the ring may be saturated, partially unsaturated, or fully unsaturated, and may be aromatic;

R¹ and R² are independently selected from H and lower alkyl;

R³ is selected from H, halogen, and substituted and unsubstituted alkyl, alkenyl, alkynyl, cycloalkyl, heterocycle, aryl, heteroaryl, CH₂OR, OR, and C(O)OR, where R is selected from substituted and unsubstituted alkyl, alkenyl, alkynyl, cycloalkyl, heterocycle, aryl, and heteroaryl, and where the total number of carbon atoms present (not including any optional substituents) ranges from 1 to 12;

R⁴ and R⁵ are independently selected from H, halogen, and substituted and unsubstituted alkyl, alkenyl, alkynyl, cycloalkyl, heterocycle, aryl, heteroaryl, CH₂OR,

OR, and C(O)OR, where R is as defined above; and where the total number of carbon atoms present (not including any optional substituents) ranges from 1 to 12;

R⁶ and R⁷ are independently selected from H, halogen, and substituted and unsubstituted alkyl, alkenyl, alkynyl, cycloalkyl, heterocycle, aryl, heteroaryl, CH₂OR, OR, and C(O)OR; where R is as defined above, and where the total number of carbon atoms present (not including any optional substituents) ranges from 1 to 12; or R⁶ and R⁷ taken together with the atoms to which they are bonded form an optionally substituted 5- or 6-membered ring optionally having up to four heteroatoms selected from O, N, and S;

R⁸ is a lipophilic moiety selected from substituted and unsubstituted alkyl, alkenyl, alkynyl, cycloalkyl, heterocycle, aryl, heteroaryl, CH₂OR, OR, and C(O)OR, where R is as defined above, and where the total number of carbon atoms present (not including any optional substituents) ranges from 6 to 20; and

R⁹ is selected from H and substituted and unsubstituted alkyl, preferably lower alkyl.

In some embodiments, R¹ or R² can be -OH or =O, and/or R⁸ can also be hydrogen;

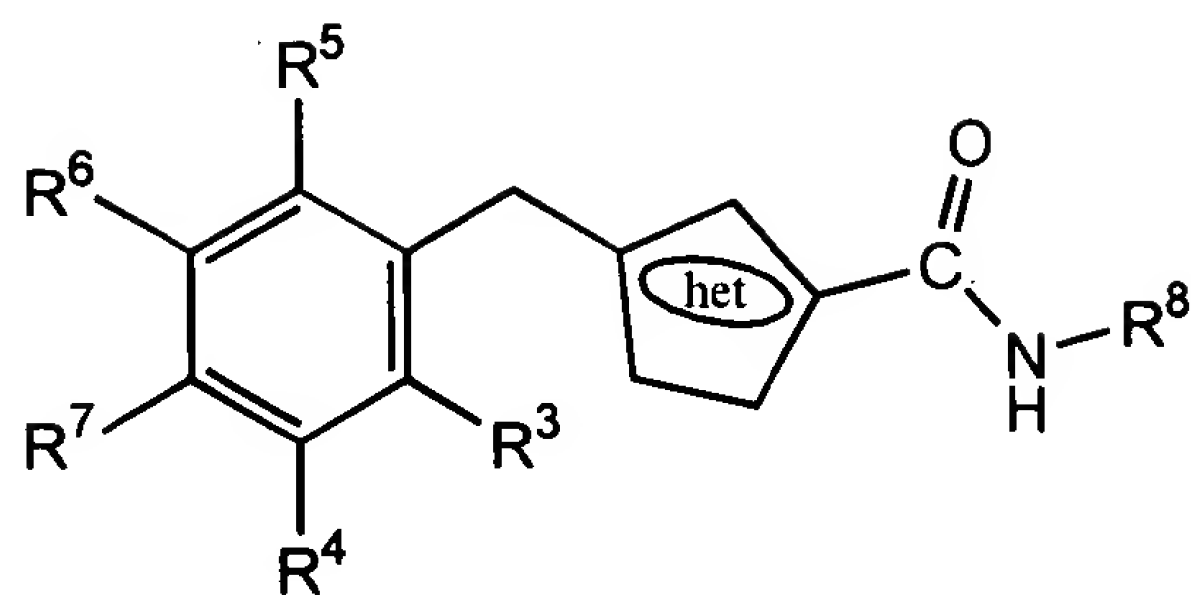
and/or R can be COR or hydrogen; and/or R⁸ can have any desired number of carbon atoms;

and/or R⁸ and R⁹ can also form a ring; and/or any adjacent R groups, such as R⁵ and R⁶ or R³ and R⁴ can form a ring, such as those described for R⁶ and R⁷;

and/or R⁶ can be COR; and/or the (het) group can be substituted or unsubstituted.

Also, in another embodiment, R⁸ and R⁹ can be selected from heterocyclic groups or any compound that forms an amide bond with the nitrogen in Formula I. That is, R⁸ and R⁹ can be any groups starting with a carbon bonded to the nitrogen of general Formula I.

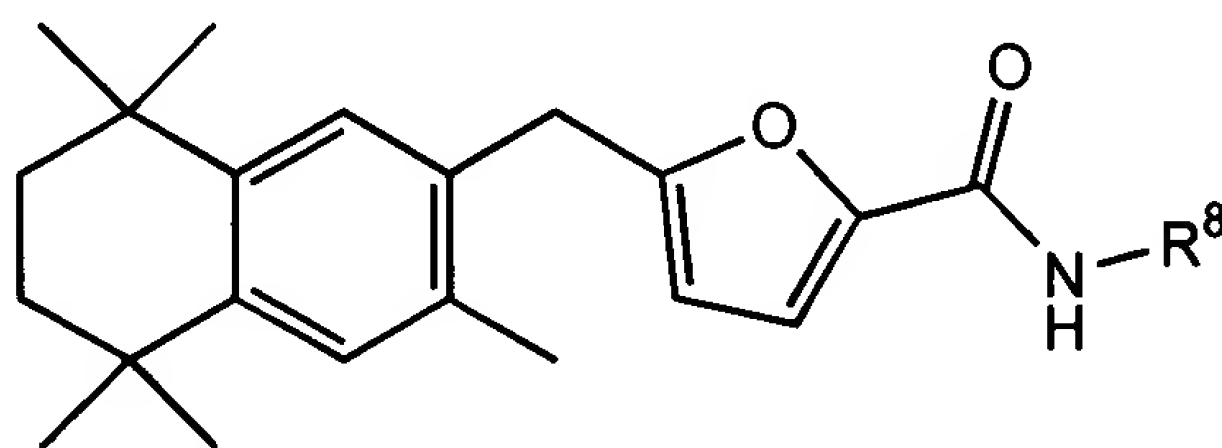
Preferred compounds of the invention are of the general formula II:



II

where the variables in the formula are as defined above.

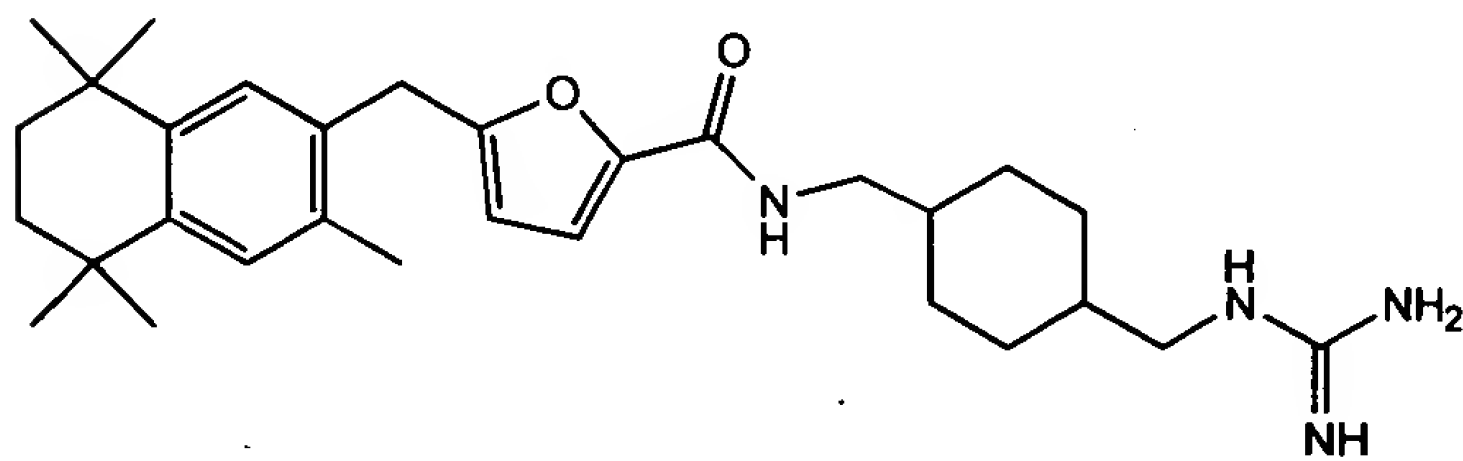
Especially preferred compounds have the formula III:



III

where R^8 is defined above. Preferred R^8 groups include: aryl, $-\text{CH}_2\text{-aryl}$, $-\text{CH}_2\text{-heteroaryl}$, $-\text{CH}_2\text{-cycloalkyl}$, and $-(\text{CH}_2)_n\text{-O-aryl}$ where n is an integer of from 1 to 4.

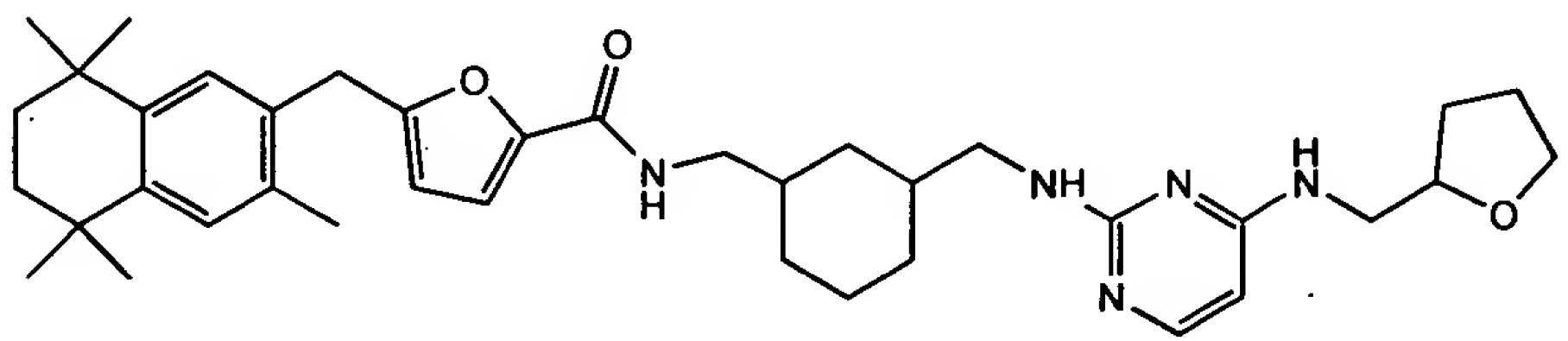
Preferred compounds of the invention include:



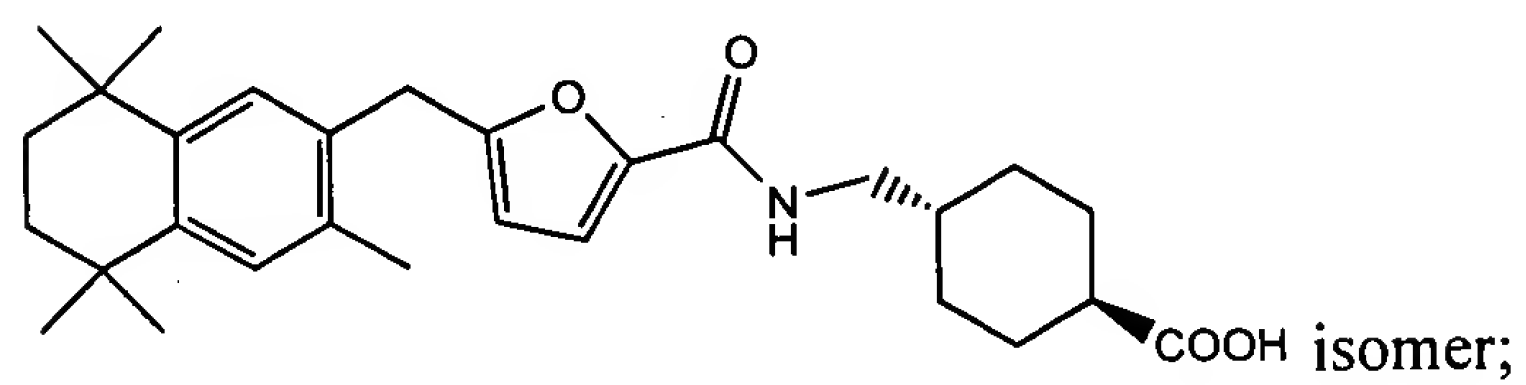
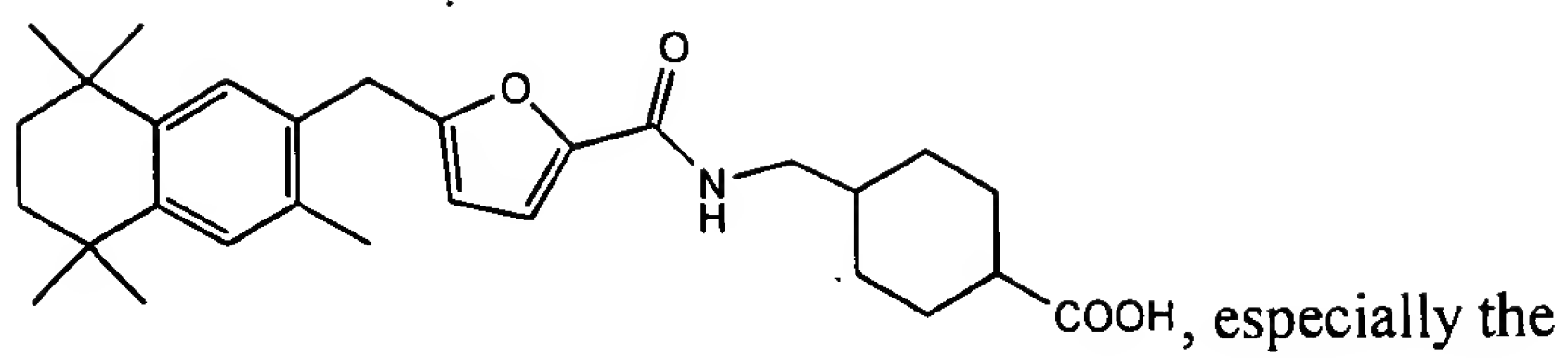
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, including both cis- and trans-

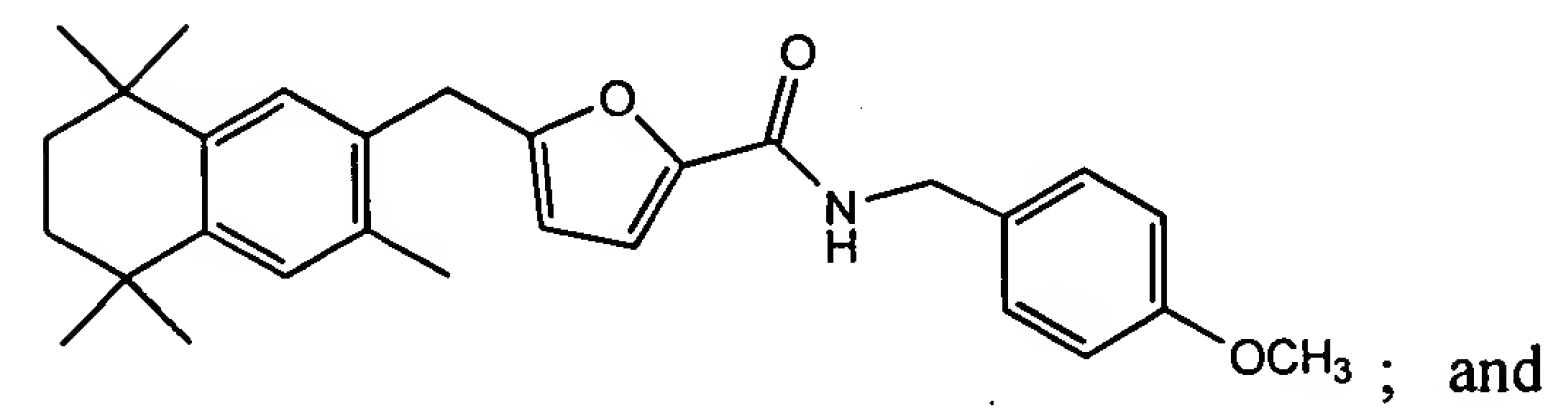
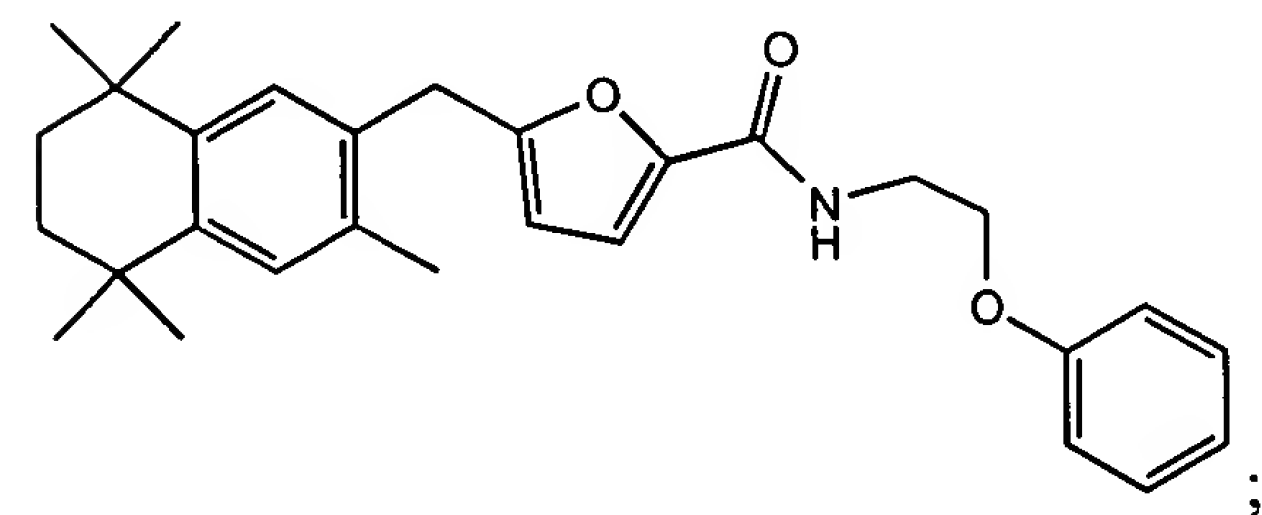
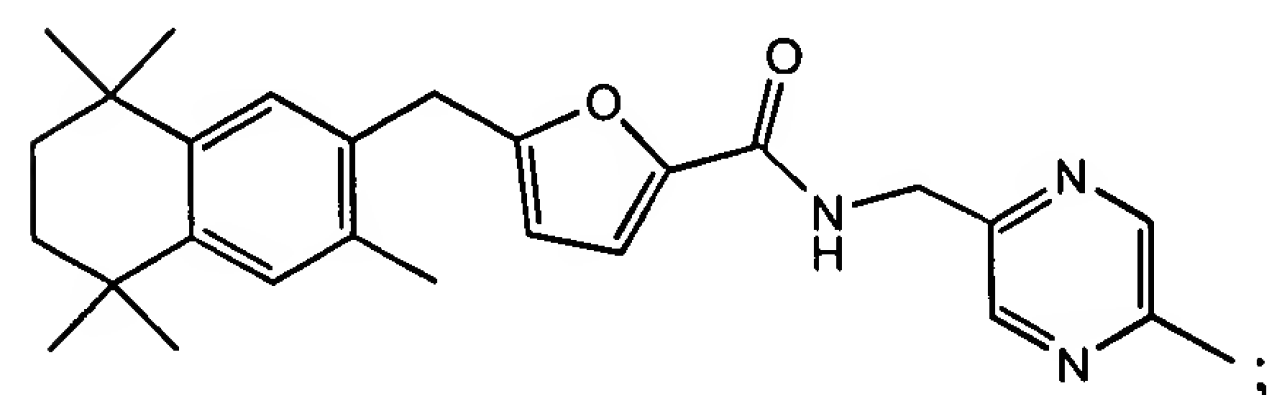
isomers at the cyclohexyl substituent;



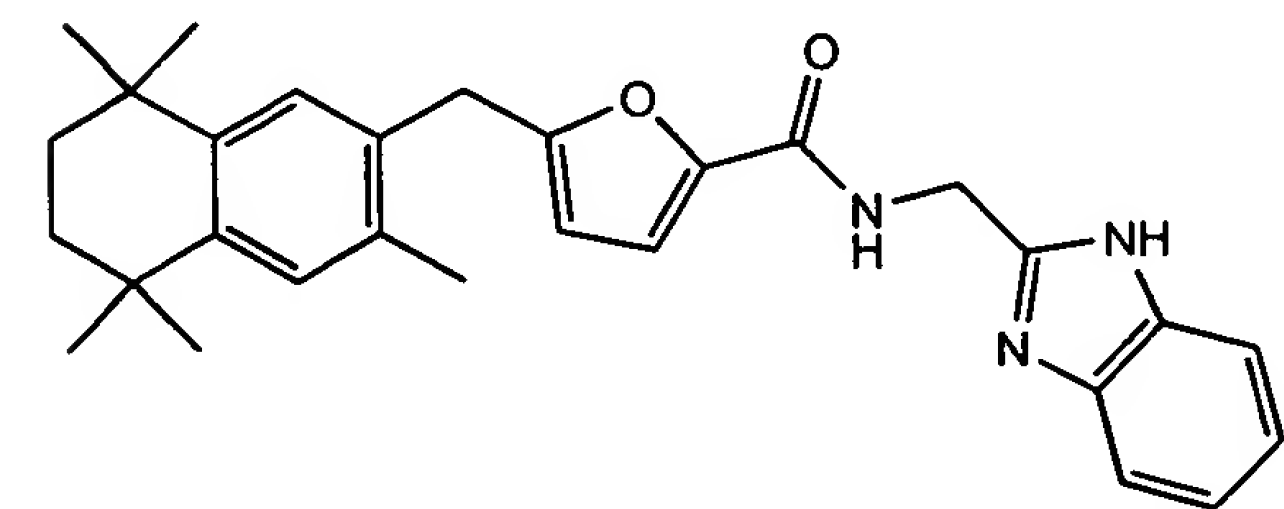
;



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In addition to compounds of the above formulae, GnRH agents of the invention include pharmaceutically acceptable salts, multimeric forms, prodrugs, and active metabolites of such compounds. Such non-peptide agents are pharmaceutically advantageous over peptide agents since they provide better biodistribution and tolerance to degradation by physiological enzymes.

The invention also relates to pharmaceutical compositions comprising a therapeutically effective amount of a GnRH agent of the invention in combination with a pharmaceutically acceptable carrier or diluent. Moreover, the invention relates to methods for regulating the secretion of gonadotropins in mammals, comprising administering therapeutically effective amounts of GnRH agents of the invention.

The invention also relates to methods and intermediates useful for making compounds of the Formula I.

Other features, objects, and advantages of the invention will become apparent from the following detailed description of the invention and its preferred embodiments.

DETAILED DESCRIPTION OF INVENTION AND PREFERRED EMBODIMENTS

Some of the compounds of the invention contain one or more centers of asymmetry, and may thus give rise to enantiomers, diastereoisomers, and other stereoisomeric forms. The invention is meant to include all such possible stereoisomers as well as their racemic and optically pure forms. When the compounds described herein contain olefinic double bonds, they are intended to encompass both E and Z geometric isomers.

The chemical formulae referred to herein may exhibit the phenomenon of tautomerism. As the structural formulae shown in this specification only depict one of the possible tautomeric forms, it should be understood that the invention nonetheless encompasses all tautomeric forms.

The term "alkyl" refers to straight- and branched-chain alkyl groups having one to twelve carbon atoms. Exemplary alkyl groups include methyl (Me), ethyl, n-propyl, isopropyl, butyl, isobutyl, sec-butyl, tert-butyl (tBu), pentyl, isopentyl, tert-pentyl,

hexyl, isohexyl, and the like. The term “lower alkyl” designates an alkyl having from 1 to 8 carbon atoms (a C₁₋₈-alkyl). Suitable substituted alkyls include fluoromethyl, difluoromethyl, trifluoromethyl, 2-fluoroethyl, 3-fluoropropyl, hydroxymethyl, 2-hydroxyethyl, 3-hydroxypropyl, and the like.

5 The term “alkenyl” refers to straight- and branched-chain alkenyl groups having from 2 to 12 carbon atoms. Illustrative alkenyl groups include prop-2-enyl, but-2-enyl, but-3-enyl, 2-methylprop-2-enyl, hex-2-enyl, and the like.

 The term “alkynyl” refers to straight- and branched-chain alkynyl groups having from 2 to 12 carbons atoms. Exemplary alkynyls include prop-2-ynyl, 3-
10 methylpent-4-ynyl, hex-2-ynyl, and the like.

 The term “carbocycle” refers to a monocyclic or polycyclic carbon ring structure (with no heteroatoms) having from 3 to 7 carbon atoms in each ring, which may be saturated, partially saturated, or unsaturated. Exemplary carbocycles include cycloalkyls and aryls.

15 The term “heterocycle” refers to a monocyclic or polycyclic ring structure with one or more heteroatoms selected from N, O, and S, and having from 3 to 7 atoms (carbon atoms plus any heteroatom(s)) in each ring, which may be saturated, partially saturated, or unsaturated. Exemplary heterocycles include tetrahydrofuranyl, tetrahydropyranyl, azetidiny, pyrrolidiny, piperidiny, piperazinyl, and the like.

20 The term “cycloalkyls” as used herein refers to saturated carbocycles having 3 to 12 carbons, including bicyclic and tricyclic cycloalkyl structures. Suitable cycloalkyls include cyclopropyl, cyclobutyl, cyclopentyl, cyclohexyl, cycloheptyl, and the like.

 The terms “aryls” and “heteroaryls” refer to monocyclic and polycyclic
25 unsaturated or aromatic ring structures, with “aryl” referring to those that are carbocycles and “heteroaryl” referring to those that are heterocycles. Examples of aromatic ring structures include phenyl, naphthyl, 1,2,3,4-tetrahydronaphthyl, furyl, thienyl, pyrrolyl, pyridyl, pyridinyl, pyrazolyl, imidazolyl, pyrazinyl, pyridazinyl, 1,2,3-triazinyl, 1,2,4-oxadiazolyl, 1,3,4-oxadiazolyl, 1-H-tetrazol-5-yl, indolyl, quinolinyl,
30 benzofuranyl, benzothiophenyl (thianaphthenyl), and the like. Such moieties may be

optionally substituted by one or more suitable substituents, for example, a substituent selected from a halogen (F, Cl, Br or I); lower alkyl; OH; NO₂; CN; CO₂H; O-lower alkyl; aryl; aryl-lower alkyl; CO₂CH₃; CONH₂; OCH₂CONH₂; NH₂; SO₂NH₂; OCHF₂; CF₃; OCF₃; and the like. Such moieties may also be optionally substituted by a fused-
5 ring structure or bridge, for example OCH₂-O.

The term "aryl-lower alkyl" means a lower alkyl bearing an aryl. Examples include benzyl, phenethyl, pyridylmethyl, naphthylmethyl, and the like. The aryl-lower alkyl may be optionally substituted.

In general, the various moieties or functional groups for variables in Formula I
10 may be optionally substituted by one or more suitable substituents. Exemplary substituents include a halogen (F, Cl, Br, or I), lower alkyl, -OH, -NO₂, -CN, -CO₂H, -O-lower alkyl, -aryl, -aryl-lower alkyl, -CO₂CH₃, -CONH₂, -OCH₂CONH₂, -NH₂, -SO₂NH₂, haloalkyl (e.g., -CF₃, -CH₂CF₃), -O-haloalkyl (e.g., -OCF₃, -OCHF₂), and the like.

15 In addition to compounds of the Formula I, GnRH agents of the invention include pharmaceutically acceptable salts, multimeric forms, prodrugs, and active metabolites of compounds of the Formula I. Such non-peptide agents are pharmaceutically advantageous over peptide agents since they provide better biodistribution and tolerance to degradation by physiological enzymes.

20 Additionally, Formula I is intended to cover, where applicable, solvated as well as unsolvated forms of the compounds. Thus, Formula I includes compounds having the indicated structure, including the hydrated as well as the non-hydrated forms.

As indicated above, GnRH agents in accordance with the invention also include active tautomeric and stereoisomeric forms of the compounds of the Formula I, which
25 may be readily obtained using techniques known in the art. For example, optically active (R) and (S) isomers may be prepared via a stereospecific synthesis, e.g., using chiral synthons and chiral reagents, or racemic mixtures may be resolved using conventional techniques.

GnRH agents further include multivalent or multimeric forms of active forms
30 of the compounds of the Formula I. Such "multimers" may be made by linking or

placing multiple copies of an active compound in close proximity to each other, e.g., using a scaffolding provided by a carrier moiety. Multimers of various dimensions (i.e., bearing varying numbers of copies of an active compound) may be tested to arrive at a multimer of optimum size with respect to receptor binding. Provision of such multivalent forms of active receptor-binding compounds with optimal spacing between the receptor-binding moieties may enhance receptor binding (see, for example, Lee et al., *Biochem.*, 1984, 23:4255). The artisan may control the multivalency and spacing by selection of a suitable carrier moiety or linker units. Useful moieties include molecular supports containing a multiplicity of functional groups that can be reacted with functional groups associated with the active compounds of the invention. A variety of carrier moieties may be used to build highly active multimers, including proteins such as BSA (bovine serum albumin) or HAS, peptides such as pentapeptides, decapeptides, pentadecapeptides, and the like, as well as non-biological compounds selected for their beneficial effects on absorbability, transport, and persistence within the target organism. Functional groups on the carrier moiety, such as amino, sulfhydryl, hydroxyl, and alkylamino groups, may be selected to obtain stable linkages to the compounds of the invention, optimal spacing between the immobilized compounds, and optimal biological properties.

Additionally, GnRH agents of the invention include pharmaceutically acceptable salts of compounds of the Formula I. The term "pharmaceutically acceptable" refers to salt forms that are pharmacologically acceptable and substantially non-toxic to the subject being administered the GnRH agent. Pharmaceutically acceptable salts include conventional acid-addition salts or base-addition salts formed from suitable non-toxic organic or inorganic acids or inorganic bases. Exemplary acid-addition salts include those derived from inorganic acids such as hydrochloric acid, hydrobromic acid, hydroiodic acid, sulfuric acid, sulfamic acid, phosphoric acid, and nitric acid, and those derived from organic acids such as p-toluenesulfonic acid, methanesulfonic acid, ethane-disulfonic acid, isethionic acid, oxalic acid, p-bromophenylsulfonic acid, carbonic acid, succinic acid, citric acid, benzoic acid, 2-acetoxybenzoic acid, acetic acid, phenylacetic acid, propionic acid, glycolic acid,

stearic acid, lactic acid, malic acid, tartaric acid, ascorbic acid, maleic acid, hydroxymaleic acid, glutamic acid, salicylic acid, sulfanilic acid, and fumaric acid. Exemplary base-addition salts include those derived from ammonium hydroxides (e.g., a quaternary ammonium hydroxide such as tetramethylammonium hydroxide), those
5 derived from inorganic bases such as alkali or alkaline earth-metal (e.g., sodium, potassium, lithium, calcium, or magnesium) hydroxides, and those derived from organic bases such as amines, benzylamines, piperidines, and pyrrolidines.

The term “prodrug” refers to a metabolic precursor of a compound of the Formula I (or a salt thereof) that is pharmaceutically acceptable. A prodrug may be
10 inactive when administered to a subject but is converted *in vivo* to an active compound of the Formula I. The term “active metabolite” refers to a metabolic product of a compound of the Formula I that is pharmaceutically acceptable and effective. Prodrugs and active metabolites of compounds of the Formula I may be determined using techniques known in the art.

15 A variety of known assays and techniques may be employed to determine the level of activity of various forms of the compounds in the GnRH system. Ligand-binding assays are used to determine interaction with the receptor of interest. Where binding is of interest, a labeled receptor may be used, where the label is a fluorescer, enzyme, radioisotope, or the like, which registers a quantifiable change upon the
20 binding of the receptor. Alternatively, the artisan may provide for an antibody to the receptor, where the antibody is labeled, which may allow for amplification of the signal. Binding may also be determined by competitive displacement of a ligand bound to the receptor, where the ligand is labeled with a detectable label. Where agonist and/or antagonist activity is of interest, an intact organism or cell may be
25 studied, and the change in an organismic or cellular function in response to the binding of the compound of interest may be measured. Various devices are available for detecting cellular response, such as a microphysiometer available from Molecular-Devices, Redwood City, California. *In vitro* and *in vivo* assays useful in measuring GnRH antagonist activity are known in the art. See, e.g., Bowers et al., “LH
30 suppression in cultured rat pituitary cells treated with 1 ng of LHRH,” *Endocrinology*,

1980, 106:675-683 (*in vitro*,) and Corbin et al., "Antioviulatory activity (AOA) in rats," *Endocr. Res. Commun.* 1975, 2:1-23 (*in vivo*). Particular test protocols that may be used are described below.

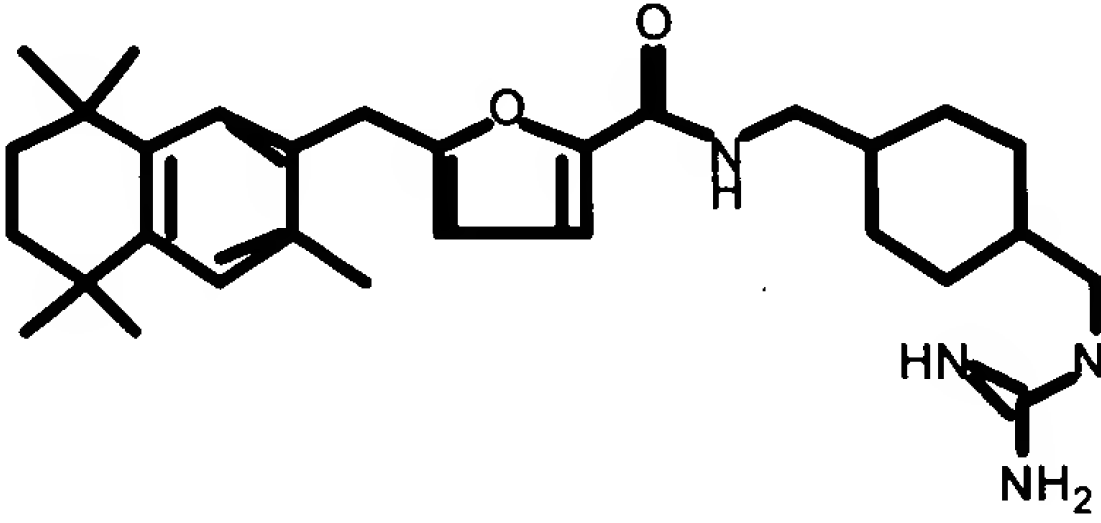
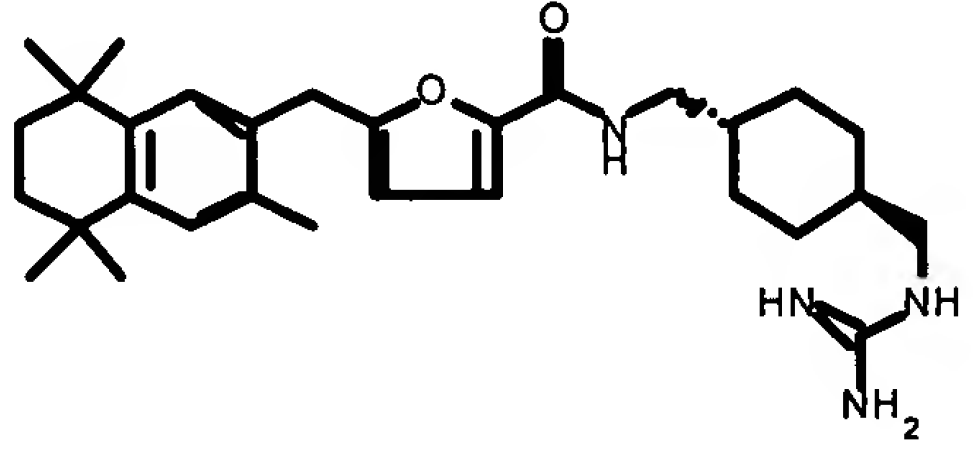
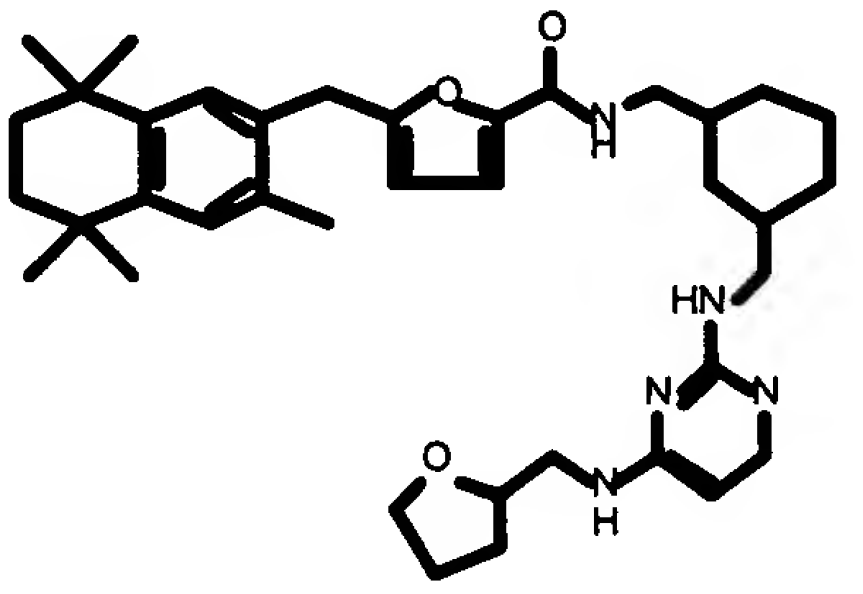
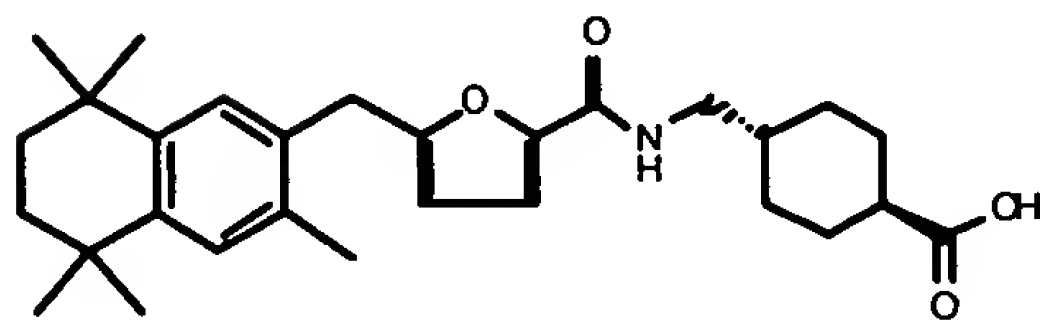
For example, GnRH-receptor antagonists may be functionally assessed by measurement of change in extracellular acidification rates as follows. The ability of compounds to block the extracellular rate of acidification mediated by GnRH in HEK 293 cells expressing human GnRH receptors is determined as a measure of the compound's antagonist activity *in vitro*. Approximately 100,000 cells/chamber are immobilized in agarose suspension medium (Molecular Devices) and perfused with unbuffered MEM media utilizing the Cytosensor® Microphysiometer (Molecular Devices). Cells are allowed to equilibrate until the basal acidification rate remains stable (approximately one hour). Control dose-response curves are performed to GnRH (10^{-11} M to 10^{-7} M). Compounds are allowed to incubate 15 minutes prior to stimulation with GnRH, and are assessed for antagonist activity. After incubation with test compounds, repeat dose-response curves to GnRH in the presence or absence of various concentrations of the test compounds are obtained. Schild regression analysis is performed on compounds to determine whether compounds antagonize GnRH-mediated increases in extracellular acidification rates through a competitive interaction with the GnRH receptor.

In another test, accumulation of total inositol phosphates may be measured by formic acid extraction from cells, followed by separation of the phosphates on Dowex columns. Cells are split using trypsin into two 12-well plates and pre-labeled with 3 H-myoinositol (0.5 Ci - 2 mCi per mL) for 16-18 hours in inositol-free medium. The medium is then aspirated and the cells rinsed with either 1X HBSS, 20 mM HEPES (pH 7.5), or serum-free DMEM, 1X HBSS, 20mM HEPES (pH 7.5) containing agonist, and 20mM LiCl is then added and the cells are incubated for the desired time. The medium is aspirated and the reaction stopped by addition of ice-cold 10mM formic acid, which also serves to extract cellular lipids. Inositol phosphates are separated by ion-exchange chromatography on Dowex columns, which are then washed with 5 mL of 10mM myoinositol and 10mM formic acid. The columns are

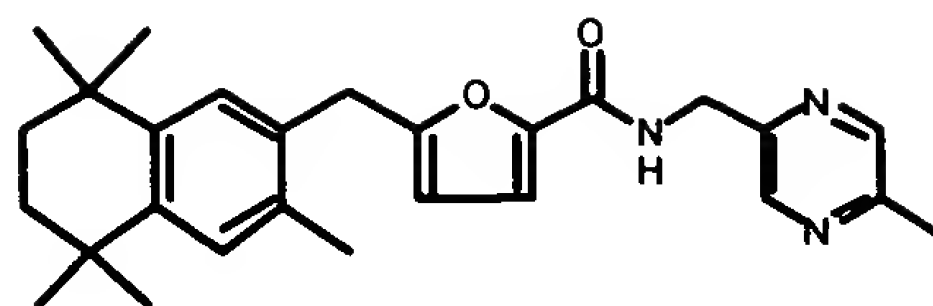
then washed with 10 mL of 60mM sodium formate and 5mM borax, and total inositol phosphates are eluted with 4.5 mL 1M ammonium formate, 0.1M formic acid.

Preferred GnRH agents of the invention include those having a K_i value of about 10 μ M or less. Especially preferred GnRH agents are those having a K_i value in the nanomolar range.

Preferred compounds of the inventions are shown in the following table:

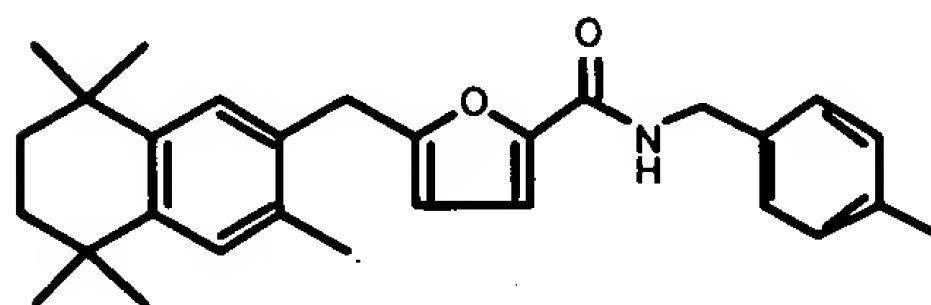
COMPOUND NO.	STRUCTURAL FORMULA	mol. weight
<u>9</u>		492.704
<u>10</u>		492.704
<u>11</u>		627.869
<u>12</u>		465.63

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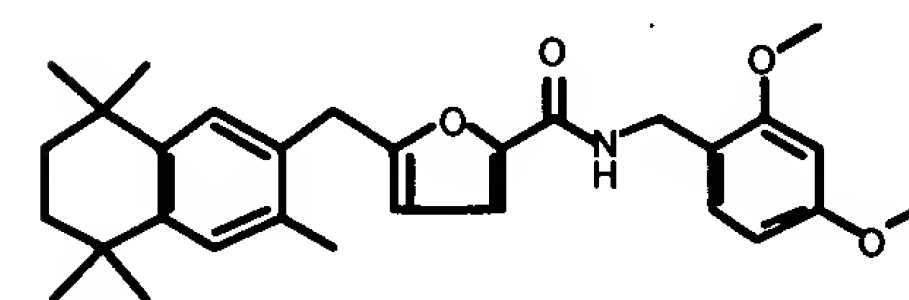
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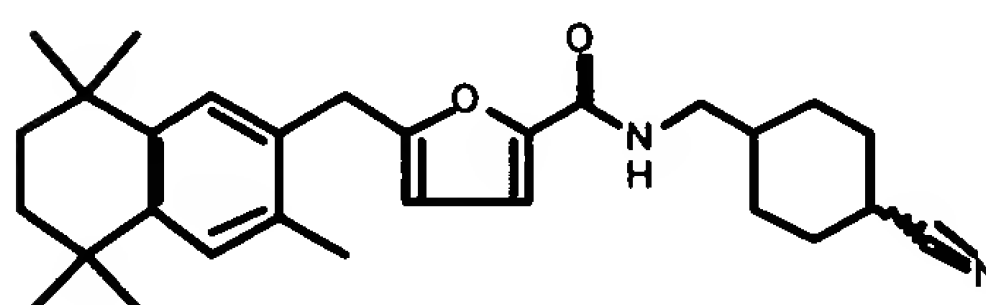
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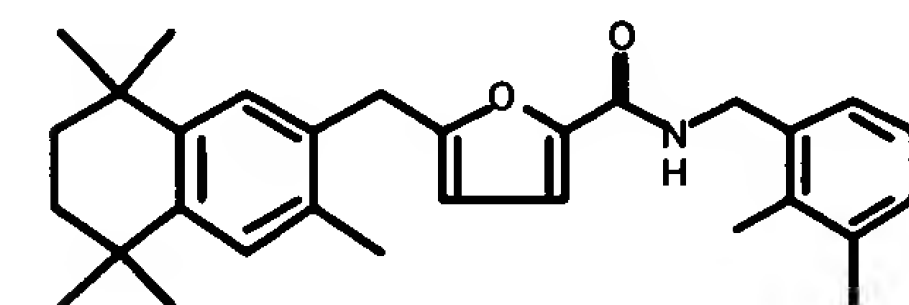
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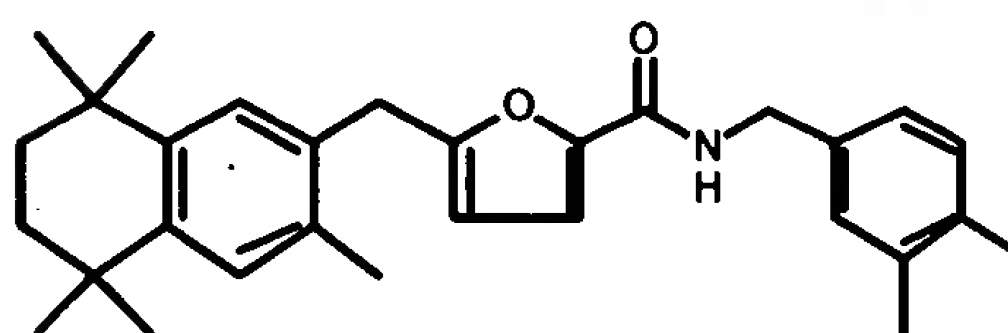
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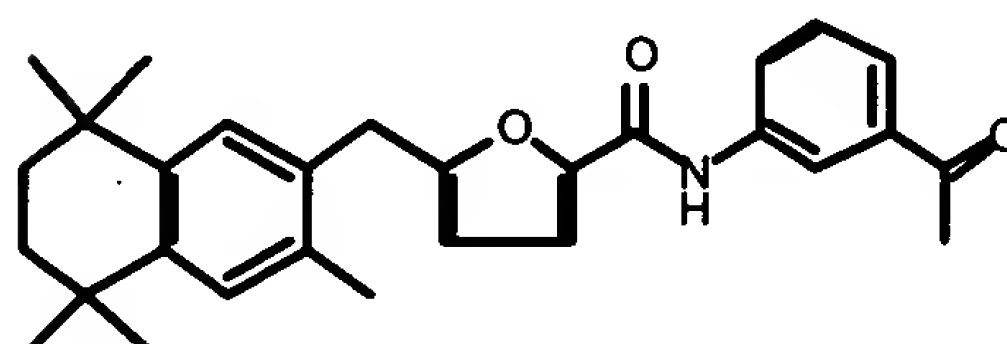
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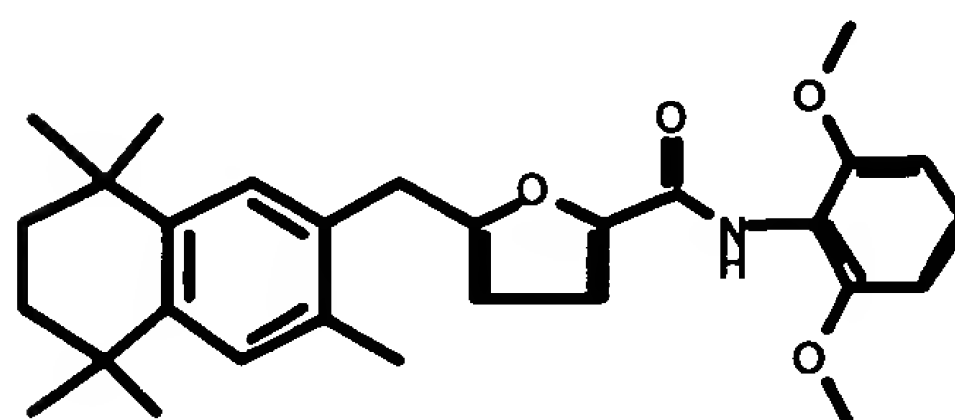
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Pharmaceutical compositions according to the invention comprise an effective GnRH-suppressing amount of at least one GnRH agent according to the invention and an inert or pharmaceutically acceptable carrier or diluent. These compositions may be prepared in a unit-dosage form appropriate for the desired mode of administration, e.g., parenteral or oral.

To treat diseases or conditions mediated by GnRH agonism or antagonism, a pharmaceutical composition of the invention is administered in a suitable formulation prepared by combining a therapeutically effective amount (i.e., a GnRH-modulating amount effective to achieve therapeutic efficacy) of at least one GnRH agent of the invention (as an active ingredient) with one or more pharmaceutically suitable carriers or diluents. Such formulations may be prepared according to conventional procedures, e.g., by appropriately mixing, granulating, and compressing or dissolving the ingredients in known manners. Optionally, one or more different active ingredients, such as different GnRH antagonists, may be employed in a pharmaceutical composition.

The pharmaceutical carrier may be either a solid or liquid. Exemplary solid carriers include lactose, sucrose, talc, gelatin, agar, pectin, acacia, magnesium stearate, stearic acid, and the like. Illustrative of liquid carriers are syrup, peanut oil, olive oil, water, and the like. Similarly, the carrier or diluent may include time-delay or time-release materials known in the art, such as glyceryl monostearate or glyceryl distearate, alone or in combination with a wax, ethylcellulose, hydroxypropylmethylcellulose, methylmethacrylate, or the like.

A variety of pharmaceutical forms can be employed. For example, if a solid carrier is used, the preparation may be in the form of a tablet, hard-gelatin capsule, powder, pellet, troche, or lozenge. The amount of solid carrier may vary widely, with an exemplary amount ranging from about 25 mg to about 1 g. If a liquid carrier is used, the preparation may be in the form of a syrup, emulsion, soft-gelatin capsule, sterile injectable solution, suspension in an ampoule or vial, or non-aqueous liquid suspension.

To obtain a stable, water-soluble dosage form, a pharmaceutically acceptable salt of a compound of Formula I may be dissolved in an aqueous solution of an organic or inorganic acid, such as 0.3M solution of succinic acid or, more preferably, citric acid. If a soluble salt form is not available, the agent may be dissolved in one or more
5 suitable cosolvents. Examples of suitable cosolvents include alcohol, propylene glycol, polyethylene glycol 300, polysorbate 80, glycerin, and the like in concentrations ranging from 0% to 60% of the total volume. In an exemplary embodiment, a compound of Formula I is dissolved in DMSO and diluted with water. The composition may also be in the form of a solution of a salt form of a compound of the
10 Formula I in an appropriate aqueous vehicle, such as water, or isotonic saline or dextrose solutions.

The pharmaceutical compositions of the present invention may be manufactured using conventional techniques, e.g., mixing, dissolving, granulating, dragee-making, levigating, emulsifying, encapsulating, entrapping, or lyophilizing
15 processes. Pharmaceutical compositions may be formulated in a conventional manner using one or more physiologically acceptable carriers comprising excipients or auxiliaries selected to facilitate processing of the active compounds into pharmaceutical preparations. An appropriate formulation is selected in view of the route of administration chosen.

20 For preparing injectable preparations, the agents of the invention may be formulated in aqueous solutions, preferably in physiologically compatible buffers such as Hanks's solution, Ringer's solution, or physiological saline buffer. For transmucosal administration, penetrants appropriate to the barrier to be permeated are used in the formulation and may be selected from those known in the art.

25 For oral administration, the agents may be formulated readily by combining the active ingredient(s) with pharmaceutically acceptable carriers known in the art. Such carriers enable the compounds of the invention to be formulated as tablets, pills, dragees, capsules, liquids, gels, syrups, slurries, suspensions, and the like, for oral ingestion by a patient to be treated. Pharmaceutical preparations for oral use can be
30 obtained by combining one or more agents with a solid excipient, optionally grinding

the resulting mixture into granules, and processing the mixture of granules after adding suitable auxiliaries, if desired, to obtain tablets or dragee cores. Suitable excipients include fillers such as sugars (e.g., lactose, sucrose, mannitol, or sorbitol) and cellulose preparations (e.g., maize starch, wheat starch, rice starch, potato starch, gelatin, gum, methyl cellulose, hydroxypropylmethylcellulose, sodium carboxymethylcellulose, and/or polyvinylpyrrolidone (PVP)). If desired, disintegrating agents may be added, such as cross-linked PVP, agar, or alginic acid or a salt thereof such as sodium alginate.

Dragee cores are provided with suitable coatings. For this purpose, concentrated sugar solutions may be used, which may optionally contain gum arabic, PVP, CarbopolTM gel, polyethylene glycol, titanium dioxide, lacquer solutions, and/or one or more suitable organic solvents. Dyestuffs or pigments may be added to the tablets or dragee coatings for identification or to characterize different combinations of active compound doses.

Pharmaceutical forms that are suitable for oral administration include push-fit capsules made of gelatin, as well as soft, sealed capsules made of gelatin and a plasticizer, such as glycerol or sorbitol. The push-fit capsules may contain the active ingredient(s) in admixture with one or more fillers such as lactose, binders such as starches, and/or lubricants such as talc or magnesium stearate, and, optionally, stabilizers. In soft capsules, the active compound may be dissolved or suspended in a suitable liquid, such as fatty oil, liquid paraffin, or liquid polyethylene glycol. In addition, stabilizers may be added. For buccal administration, the compositions may take the form of tablets or lozenges formulated in a conventional manner.

For administration by inhalation, the compounds for use according to the present invention may be conveniently delivered in the form of an aerosol spray presentation from pressurized packs or a nebuliser, with the use of a suitable propellant, e.g., dichlorodifluoromethane, trichlorofluoromethane, dichlorotetrafluoroethane, carbon dioxide, or another suitable gas. In the case of a pressurized aerosol, the dosage unit may be determined by providing a valve to deliver a metered amount. Capsules and cartridges of, e.g., gelatin for use in an inhaler or

insufflator may be formulated containing a powder mix of the agent and a suitable powder base such as lactose or starch.

The agents may be formulated for parenteral administration by injection, e.g., by bolus injection or continuous infusion. Formulations for injection may be prepared
5 in unit-dosage form, e.g., in ampoules, or in multi-dose containers, with an added preservative. The compositions may take such forms as suspensions, solutions, or emulsions in oily or aqueous vehicles, and may contain formulatory agents such as suspending, stabilizing, and/or dispersing agents.

Pharmaceutical formulations for parenteral administration include aqueous
10 solutions of the active compounds in water-soluble form. Additionally, suspensions of the active compounds may be prepared as appropriate oily injection suspensions. Suitable lipophilic solvents or vehicles include fatty oils such as sesame oil, or synthetic fatty acid esters such as ethyl oleate or triglycerides, or liposomes. Aqueous injectable suspensions may contain substances increasing the viscosity of the
15 suspension, such as sodium carboxymethyl cellulose, sorbitol, or dextran. Optionally, the suspension may also contain suitable stabilizers or agents increasing the solubility of the compounds to allow for the preparation of highly concentrated solutions.

Alternatively, the active ingredient may be in powder form for constitution with a suitable vehicle, e.g., sterile pyrogen-free water, before use. The compounds may
20 also be formulated as rectal compositions, such as suppositories or retention enemas, e.g., containing conventional suppository bases such as cocoa butter or other glycerides.

In addition to the formulations described above, the compounds may also be formulated as a depot preparation. Such long-acting formulations may be administered
25 by implantation (for example, subcutaneously or intramuscularly) or by intramuscular injection. Thus, for example, the compounds may be formulated with suitable polymeric or hydrophobic materials (for example, as an emulsion in an acceptable oil) or ion-exchange resins, or as sparingly soluble derivatives, for example, as a sparingly soluble salt.

An exemplary pharmaceutical carrier for the hydrophobic compounds of the invention is a cosolvent system comprising benzyl alcohol, a nonpolar surfactant, a water-miscible organic polymer, and an aqueous phase. The cosolvent system may be the VPD co-solvent system (VPD is a solution of 3% w/v benzyl alcohol, 8% w/v of the nonpolar surfactant polysorbate 80, and 65% w/v polyethylene glycol 300, made up to volume in absolute ethanol). The VPD co-solvent system (VPD:5W) is comprised of VPD diluted 1:1 with a 5% dextrose-in-water solution. This co-solvent system dissolves hydrophobic compounds well, and the resulting formulation produces low toxicity upon systemic administration. As will be apparent, the proportions of a suitable co-solvent system may be varied in light of the solubility and toxicity characteristics. Furthermore, the identity of the co-solvent components may be varied: for example, other low-toxicity nonpolar surfactants may be used instead of polysorbate 80; the fraction size of polyethylene glycol may be varied; one or more other biocompatible polymers (e.g., PVP) may be added or replace polyethylene glycol; and other sugars or polysaccharides may be substituted for dextrose.

Alternatively, other delivery systems for hydrophobic pharmaceutical compounds may be employed. Liposomes and emulsions are known examples of delivery vehicles or carriers for hydrophobic drugs and may be used to formulate suitable preparations. Certain organic solvents such as dimethylsulfoxide also may be employed, although this may cause an increase in toxicity. Additionally, delivery may be achieved using a sustained-release system, such as semipermeable matrices of solid hydrophobic polymers containing the therapeutic agent. Various sustained-release materials are available and known to those skilled in the art. Sustained-release capsules may, depending on their chemical nature, release the compounds for a period lasting from a few weeks or up to over 100 days. Depending on the chemical nature and the biological stability of the therapeutic agent, additional techniques for protein stabilization may be readily employed.

The pharmaceutical compositions also may comprise suitable solid- or gel-phase carriers or excipients. Examples of such carriers or excipients include calcium

carbonate, calcium phosphate, sugars, starches, cellulose derivatives, gelatin, and polymers such as polyethylene glycols.

Some of the compounds of the invention may be provided as salts with pharmaceutically compatible counter-ions. Pharmaceutically acceptable salts may be
5 formed with many acids, including hydrochloric, sulfuric, acetic, lactic, tartaric, malic, succinic, and like acids. Salts tend to be more soluble in aqueous or other protonic solvents than are the corresponding free-base forms.

It will be appreciated that the actual dosages of the agents used in the compositions of this invention will vary according to the particular complex being
10 used, the particular composition formulated, the mode of administration, and the particular site, host, and disease being treated. Optimal dosages for a given set of conditions may be ascertained by those skilled in the art using conventional dosage-determination tests in view of the experimental data for a given compound. For oral administration, an exemplary daily dose generally employed will be from about 0.001
15 to about 1000 mg/kg of body weight, with courses of treatment repeated at appropriate intervals. Administration of prodrugs may be dosed at weight levels that are chemically equivalent to the weight levels of the fully active compounds.

Examples of specific pharmaceutical preparations in accordance with the invention are provided below.

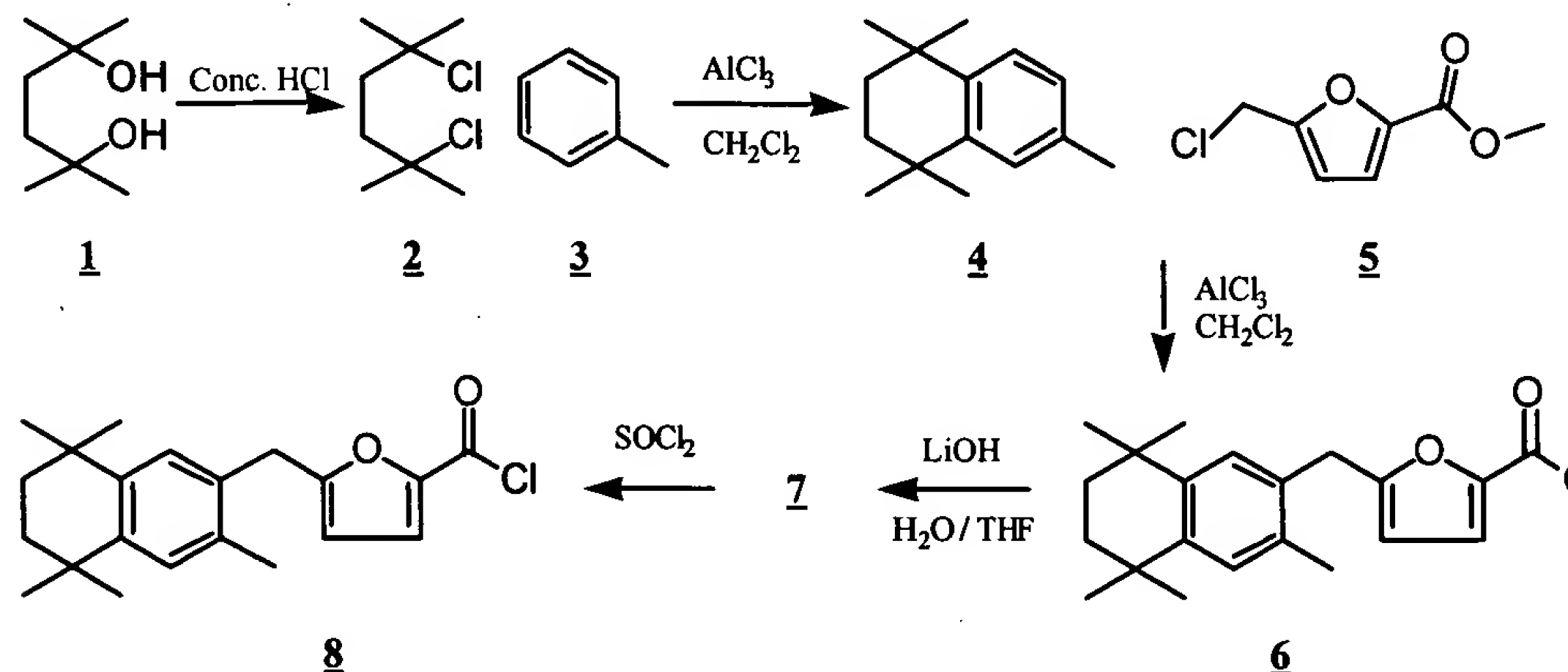
20 Parenteral Composition: To prepare a pharmaceutical composition of this invention suitable for administration by injection, 100 mg of a pharmaceutically acceptable water-soluble salt of a compound of Formula I is dissolved in DMSO and then mixed with 10 mL of 0.9% sterile saline. The resulting mixture is incorporated into a unit-dosage form suitable for administration by injection.

25 Oral Composition: To prepare an orally administerable pharmaceutical composition, 100 mg of a compound of Formula I is mixed with 750 mg of lactose. The resulting mixture is incorporated into a unit-dosage form suitable for oral administration, such as a hard-gelatin capsule.

SYNTHESIS OF GnRH REAGENTS AND COMPOUNDS

30 A. Building Block Example:

Naphthalene-Based Building Blocks: A useful acylating agent is prepared by sequential Friedel-Crafts alkylations and is shown below:



Compound 2 can be prepared as follows:

5 2,5-Dimethyl-2,5-hexanediol (200 grams, 1.37 mole) was added as a solid portion wise to 3 liters of concentrated hydrochloric acid in a large Erlenmeyer flask. The diol quickly dissolved in the hydrochloric acid and the desired product 2,5 dichloro-2,5 dimethylhexane precipitated out of solution as it was formed. The reaction was stirred at room temperature for 4 hours. One liter of 50% ethyl acetate in
10 hexanes were added and the organic layer separated and washed several times with water (until neutral by pH paper). The organic solvents were removed in vacuo at room temperature. The crude 2,5 dichloro-2,5 dimethylhexane was dissolved in hexanes and plugged through a pad of silica gel (10:1 ratio) and eluted with hexanes. This final filtration step gives a white solid after removal of the organic solvent in
15 vacuo. Recovery of pure 2,5 dichloro-2,5 dimethylhexane was 230 grams 92% yield. ¹H NMR (CDC13, delta): 1.96 (4H, s); 1.61 (12H, s).

Using a similar procedure 2,4-dimethyl-2,4-pentanediol was converted to 2,4-dichloro-2,4-deimthylpentane. ¹H NMR (CDC13, delta): 2.42 (2H, s); 1.73 (12H, s).

20 1,1,4,4,6-pentamethyl-1,2,3,4-tetrahydronaphthalene **4**: To a solution of 2,5 dichloro-2,5 dimethylhexane **2** (10 g, 54.7 mmol) in toluene (270 Ml, 0.2 M) is slowly added aluminum trichloride (5.47 g, 41 mmol) as a solid over a 15-minute period. The reaction is complete after 10 minutes as assayed by tlc in hexanes. The unreacted aluminum trichloride is quenched slowly with water over 10 minutes. Additional

toluene (250 mL) is added to extract the product from the aqueous layer. The organic layer is passed through a pad of silica gel (40 g) and eluted with toluene. The organic layer is evaporated *in vacuo* to dryness to yield 1,1,4,4,6-pentamethyl-1,2,3,4-tetrahydronaphthalene **4** (11g, 97% yield). NMR 1.29 (s, 6H), 1.28 (s, 6H), 1.69 (s, 4H), 2.32 (s, 3H), 7.22 (d, 1H), 7.12 (s, 1H), 6.97 (dd, 1H).

Methyl 5-[(3,5,5,8,8-pentamethyl-5,6,7,8-tetrahydro-2-naphthalenyl)methyl]-2-furoate **6**: To a solution containing 1,1,4,4,6-pentamethyl-1,2,3,4-tetrahydronaphthalene **4** (20 g, 99 mmol) and methyl 5-(chloromethyl)-2-furoate **5** (17.28 g, 99 mmol) in methylene chloride (500 mL, 0.2 M), aluminum trichloride (16.46 g, 124 mmol) is added slowly as a solid at the reflux temperature of methylene chloride. The solution is refluxed for an additional two hours. The reaction is monitored by tlc in 10% ethyl acetate/hexanes solution. The reaction is cooled to room temperature and the unreacted aluminum trichloride is quenched with water over 15 minutes. The crude product is extracted with methylene chloride and passed through silica gel (80g) and eluted with methylene chloride. The solvent is evaporated *in vacuo* to syrup. The crude product is purified with silica gel (300 g) via a plug filtration column. Methyl 5-[(3,5,5,8,8-pentamethyl-5,6,7,8-tetrahydro-2-naphthalenyl)methyl]-2-furoate **6** is eluted with 2 % ethyl acetate/hexanes to afford 15.4 g (46 % yield). NMR 1.25 (s, 6H), 1.28 (s, 6H), 1.67 (s, 4H), 2.23 (s, 3H), 3.89 (s, 3H), 3.97 (s, 2H), 5.95 (d, 1H), 7.09 (m, 3H).

5-[(3,5,5,8,8-pentamethyl-5,6,7,8-tetrahydro-2-naphthalenyl)methyl]-2-furoic acid **7**: To a solution containing methyl 5-[(3,5,5,8,8-pentamethyl-5,6,7,8-tetrahydro-2-naphthalenyl)methyl]-2-furoate **6** (15.1 g, 44 mmol) in MeOH (175 mL) and water (175 mL), a solution of NaOH (3.53g, 88.3 mmol) in water (29mL) is added. The reaction mixture is stirred overnight. After completion as judged by tlc, the solution is acidified with 1M HCl to pH 2. The crude product is extracted into organic layer using ethyl acetate, and concentrated to afford 5-[(3,5,5,8,8-pentamethyl-5,6,7,8-tetrahydro-2-naphthalenyl)methyl]-2-furoic acid **7** (15.0 g, 99 % yield). NMR 1.26 (s, 6H), 1.28 (s, 6H), 1.68 (s, 4H), 2.24 (s, 3H), 4.00 (s, 2H), 6.01 (d, 1H), 7.10 (s, 2H), 7.23 (d, 1H).

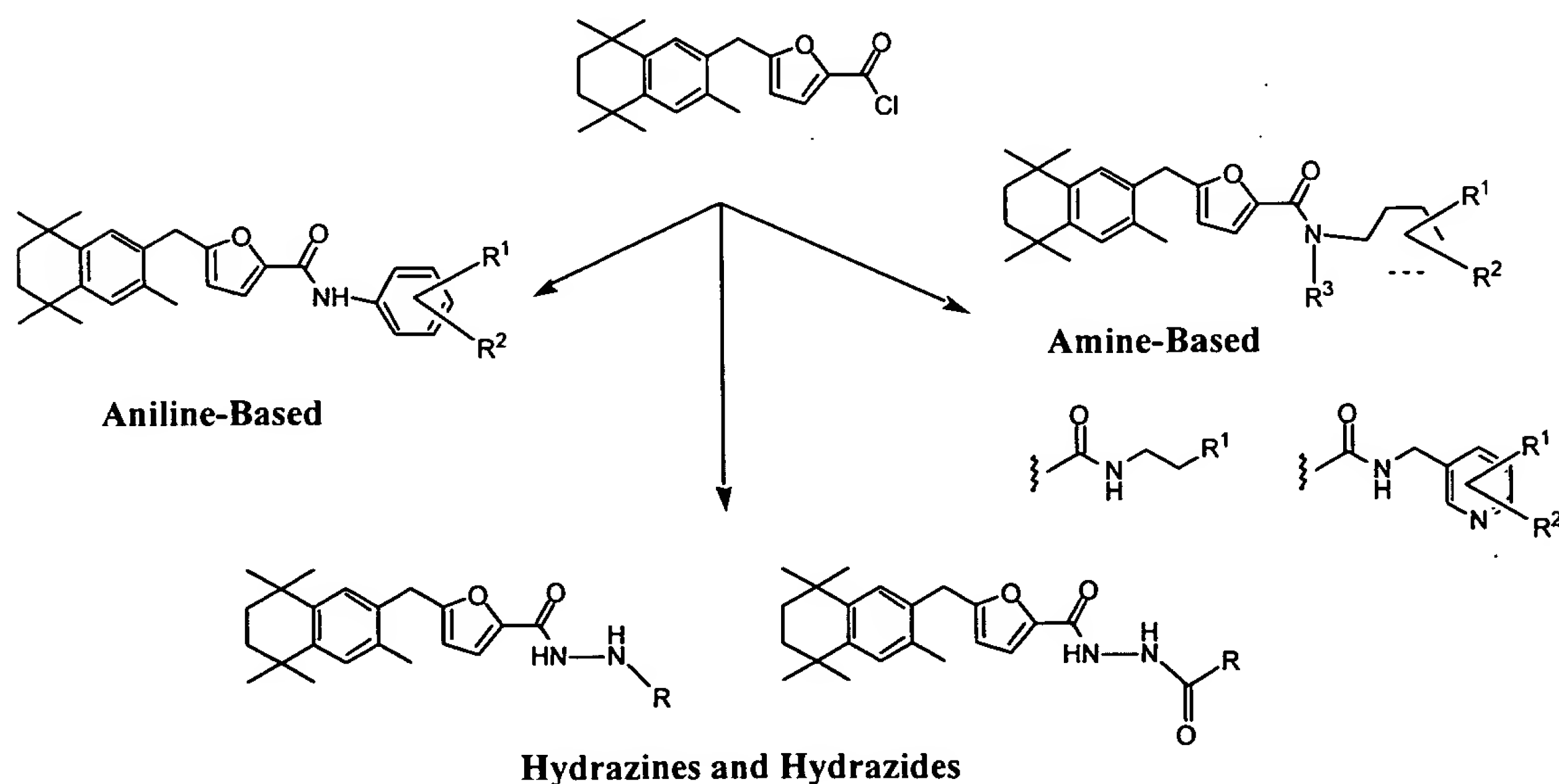
5-[(3,5,5,8,8-pentamethyl-5,6,7,8-tetrahydro-2-naphthalenyl)methyl]-2-furoyl chloride **8**: To a solution containing 5-[(3,5,5,8,8-pentamethyl-5,6,7,8-tetrahydro-2-naphthalenyl)methyl]-2-furoic acid **7** (20.15 g, 61.77 mmol) in methylene chloride (310 mL), thionyl chloride (45 mL, 617 mmol) is added. The reaction is refluxed for 5 hours and another batch of thionyl chloride (45 mL, 617 mmol) is added. The reaction is stirred overnight at room temperature. The solution is concentrated to a syrup and passed through a pad of silica gel (50 g), washed with 3% hexanes, and concentrated *in vacuo* to afford 5-[(3,5,5,8,8-pentamethyl-5,6,7,8-tetrahydro-2-naphthalenyl)methyl]-2-furoyl chloride **8** (17 g, 80% yield). NMR 1.26 (s, 6H), 1.28 (s, 6H), 1.68 (s, 4H), 2.25 (s, 3H), 4.00 (s, 2H), 6.11 (d, 1H), 7.10 (s, 1H), 7.11 (s, 1H), 7.41 (d, 1H).

Additional building blocks can be prepared under these reaction conditions which contain a variety of functional groups contained in the general formula shown above.

15 B. Acylation Examples:

The next scheme shows several examples which can use the general synthetic procedure for acylations given below.

Reagent Examples Coupling to Y



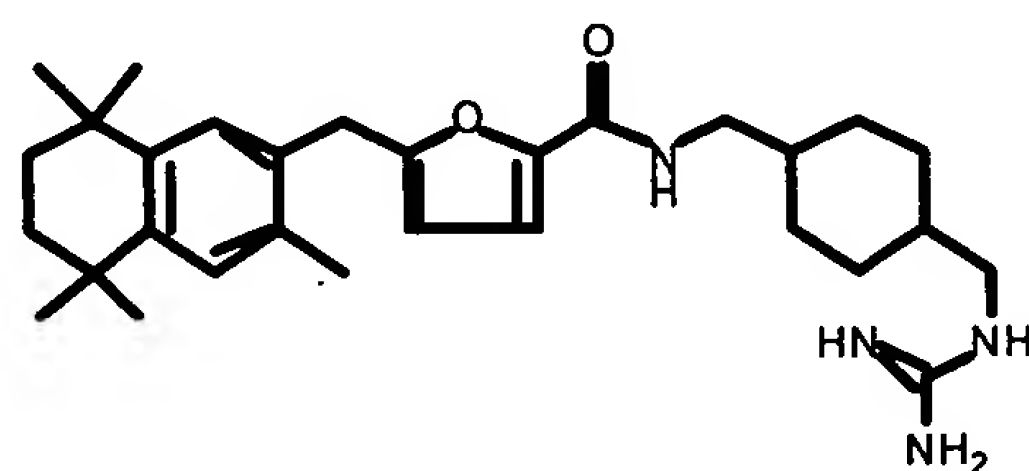
Amines are dissolved or suspended in dichloromethane, dichloroethane, ethyl acetate, acetonitrile, or the like (0.2M concentration) followed by the addition of the acid chloride reagent (1.00 mmol. equiv.). To the mixture is added triethyl amine (5.00 mmol. equiv.) and the reaction stirred at room temperature for 12-48 hours. The solvents are removed *in vacuo*. The product is purified by column chromatography on silica gel and eluted with an appropriate elution solvent (e.g., 3:1 hexanes:ethyl acetate). The solvents are removed *in vacuo* to yield the acylated product.

As an alternative, the reaction mixture is diluted with dichloromethane (five times the amount of dichloromethane used) and washed with saturated sodium bicarbonate. The organic layer is dried over magnesium sulfate and filtered. The product is purified by column chromatography on silica gel and eluted with an appropriate elution solvent (e.g., 3:1 hexanes:ethyl acetate). The solvents are removed *in vacuo* to yield the acylated product.

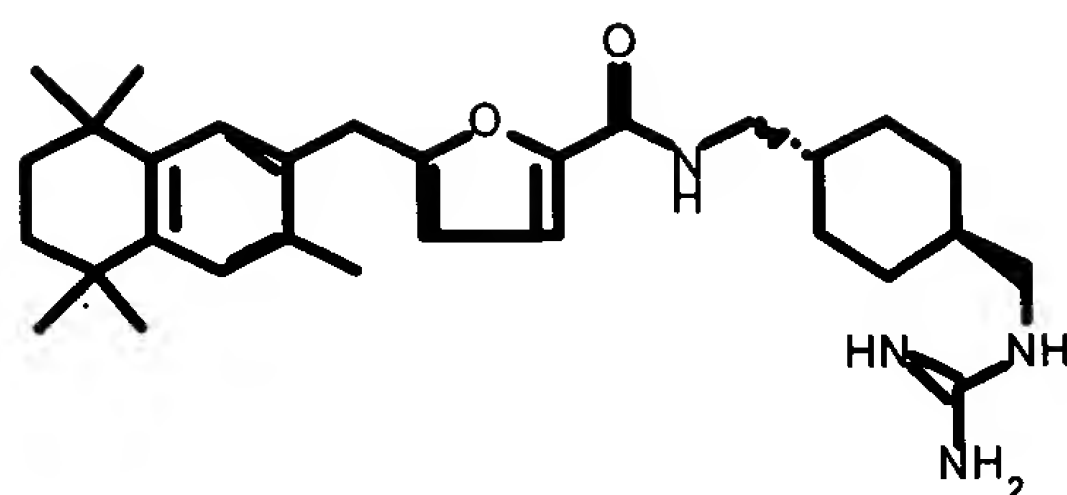
Using the general reaction protocol, large numbers of compounds can be readily prepared and assayed for their activities either as pure or impure materials. The reaction protocol works well on anilines, amines, benzyl amines, hydrazines, hydrazides, alcohols and the like.

Specific examples showing a variety of structures acylated according to a general procedure are shown below:

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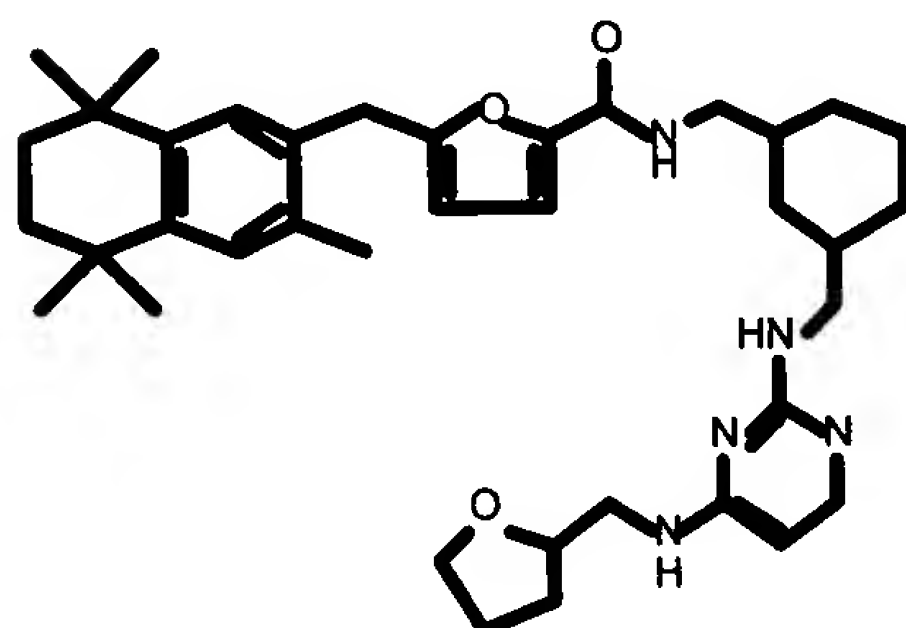
COMPOUND NO.	STRUCTURE	mol. weight
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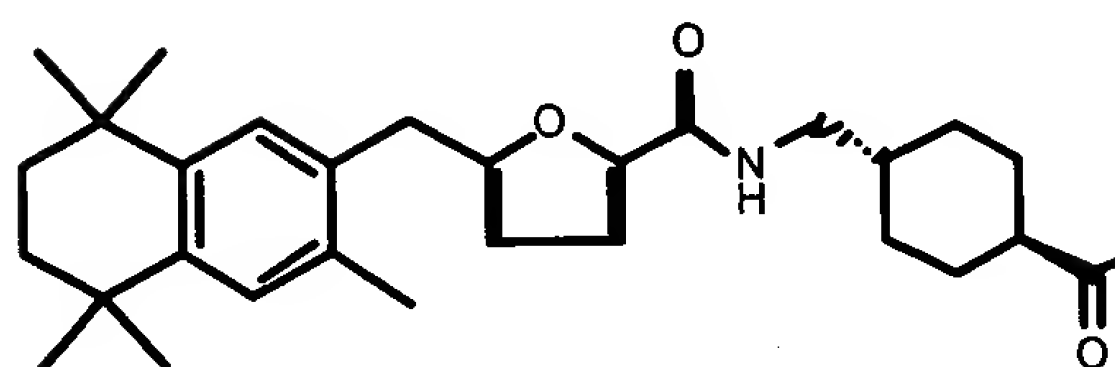
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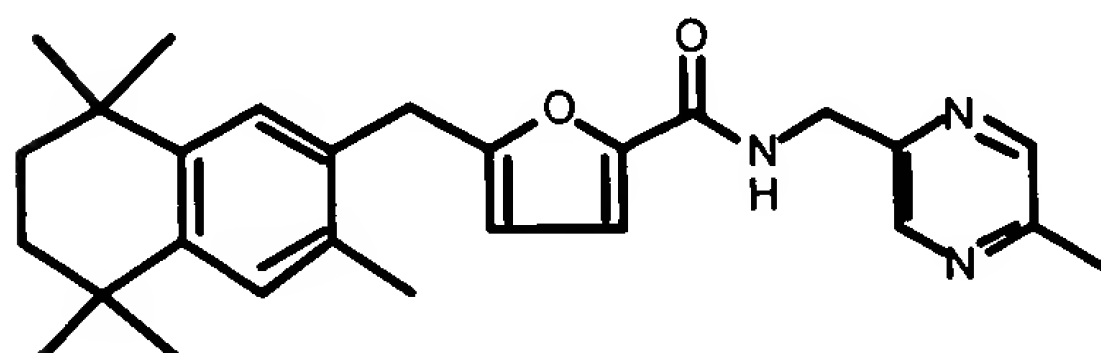
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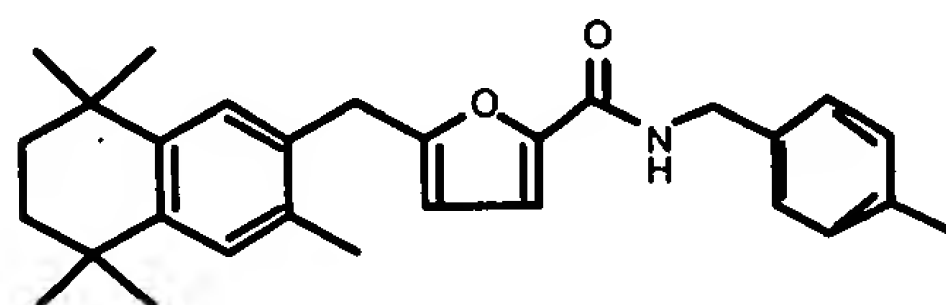
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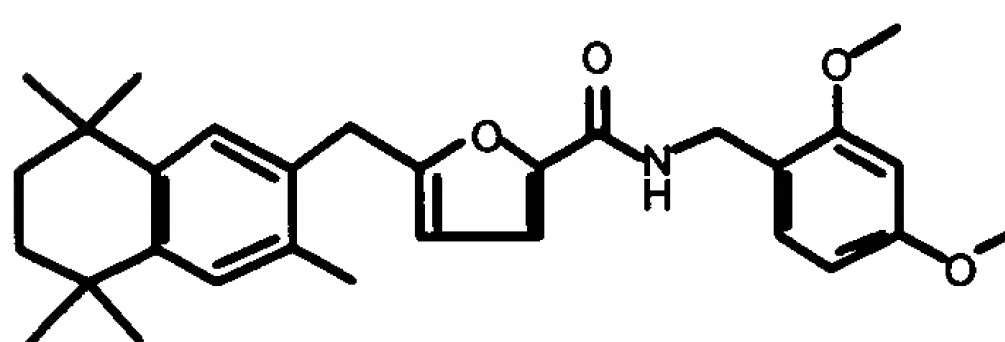
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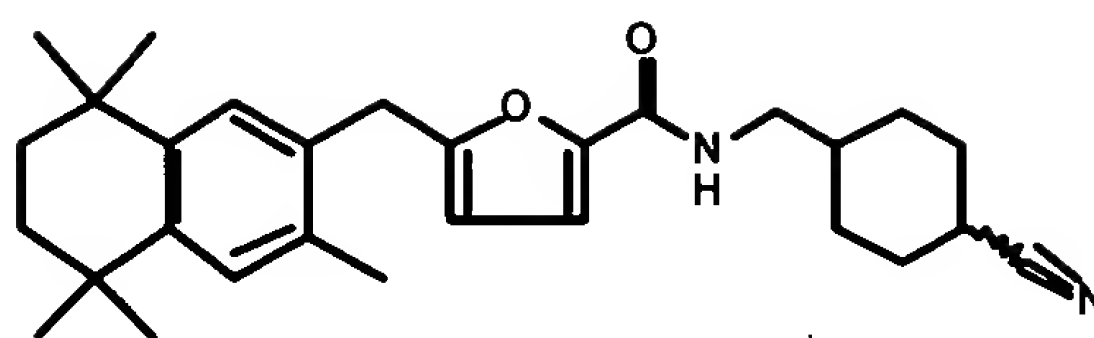
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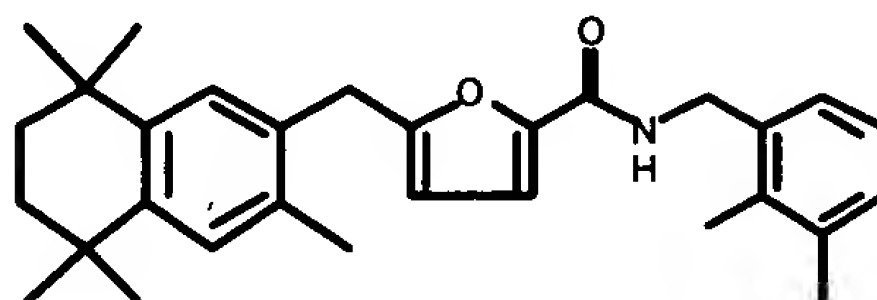
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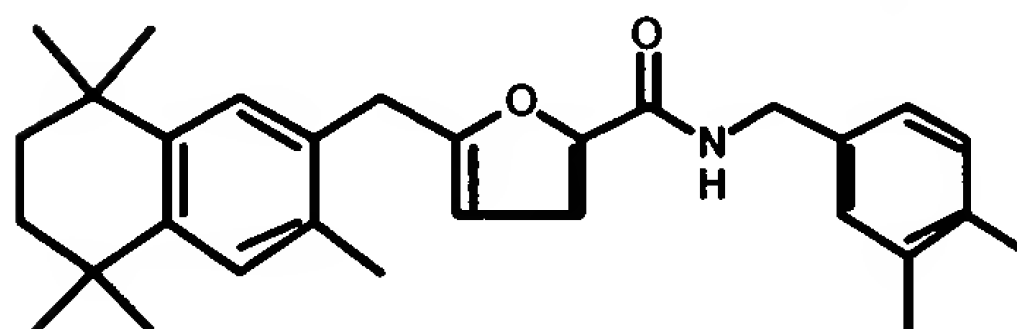
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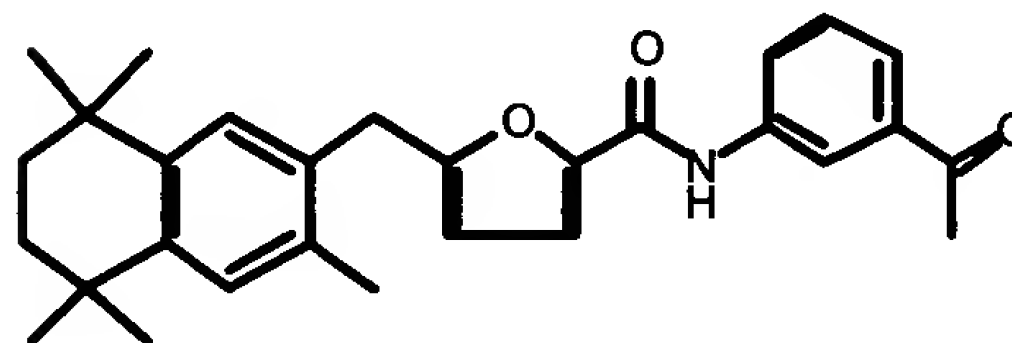
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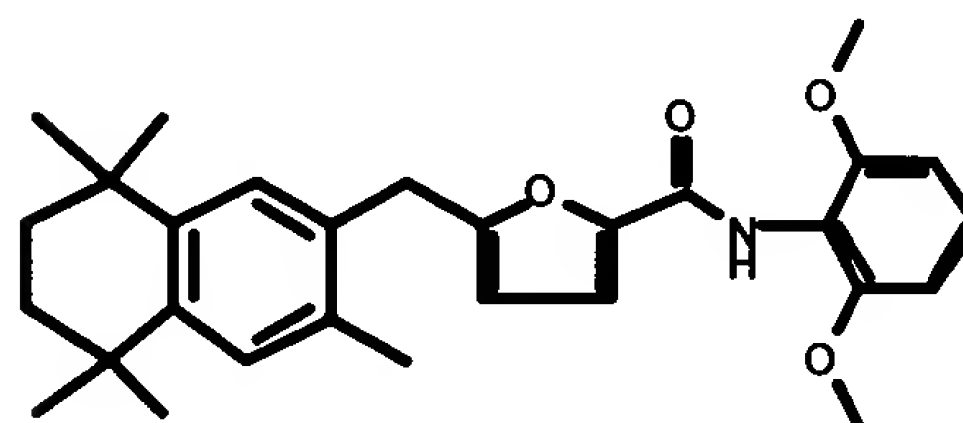
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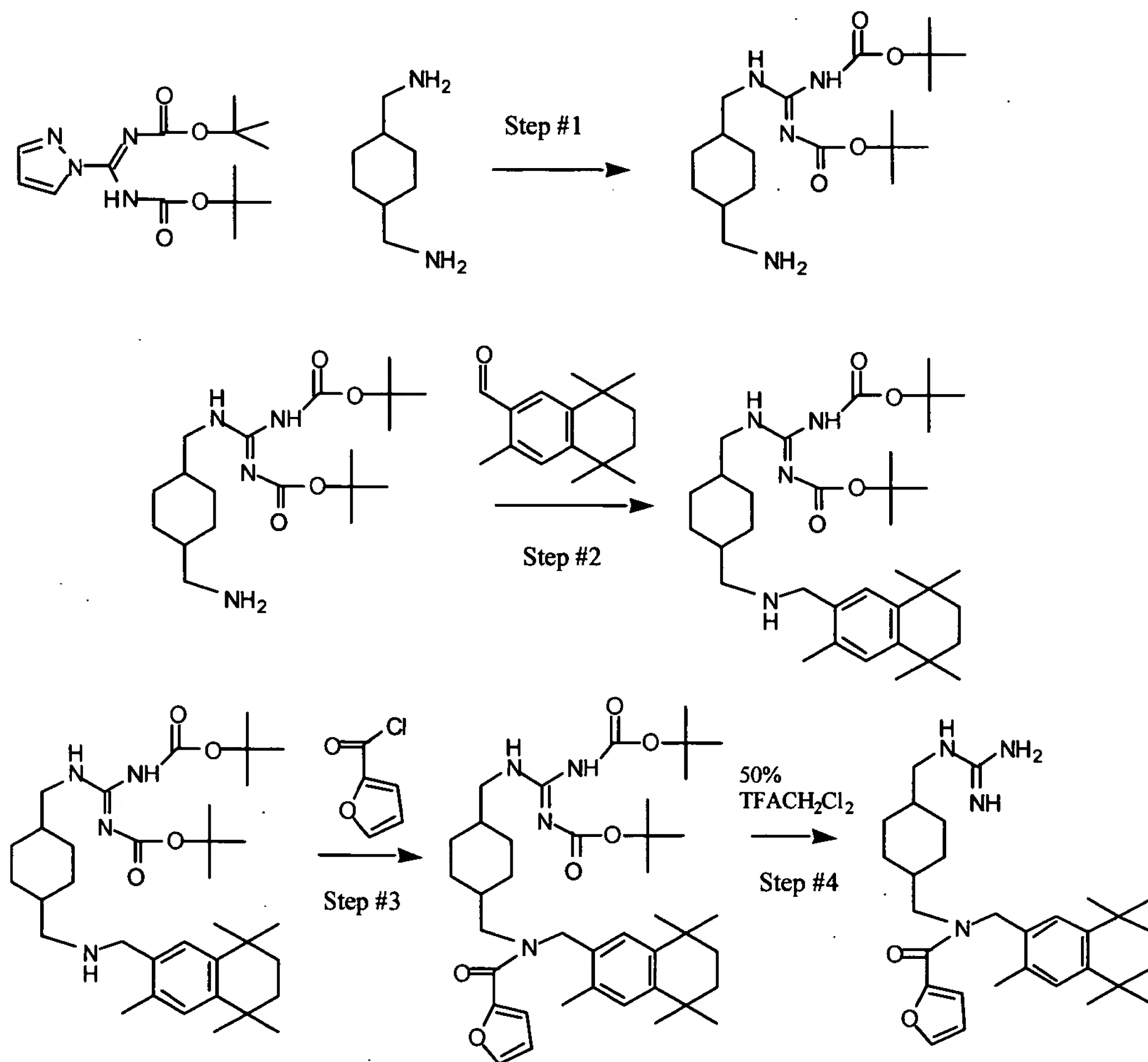
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C. Synthesis and Acylation of Guanidine-Containing Compounds:



Step 1 -- Preparation of Protected Compound by 1-(N,N'-diBoc)-

- 5 guanidinomethylation: Alternative Steps 1(A) and 1(B) below provide two general 1-(N,N'-diBoc)-guanidinomethylation procedures.

10 Step 1(A): To a solution of diamine (2.00 mmol equiv.) in THF (0.7 M) is added a solution of 1-H-pyrazole-1-(N,N-bis(tert-butoxycarbonyl)carboxamidine) (1.00 mmol equiv.) in THF (0.7M). The solution is stirred at room temperature for 3 hours (h), or until no further transformation can be observed by tlc (thin-layer chromatography). The solvent is removed under reduced pressure to give a syrupy residue, which is taken up in ethyl acetate (~1.5 times the volume amount of THF used in the reaction or the volume of solvent needed to dissolve the amount of residue

obtained) and washed with water until neutral pH. The organic layer is washed with brine, dried over MgSO₄, and concentrated. The product is purified by column chromatography on silica gel and eluted with an appropriate elution solvent (which may be readily determined, e.g., using 5% MeOH in dichloromethane as a starting point). The solvents are removed *in vacuo* to afford the 1-(N,N'-diBoc)-
5 guanidinomethyl-linked-amine. In addition other reagents can be used to place a protected N,N'-diBoc-guanidine unit on diamines, such as 1,3-bis(tert-butoxycarbonyl)-2-methyl-2-thiopseudourea (CAS No. 107819-90-0). Alternatively, the 1-H-pyrazole-1-(N,N-bis(tert-butoxycarbonyl)carboxamidine) can be added
10 directly as a solid, rather than as a solution as described above.

Step 1(B): To a solution of diamine (1.00 mmol equiv.) in THF (0.07M) is added portionwise as a solid (over a 10-minute time period) 1-H-pyrazole-1-(N,N-bis(tert-butoxy-carbonyl)carboxamidine) (1.00 mmol equiv.). The solution is stirred at room temperature for 0.5 hour. The solvent is removed under reduced pressure to give
15 a syrupy residue, which is taken up in ethyl acetate (0.5 times the volume amount of THF used in the reaction, or the volume of solvent needed to dissolve the amount of residue obtained) and washed twice with water. The layers are separated, and the product is purified by column chromatography on silica gel and eluted with 100% ethyl acetate to remove any non-polar impurities and then with 100% isopropyl alcohol to
20 give the pure product. The solvents are removed *in vacuo* to afford the desired product. Typical TLC conditions are 15:85:0.1 methanol/chloroform/acetic acid. Typical yields range from 40% to 44% of the desired protected compound.

Step 2 -- Reductive Amination (optional): Reductive amination may be accomplished in a suitable manner. For reductive amination of aldehydes and ketones
25 with sodium triacetoxymethylborohydride, see generally: Abdel-Magid et al., *J. Org. Chem.*, 1996, 61:3849. Two alternate reductive-aminations procedures are described below.

Step 2(A): 3,5,5,8,8-Pentamethyl-5,6,7,8-tetrahydro-2-naphth-aldehyde (1.00 mmol equiv.) and 1-(N,N'-diBoc)-guanidinomethyl-linked-amine (1.00 mmol equiv.) are dissolved in methanol (0.09M). Then, 1% glacial acetic acid in methanol solution
30 (10% of the volume of methanol used) is added followed by NaCNBH₃ (1.00 mmol

equiv.), and the reaction contents are stirred overnight. The reaction is assayed by TLC to reveal three components (aldehyde, desired product, and starting guanidine derivative). The reaction is terminated by adding water (50% of the volume of methanol used), extracted with dichloromethane (10 times the volume of methanol used), and washed with saturated sodium bicarbonate. The organic layer is dried over magnesium sulfate, filtered, and concentrated. The product is purified by column chromatography on silica gel and eluted with an appropriate elution solvent (e.g., 3:1 ethyl acetate in hexanes to remove the unreacted aldehyde, followed by elution with 1:1 ethyl acetate in hexanes), obtaining the desired reductive amination product. In some cases, warming to reflux for 2 hours will facilitate the imine formation reaction. See also, Abdel-Magid et al., *J. Org. Chem.*, 1996, 61:3849, which describes the reductive amination of aldehydes and ketones with sodium triacetoxyborohydride.

Step 2(B): 3,5,5,8,8-Pentamethyl-5,6,7,8-tetrahydro-2-naphth-aldehyde (1.00 mmol equiv.) and 1-(N,N'-diBoc)-guanidinomethyl-linked-amine (1.00 mmol equiv.) are dissolved in methanol (0.09M). Then, NaBH₄ (1.00 mmol equiv.) is added (in ethanol via the additional small-scale procedures given below, or carefully as a solid) and the reaction contents are stirred overnight. The reaction is assayed by TLC to reveal three components (aldehyde, desired product and starting guanidine derivative). The reaction is terminated by the addition of water (50% of the volume of methanol used), extracted with dichloromethane (10 times the volume of methanol used), and washed with saturated sodium bicarbonate. The organic layer is dried over magnesium sulfate, filtered, and concentrated. The product is purified by column chromatography on silica gel and eluted with an appropriate elution solvent (as can be readily determined by the skilled artisan or, for example, with 3:1 ethyl acetate in hexanes to remove the unreacted aldehyde followed by elution with 1:1 ethyl acetate in hexanes) to obtain the desired reductive-amination product. In some cases, warming to reflux for 2 hours should facilitate the imine-formation reaction.

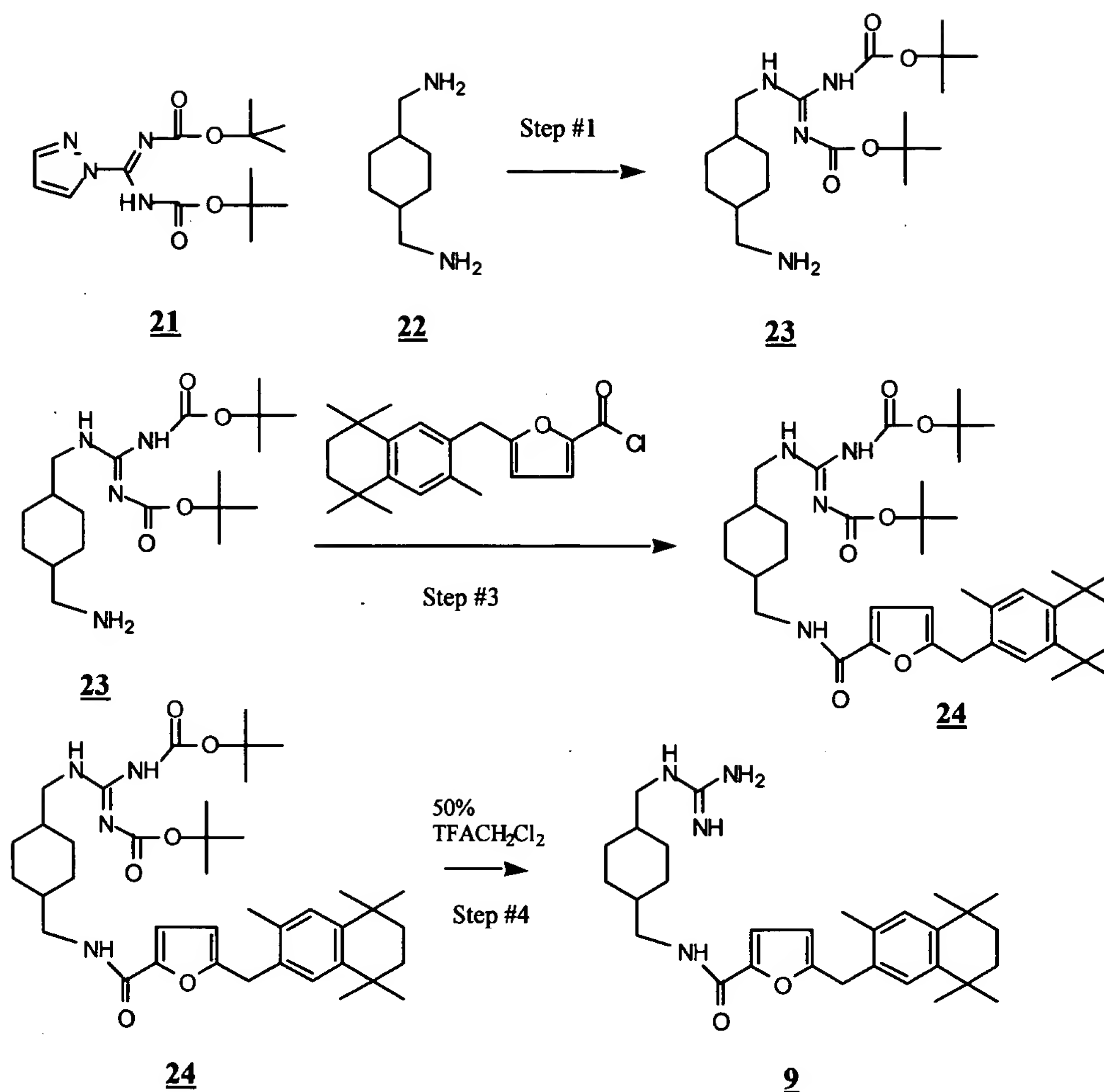
Step 3 -- Acylation: The products from the reductive amination (1.00 mmol equiv.) are dissolved in dichloromethane (~0.2 to 0.05M, depending on solubilities of the substrates), followed by the addition of triethylamine (2.00 mmol equiv.) and 2-

furoyl chloride reagent **8** (1.00 mmol equiv.). The reaction contents are stirred overnight at room temperature (RT). The reaction mixture is diluted with dichloromethane (5 times the amount of dichloromethane used) and washed with saturated sodium bicarbonate. The organic layer is dried over magnesium sulfate and
5 filtered. The product is purified by column chromatography on silica gel and eluted with an appropriate elution solvent (e.g., 3:1 hexanes:ethyl acetate). The solvents are removed *in vacuo* to yield the acylated product.

Step 4 -- Basic Group Deprotection: The product from the acylation step (1.00 mmol equiv.) is dissolved in a solution of 25-50% TFA in dichloromethane (0.02M),
10 and the reaction contents are stirred at room temperature (15-20 minutes; solution becomes slight reddish-orange). The reaction contents are stirred for an additional 1 hour and 20 minutes or until the BOC deprotection is complete. The reaction is terminated by concentration *in vacuo*, followed by the addition of water/acetonitrile (0.006M) and lyophilization overnight. The final compound is purified by high-
15 performance liquid chromatography (HPLC) methodology. The solvents are removed *in vacuo* (yields range from 30% to 50%) to give the product.

An alternate procedure for removing of N, N'-bis-BOC guanidines using tin tetrachloride, which can give the corresponding guanidinium chloride salts, is described in Miel et al., *Tetrahedron Letters*, 1997, 38:7865-7866.

20 Compound **9** may be prepared according to the steps shown above with the exclusion of step #2, as shown in the following scheme:



Preparation of Reagents: Reagents useful for synthesizing compounds may be obtained or prepared according to techniques known in the art. For example, the preparation of free amines from common salt forms and stock reagent solutions can be useful for small-scale reactions. See also Abdel-Magid et al., "Reductive amination of

5 aldehydes and ketones with sodium triacetoxyborohydride," *J. Org. Chem.*, 1996, 61:3849.

Methanolic solutions of the free bases can be prepared from hydrochloride, dihydrochloride, hydrobromide, or other salts when the free base is soluble in

10 methanol. In this procedure, once the sodium methoxide is added, care should be taken to prevent exposure to air, since amine free bases, particularly primary amines, absorb carbon dioxide from the air to form salts. A 10-mL quantity of a 0.1M solution

of a free base in methanol may be prepared as follows. Weigh 1.0 mmol of a monohydrochloride salt into a tared Erlenmeyer flask containing a stirring bar, and add 7 mL of methanol. To the stirred slurry, add 229 mL (1.0 mmol, 1 equiv.) of sodium methoxide in methanol (25 wt %, 4.37M), stopper the flask, and stir the mixture vigorously for 2 hours. The slurry will sometimes change in appearance as a finer, milky precipitate of sodium chloride is formed. Filter the slurry through a 15-mL medium fritted glass funnel, wash the filter case with 1-2 mL methanol, transfer the filtrate to a 20-mL vial, and dilute to 10 mL with methanol. The theoretical yield of sodium chloride is nearly 59 mg, but the recovery is usually not quantitative, owing to a slight solubility in methanol. For a dihydrochloride salt, a second equivalent of sodium methoxide is required (458 mL).

A 0.5M solution of sodium borohydride in ethanol may be prepared as follows. Sodium borohydride (520 mg, 13.8 mmol) is stirred in pure (non-denatured) anhydrous ethanol (25 mL) for ~2-3 minutes. The suspension is filtered through a medium fritted glass funnel to remove a small amount of undissolved solid (typically about 5% of the total mass of borohydride, or 25 mg). The filtrate should appear as a colorless solution that evolves only a little hydrogen. This solution should be used immediately, as it decomposes significantly over a period of a few hours, resulting in the formation of a gelatinous precipitate. Sodium borohydride is hygroscopic, so avoid exposure to air by making the solution at once after weighing the solid. Sodium borohydride has a solubility of about 4% in ethanol at room temperature. This corresponds to a little over 0.8M. However, sometimes a small percentage of the solid remains undissolved regardless of the concentration being prepared, even after stirring for ≥ 5 minutes.

To perform small-scale synthesis of compounds of the Formula I, the reactions described below may be performed to prepare various reactants useful in the reaction scheme described above. As with the rest of the specification, all temperatures in the following description are in degrees Celsius and all parts and percentages are by weight, unless indicated otherwise.

Various starting materials and other reagents may be purchased from commercial suppliers, such as Aldrich Chemical Company or Lancaster Synthesis Ltd.,

and used without further purification, unless otherwise indicated. Tetrahydrofuran (THF) and N,N-dimethylformamide (DMF) are purchased from Aldrich in SureSeal[®] bottles and used as received. All solvents are purified by using standard methods in the art, unless otherwise indicated.

5 The reactions set forth below are performed under a positive pressure of nitrogen or with a drying tube, at ambient temperature (unless otherwise stated), in anhydrous solvents, and the reaction flasks are fitted with rubber septa for the introduction of substrates and reagents via syringe. Glassware is oven-dried and/or heat-dried. Analytical thin-layer chromatography is performed on glass-backed silica
10 gel 60°F 254 plates (Analtech (0.25 mm)) and eluted with the appropriate solvent ratios (v/v). The reactions are assayed by TLC and terminated as judged by the consumption of starting material.

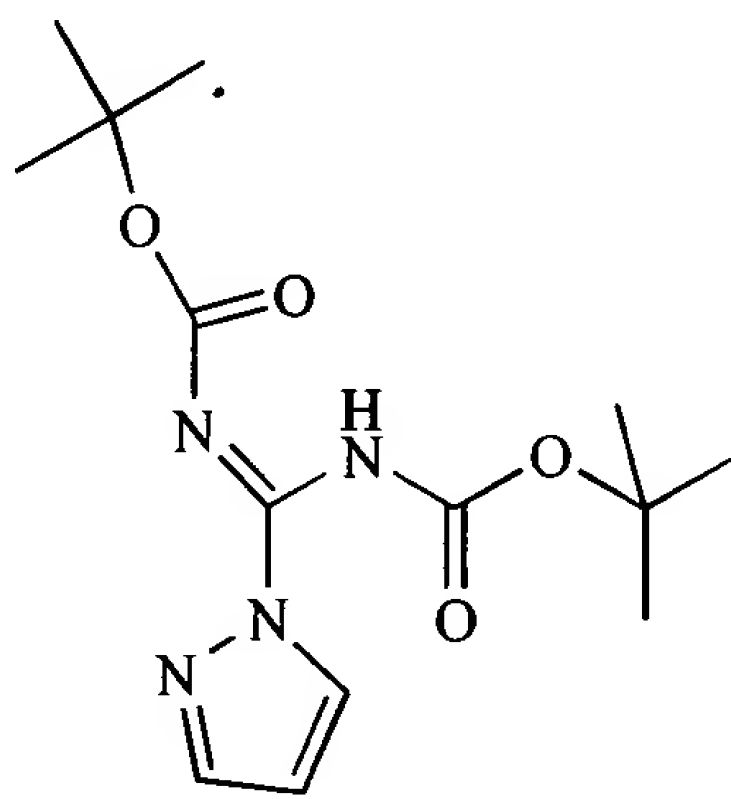
 The tip plates are visualized with a *p*-anisaldehyde spray reagent or phosphomolybdic acid reagent (Aldrich Chemical, 20 wt% in ethanol) and activated
15 with heat. Work-ups are typically done by doubling the reaction volume with the reaction solvent or extraction solvent and then washing with the indicated aqueous solutions using 25% by volume of the extraction volume (unless otherwise indicated). Product solutions are dried over anhydrous Na₂SO₄ prior to filtration, and evaporation of the solvents is under reduced pressure on a rotary evaporator and noted as solvents
20 removed *in vacuo*. Flash column chromatography (Still et al., *A.J. Org. Chem.*, 1978, 43:2923) is conducted using Baker-grade flash silica gel (47-61mm) and a silica gel:crude material ratio of about 20:1 to 50:1, unless otherwise stated. Hydrogenolysis is done at the pressure indicated or at ambient pressure.

¹H-NMR spectra are recorded on a Bruker instrument operating at 300 MHz,
25 and ¹³C-NMR spectra are recorded operating at 75 MHz. NMR spectra are obtained as CDCl₃ solutions (reported in ppm), using chloroform as the reference standard (7.25 ppm and 77.00 ppm) or CD₃OD (3.4 and 4.8 ppm and 49.3 ppm), or an internal tetramethylsilane standard (0.00 ppm) when appropriate. Other NMR solvents are used as needed. When peak multiplicities are reported, the following abbreviations are
30 used: s = singlet, d = doublet, t = triplet, m = multiplet, br = broadened, dd = doublet

of doublets, dt = doublet of triplets. Coupling constants, when given, are reported in Hertz.

Infrared spectra are recorded on a Perkin-Elmer FT-IR Spectrometer as neat oils, as KBr pellets, or as CDCl₃ solutions, and when reported are in wave numbers (cm⁻¹). The mass spectra are obtained using LSIMS or electrospray. All melting points are uncorrected.

Preparation of the Building Block 1-H-pyrazole-1-carboxamidine:

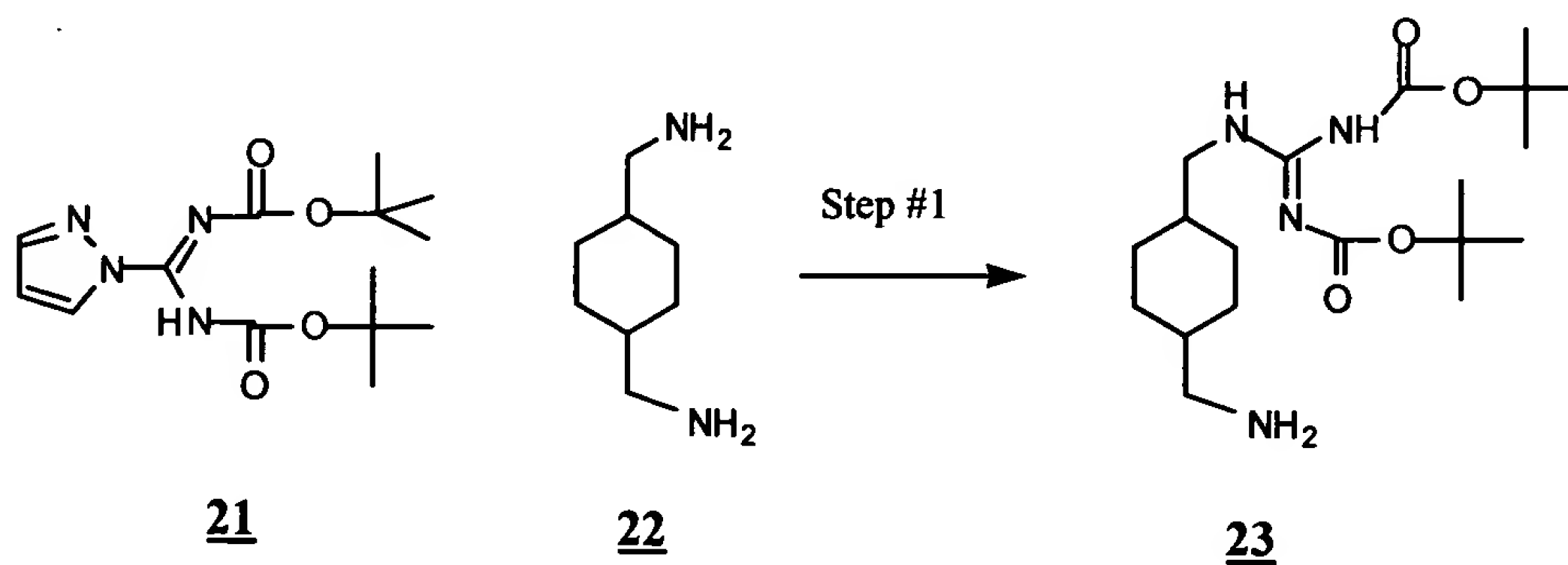


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1-H-pyrazole-1-carboxamidine is prepared according to Bernatowicz et al., *J. Org. Chem.*, 1992, 57:2497-2502 (and references therein), and protected with di-tert-butyl dicarbonate to give 1-H-pyrazole-1-(N,N-bis(tert-butoxycarbonyl)carboxamidine) according to Drake et al., *Synth.*, 1994, 579-582.

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Preparation of 1-(N,N'-diBoc)-guanidinomethyl-4-aminomethylcyclohexane:

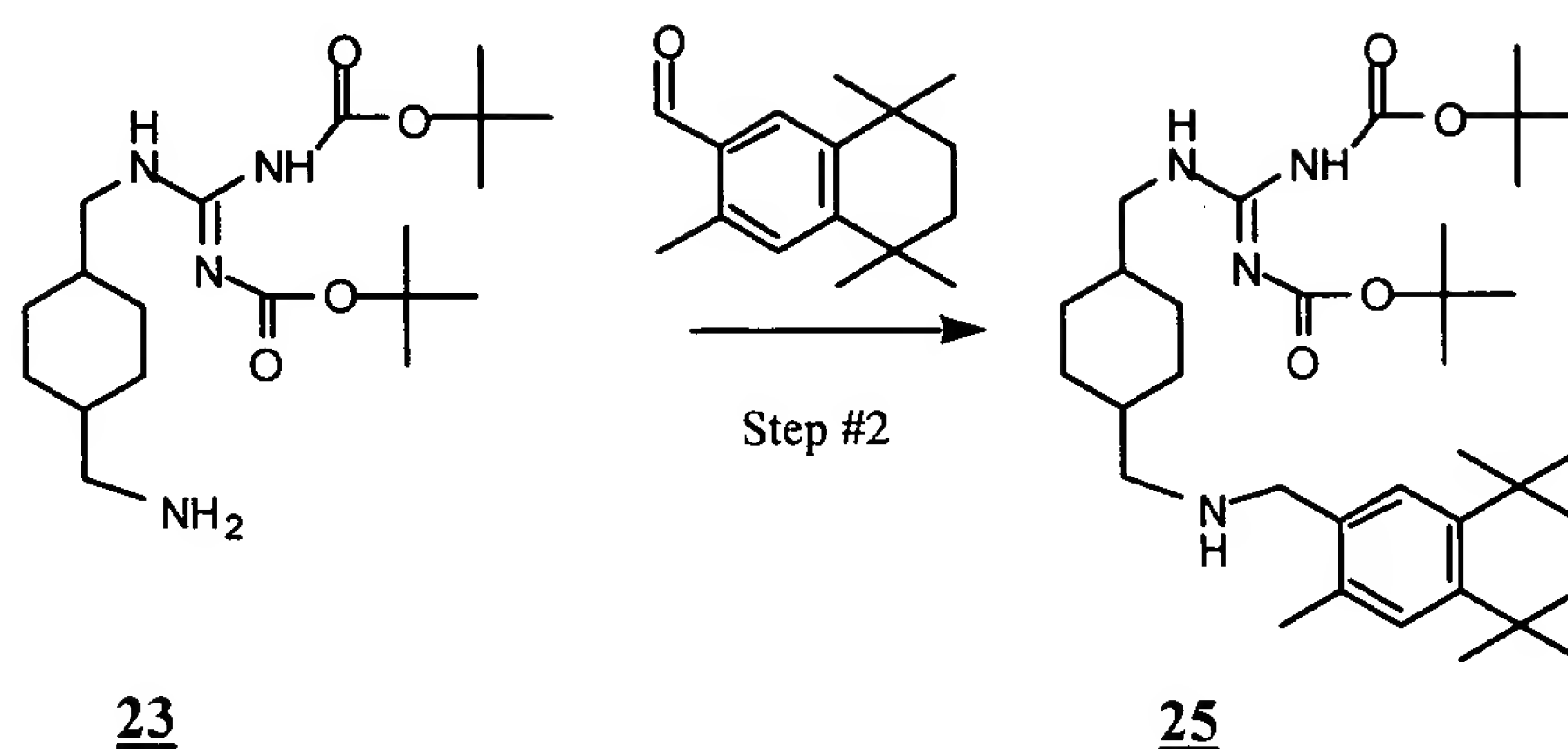


To a solution of 1,4-bis-aminomethyl-cyclohexane 22 (20 g, 0.14 mol) in THF (200 mL) is added a solution of 1-H-pyrazole-1-(N,N-bis(tert-

butoxycarbonyl)carboxamidine) **21** (22.0 g, 0.07 mol) in THF (100 mL). (Note that 1-H-pyrazole-1-(N,N-bis(tert-butoxycarbonyl) carboxamidine) does not need to be dissolved in THF; rather it may be added neat as a solid to the process.) The solution is stirred at room temperature for 3 hours. The solvent is removed under reduced pressure to give a syrupy residue, which is taken up in ethyl acetate (500 mL) and washed with water until neutral pH. The organic layer is washed with brine, dried over MgSO₄, and concentrated. The product is purified by column chromatography on silica gel and eluted with 5% MeOH in dichloromethane. The solvents are removed *in vacuo* to afford 11.6 g (43 % yield) of 1-(N,N'-diBoc)-guanidinomethyl-4-aminomethyl cyclohexane (Compound **23**). ¹H NMR (CDCl₃) δ 11.5 (br s, 1H), 8.35 (br s, 1H), 3.26 (dt, 2H), 2.52 (dd, 2H), 1.82-0.97 (m, 28H, with singlet at 1.5).

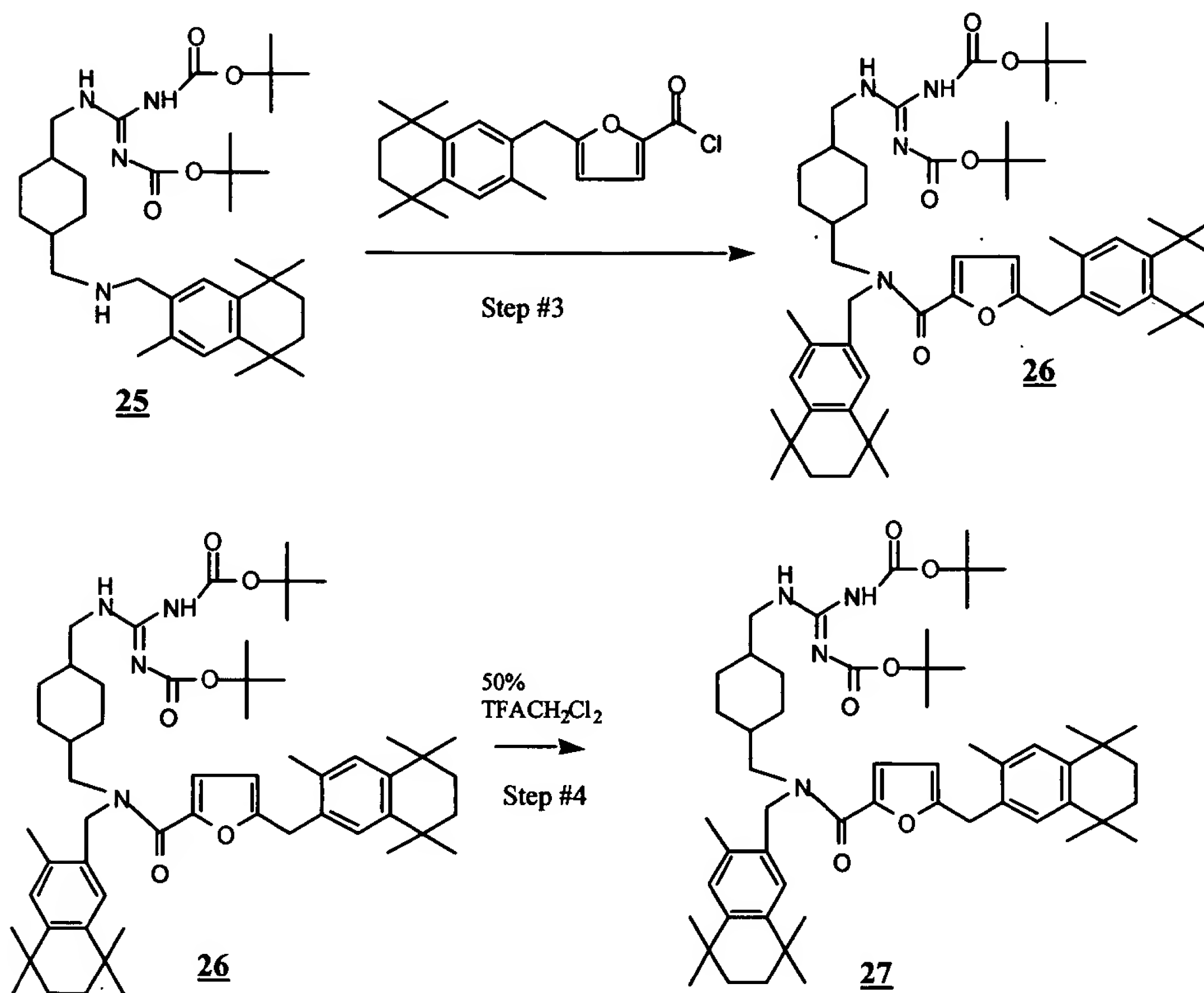
An alternate preparation of 1-(N,N'-diBoc)-guanidinomethyl-3-aminomethylcyclohexane is as follows. To a solution of *cis/trans* 1,4-bis-aminomethyl-cyclohexane (9.0 g, 63.3 mmol) in THF (903 mL, 0.07M) is added portionwise as a solid (over a 10-minute period) 1-H-Pyrazole-1-(N,N-bis(tert-butoxycarbonyl)carboxamidine) (19.6 g, 63.3 mmol). The solution is stirred at room temperature for 0.5 hour. The solvent is removed under reduced pressure to give a syrupy residue, which is taken up in ethyl acetate (500 mL) and washed twice with water. The layers are separated and the product is purified by column chromatography on silica gel and eluted with 100% ethyl acetate to remove any non-polar impurities, followed by elution with 100% isopropyl alcohol, to give the pure product. The solvents are *removed in vacuo* to afford 10.2 g (42 % yield) of 1-(N,N'-diBoc)-guanidinomethyl-4-aminomethylcyclohexane. ¹H NMR (CDCl₃) δ 11.5 (br s, 1H), 8.35 (br s, 1H), 3.26 (dt, 2H), 2.52 (dd, 2H), 1.82-0.97 (m, 28H, with singlet at 1.5).

Reductive Amination:



3,5,5,8,8-Pentamethyl-5,6,7,8-tetrahydro-2-naphth-aldehyde (0.2021 g, 0.88
5 mmol) and 1-(N,N'-diBoc)-guanidinomethyl-4-aminomethylcyclohexane (Compound
23, 0.337 g, 0.88 mmol) are dissolved in methanol (10 mL). Then, 1% glacial acetic
acid in methanol (100 μ L) solution is added followed by NaCNBH₃ (55.4 mg, 0.88
mmol, 1.0 equiv.), and the reaction contents are stirred overnight. The reaction is
assayed by TLC to reveal three components (aldehyde, desired product, and starting
10 guanidine derivative). The reaction is terminated by the addition of water (~5 mL),
extracted with dichloromethane (~100 mL), and washed with saturated sodium
bicarbonate. The organic layer is dried over magnesium sulfate, filtered, concentrated,
and subjected to column chromatography eluting with 3:1 ethyl acetate in hexanes to
remove the unreacted aldehyde, followed by eluting with 1:1 ethyl acetate in hexanes,
15 yielding the desired product (Compound **25**, cyclohexyl, cis/trans mixture). The
solvents are removed *in vacuo* (typical general yields range from 50 to 80 %).

Preparation of the Acylated Derivative Followed by Deprotection of Guanidine:



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The product from the reductive amination **25** (1.0 equiv.) is dissolved in dichloromethane (10-15 mL), followed by the addition of triethylamine (2 equiv.), and 2-furoyl chloride reagent (1.0 equiv.). The reaction contents are stirred overnight at room temperature. The reaction is diluted with dichloromethane (50 mL) and washed with saturated sodium bicarbonate. The organic layer is dried over magnesium sulfate, filtered, and purified by column chromatography and eluted using 3:1 hexanes in ethyl acetate. The solvents are removed *in vacuo* to give Compound **26**.

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The product from the acylation reaction **26** (1.0 equiv.) is dissolved in a solution of 50% TFA in dichloromethane (20-25 mL), and the reaction contents are stirred at room temperature (15-20 minutes; solution becomes slight reddish-orange). The reaction contents are stirred for an additional 1 hour and 20 minutes until the

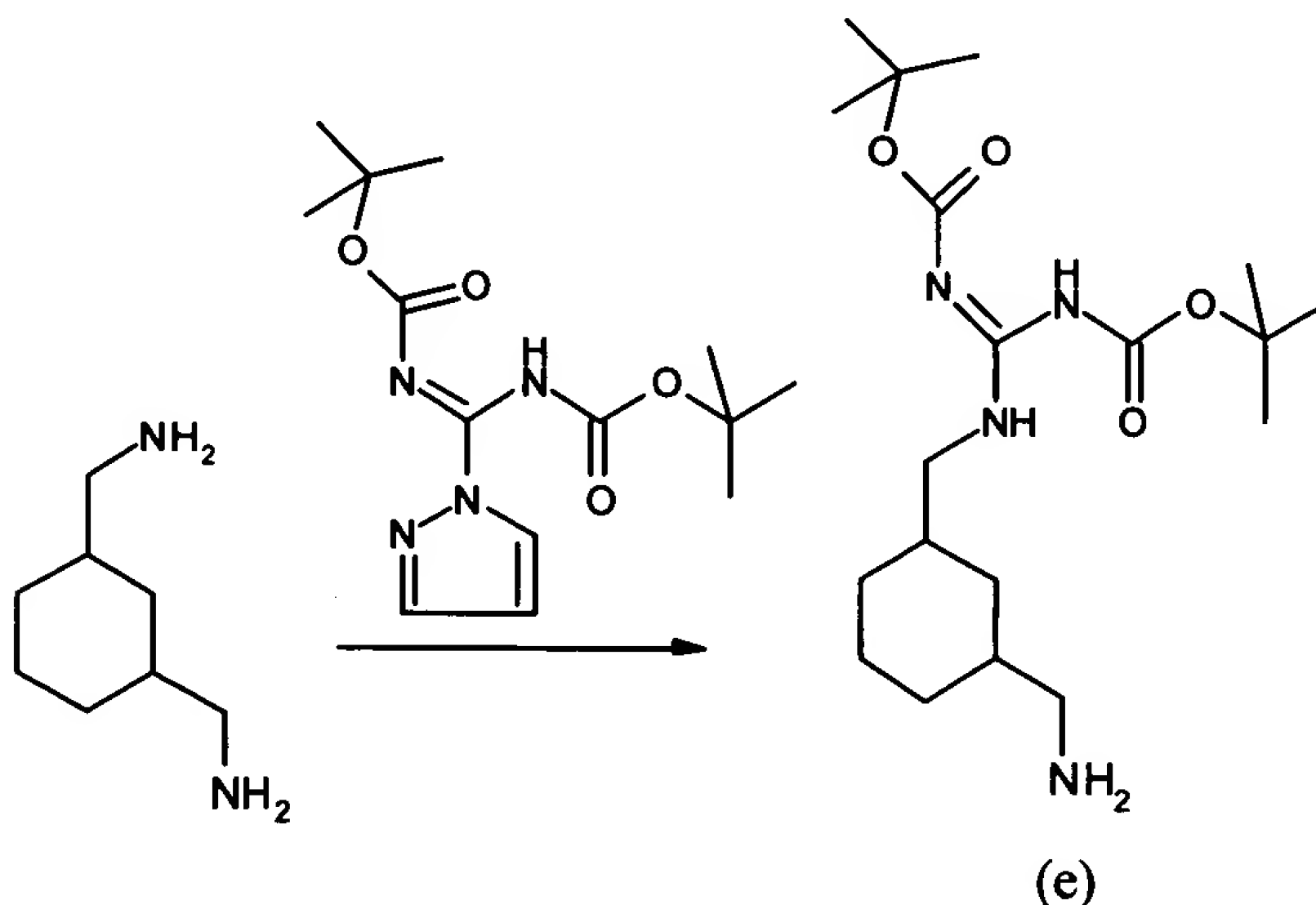
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deprotection is complete. The reaction is terminated by concentration *in vacuo*, followed by the addition of water/acetonitrile (~50 mL) and lyophilization overnight. The final compound is purified by HPLC methods. The solvents are removed *in vacuo* to give Compound **27**.

5 The following discussion relates to the preparation of exemplary Compounds (e)-(k). Compounds (e)-(k) may be used as described above to produce the corresponding deprotected (free guanidiny) compounds, through hydrolysis under acid conditions.

Preparation of 1-(N,N'-diBoc)-guanidinomethyl-3-aminomethylcyclohexane:

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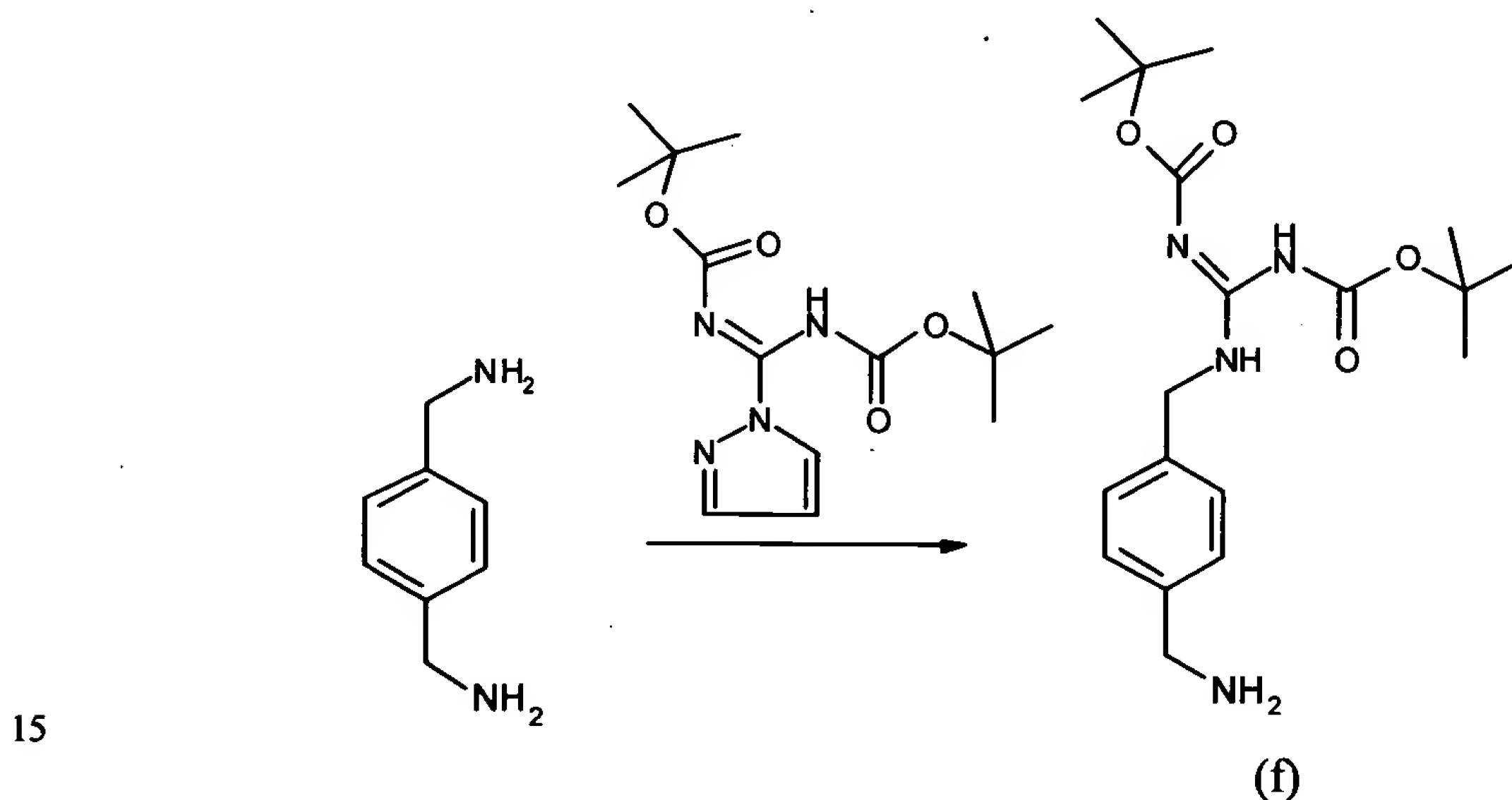
To a solution of *cis/trans*-1,3-bis-aminomethylcyclohexane (7.5 g, 52.8 mmol) in THF (30 mL) is added a solution of 1,3-bis(tert-butoxycarbonyl)-2-methyl-2-thiopseudourea (7.65 g, 26.3 mmol) in THF (40 mL) within 0.5 hour. The solution is stirred at room temperature for 5 hours. The solvent is removed under reduced pressure, and the product is purified by column chromatography on silica gel using a mixture of methylene chloride/methanol as the eluant, to afford 2.2 g (22% yield) of 1-(N,N'-diBoc)-guanidinomethyl-3-aminomethylcyclohexane (Compound (e)). ¹H NMR (CDCl₃) δ 11.53 (br s, 1H), 8.40 (br s, 1H), 3.28-3.30 (m, 2H), 2.54-2.61 (m, 2H), 1.81 (br s, 2H), 1.27-1.58 (m, 26H), 0.89 (m, 1H), 0.65 (m, 1H).

15

20

Alternatively, Compound (e) may be prepared as follows. To a solution of *cis/trans* 1,3-bis-aminomethylcyclohexane (10.0 g, 70.3 mmol) in THF (1000 mL, 0.07M) is added portionwise as a solid (over a 10-minute period) 1-H-Pyrazole-1-(N,N-bis(tert-butoxycarbonyl)carboxamidine) (21.8 g, 70.3 mmol). The solution is stirred at room temperature for 0.5 hour. The solvent is removed under reduced pressure to give a syrupy residue, which is taken up in ethyl acetate (500 mL) and washed twice with water. The layers are separated, and the product is purified by column chromatography on silica gel and eluted with 100% ethyl acetate to remove any non-polar impurities, followed by elution with 100% isopropyl alcohol to give the pure product. The solvents are removed *in vacuo* to afford 11.4 g (41 % yield) of 1-(N,N'-diBoc)-guanidinomethyl-3-aminomethylcyclohexane. ¹H NMR (CDCl₃) δ 11.53 (br s, 1H), 8.40 (br s, 1H), 3.28-3.30 (m, 2H), 2.54-2.61 (m, 2H), 1.81 (br s, 2H), 1.27-1.58 (m, 26H), 0.89 (m, 1H), 0.65 (m, 1H).

Preparation of 1-(N,N'-diBoc)-guanidinomethyl-4-aminomethylbenzene:

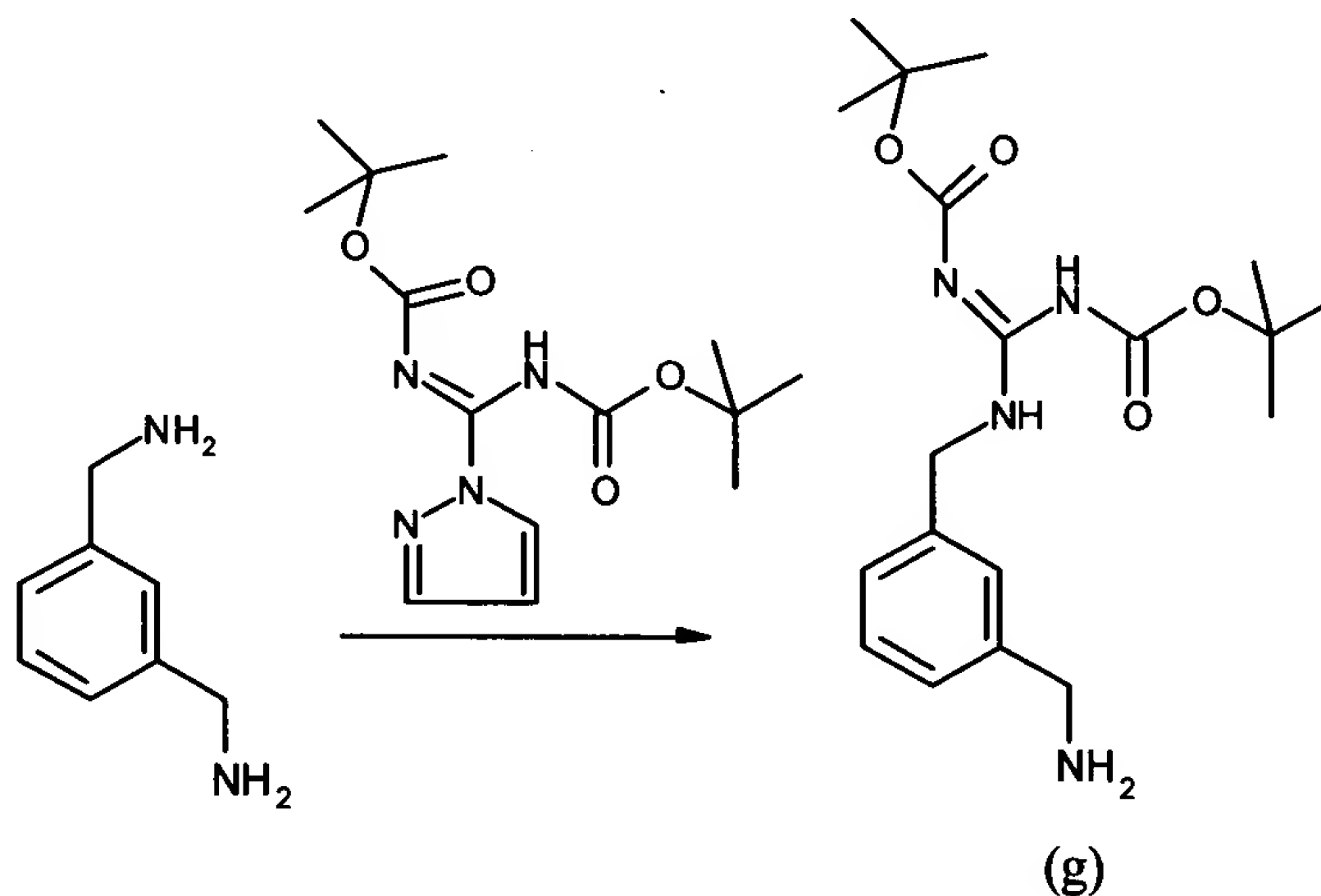


To a solution of *p*-xylylenediamine (6.44 g, 47.4 mmol) in THF (30 mL) is added a solution of 1,3-bis(tert-butoxycarbonyl)-2-methyl-2-thiopseudourea (6.63 g, 22.9 mmol) in THF (40 mL) within 0.5 hour. The solution is stirred at room temperature for 5 hours. The solvent is removed under reduced pressure, and the product is purified by column chromatography on silica gel using a mixture of methylene chloride/methanol as the eluant, to afford 8.0 g (92% yield) of 1-(N,N'-diBoc)-

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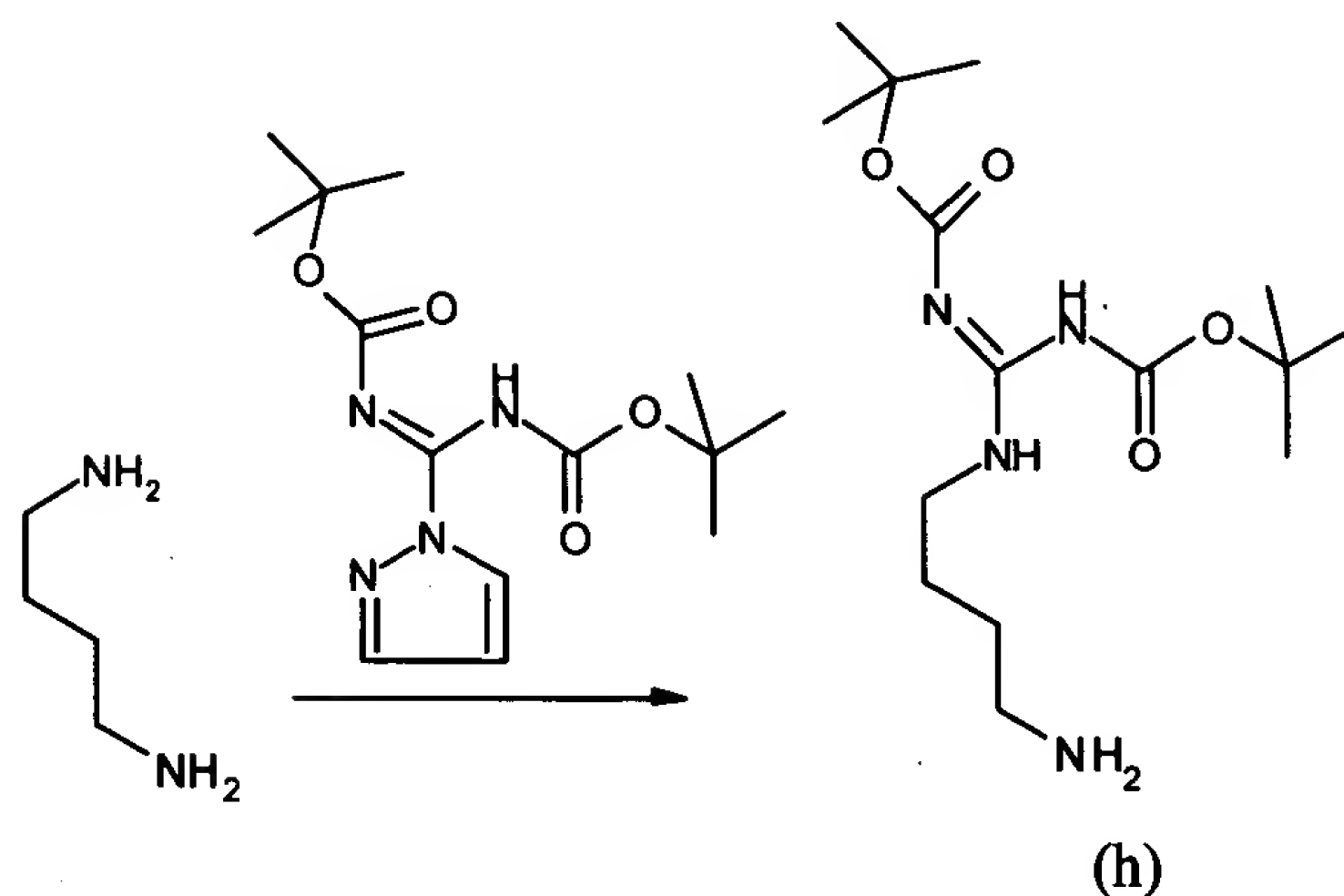
guanidinomethyl-4-aminomethyl benzene (Compound (f)). ^1H NMR (CDCl_3) δ 11.54 (br s, 1H), 8.56 (br s, 1H), 7.29 (s, 4H), 4.60 (d, 2H), 3.86 (s, 2H), 1.64 (br s, 2H), 1.52 (s, 9H), 1.48 (s, 9H).

Preparation of 1-(N,N'-diBoc)-guanidinomethyl-3-aminomethylbenzene:



To a solution of *m*-xylylenediamine (7.14 g, 52.5 mmol) in THF (30 mL) is added a solution of 1,3-bis(tert-butoxycarbonyl)-2-methyl-2-thiopseudourea (7.57 g, 26.1 mmol) in THF (40 mL) within 0.5 hour. The solution is stirred at room temperature for 5 hours. The solvent is removed under reduced pressure, and the product is purified by column chromatography on silica gel using a mixture of methylene chloride/methanol as the eluant, to afford 7.9 g (80% yield) of 1-(N,N'-diBoc)-guanidinomethyl-3-aminomethylbenzene (Compound (g)). ^1H NMR (CDCl_3) δ 11.54 (br s, 1H), 8.58 (br s, 1H), 7.19-7.34 (m, 4H), 4.62 (d, 2H), 3.86 (s, 2H), 1.83 (br s, 2H), 1.52 (s, 9H), 1.48 (s, 9H).

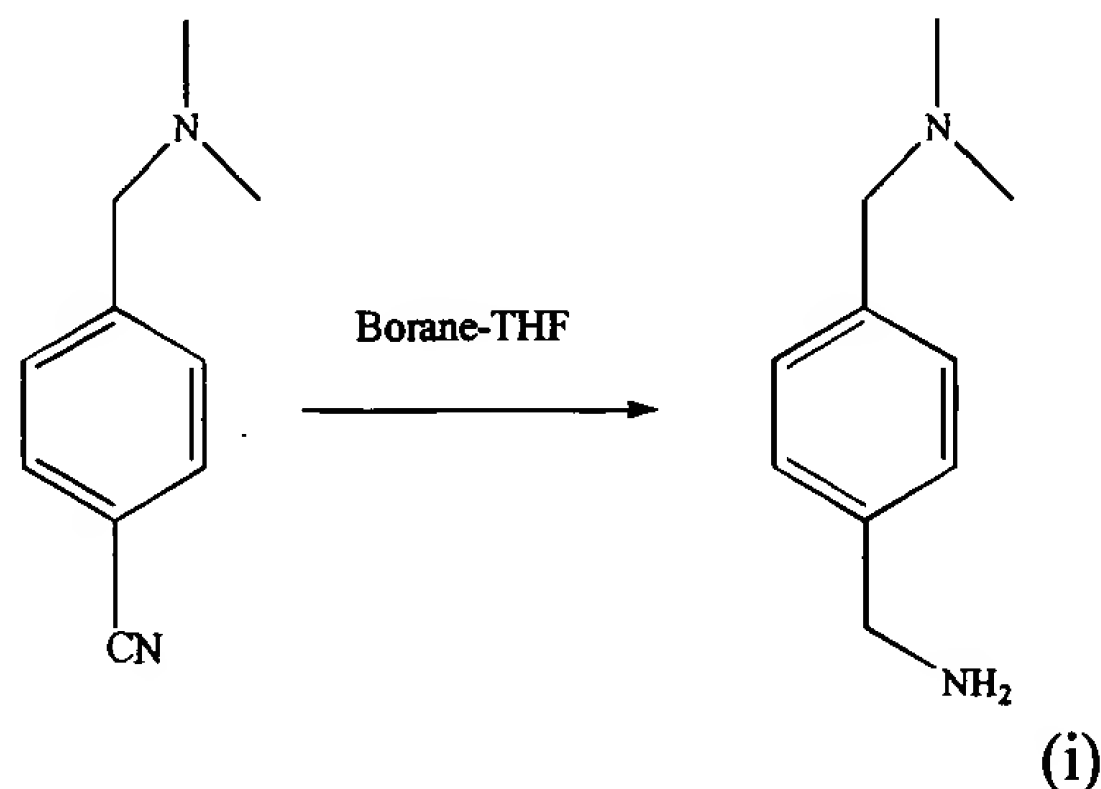
Preparation of 1-(N,N'-diBoc)-guanidine-4-aminobutane:



To a solution of 1,4-diaminobutane (4.15 g, 47.1 mmol) in THF (30 mL) is
 5 added a solution of 1,3-bis(tert-butoxycarbonyl)-2-methyl-2-thiopseudourea (6.83 g,
 23.6 mmol) in THF (40 mL) within 0.5 hour. The solution is stirred at room
 temperature for 5 hours. The solvent is removed under reduced pressure, and the
 product is purified by column chromatography on silica gel using a mixture of
 methylene chloride/methanol as the eluant, to afford 3.0 g (40% yield) of 1-(N,N'-
 10 diBoc)-guanidino-4-aminobutane (Compound (h)). ¹H NMR (CDCl₃) δ 11.49 (br s,
 1H), 8.35 (br s, 1H), 3.42-3.47 (m, 2H), 2.72-2.76 (t, 2H), 0.86-1.65 (m, 24H).

An alternate procedure for preparing Compound (h) is as follows. To a
 solution of 1,4-diaminobutane (6.0 g, 68.1 mmol) in THF (972 mL, 0.07M) is added
 portionwise as a solid (over a 10-minute period) 1-H-pyrazole-1-(N,N-bis(tert-
 15 butoxycarbonyl)carboxamidine) (21.5 g, 68.1 mmol). The solution is stirred at room
 temperature for 0.5 hour. The solvent is removed under reduced pressure to give a
 syrupy residue, which is taken up in ethyl acetate (500 mL) and washed twice with
 water. The layers are separated and the product is purified by column chromatography
 on silica gel and eluted with 100% ethyl acetate to remove any non-polar impurities
 20 and then with 100% isopropyl alcohol to give the pure product. The solvents are
 removed *in vacuo* to afford 10.0 g (44 % yield) of 1-(N,N'-diBoc)-guanidino-4-
 aminobutane. ¹H NMR (CDCl₃) δ 11.49 (br s, 1H), 8.35 (br s, 1H), 3.42-3.47 (m,
 2H), 2.72-2.76 (t, 2H), 0.86-1.65 (m, 24H).

Preparation of 1-N,N-dimethylaminomethyl-4-aminomethylbenzene:

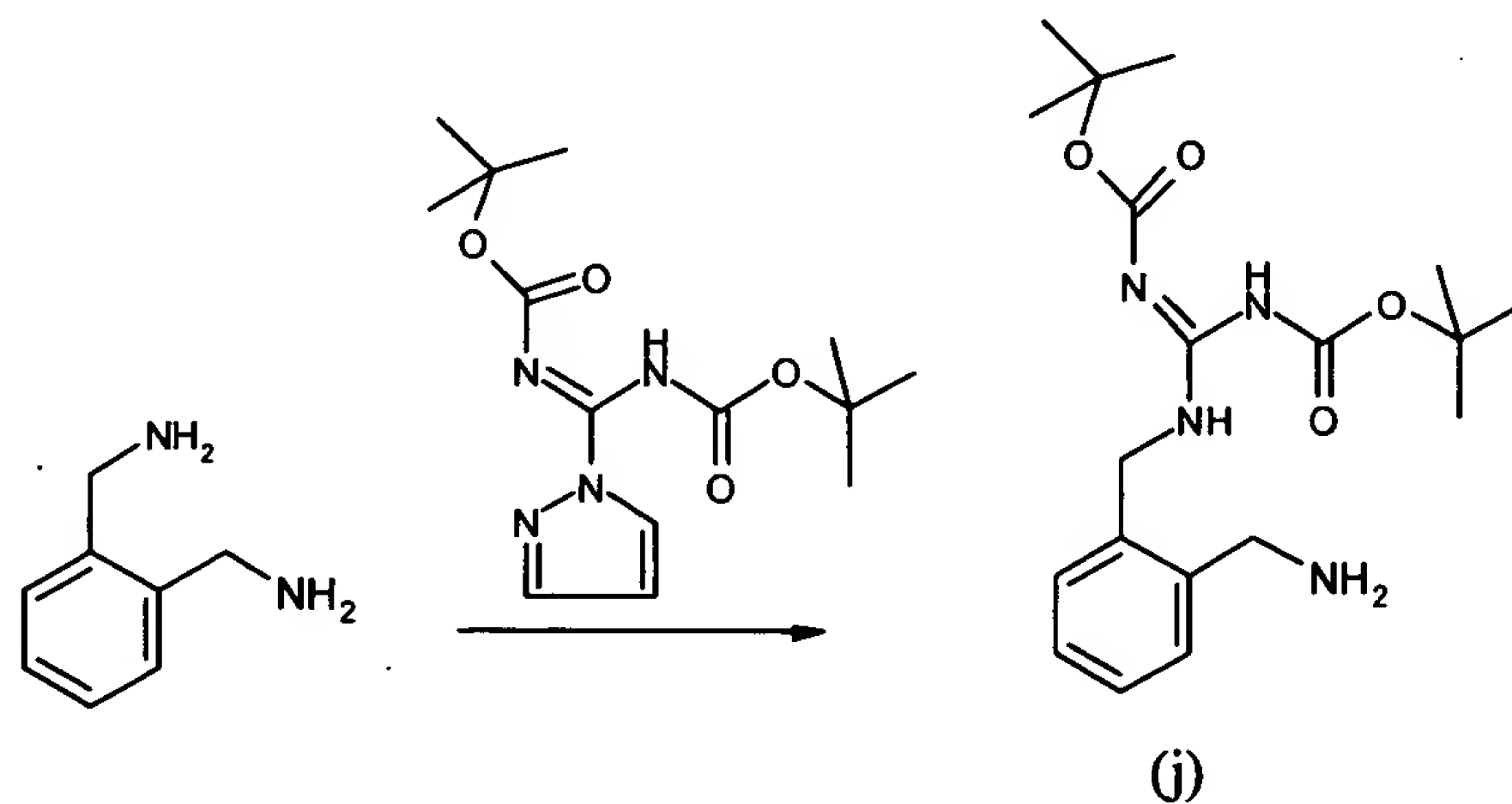


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To a solution of 1-N,N-dimethyl aminomethyl-4-carbonitrile benzene (4.8 g, 30 mmol) in THF is added a solution of 1 M borane tetrahydrofuran complex (90 mL). The mixture is heated at reflux temperature for 16 hours under nitrogen. After cooling to room temperature, a 1M solution of HCl in methanol (100 mL) is added. The reaction mixture is heated at reflux for 3 hours. The product, which precipitates, is collected by filtration, washed with diethyl ether, and dried *in vacuo* to give 5.9 g (83% yield) of the product as the hydrochloride salt (Compound (i)): ¹H NMR (DMSO-d₆) δ 8.65 (br s, 3H), 7.55 (dd, 4H), 4.25 (s, 2H), 3.98 (s, 2H), 2.62 (s, 6H).

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Preparation of 1-(N,N'-diBoc)-guanidinomethyl-2-aminomethylbenzene:



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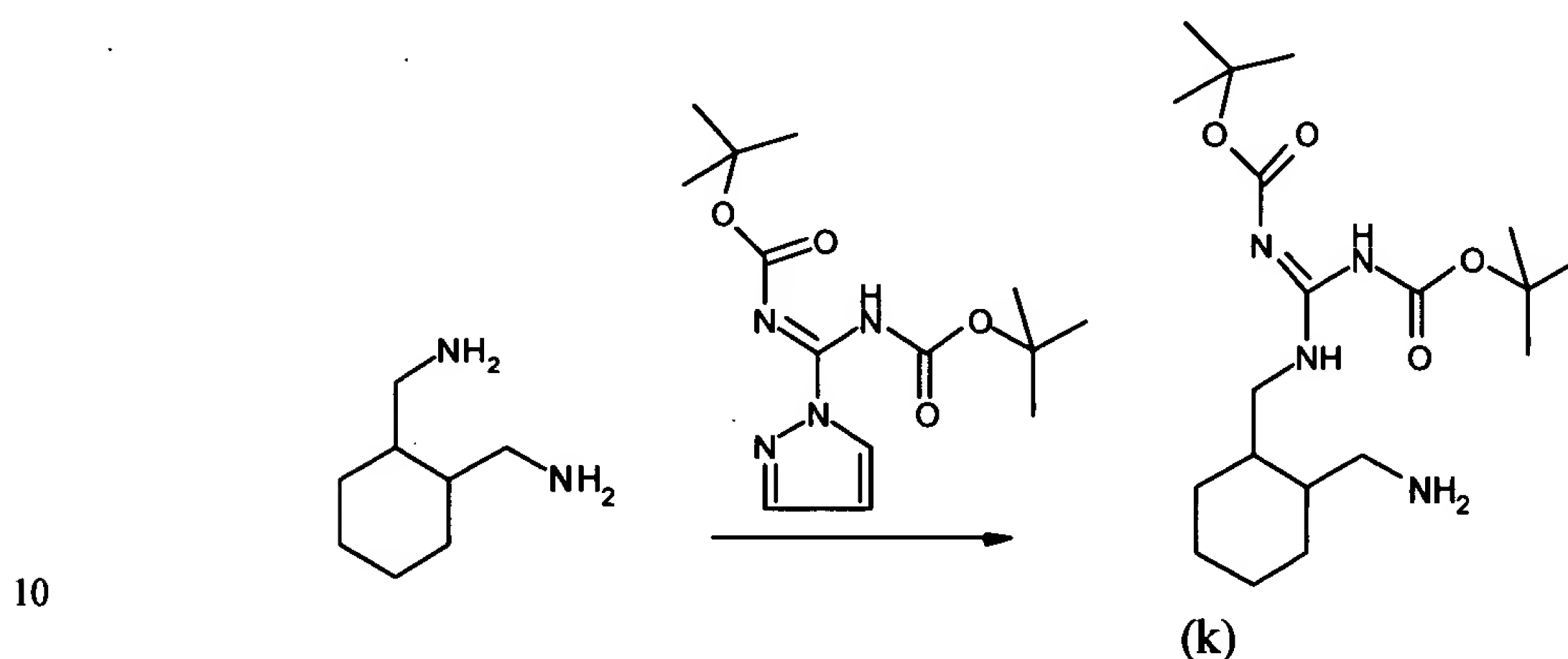
To a solution of *o*-xylylenediamine (7.14 g, 52.5 mmol) in THF (30 mL) is added a solution of 1,3-bis(tert-butoxycarbonyl)-2-methyl-2-thiopseudourea (7.57 g, 26.1 mmol) in THF (40 mL) within 0.5 hour. The solution is stirred at room

20

temperature for 5 hours. The solvent is removed under reduced pressure, and the product is purified by column chromatography on silica gel using a mixture of methylene chloride/methanol as the eluant, to afford 1-(N,N'-diBoc)-guanidinomethyl-3-aminomethyl benzene (Compound (j)).

5 Alternatively, Compound (j) may be prepared in a manner analogous to the alternative preparation described above for Compound (e).

Preparation of 1-(N,N'-diBoc)-guanidinomethyl-2-aminomethylcyclohexane:

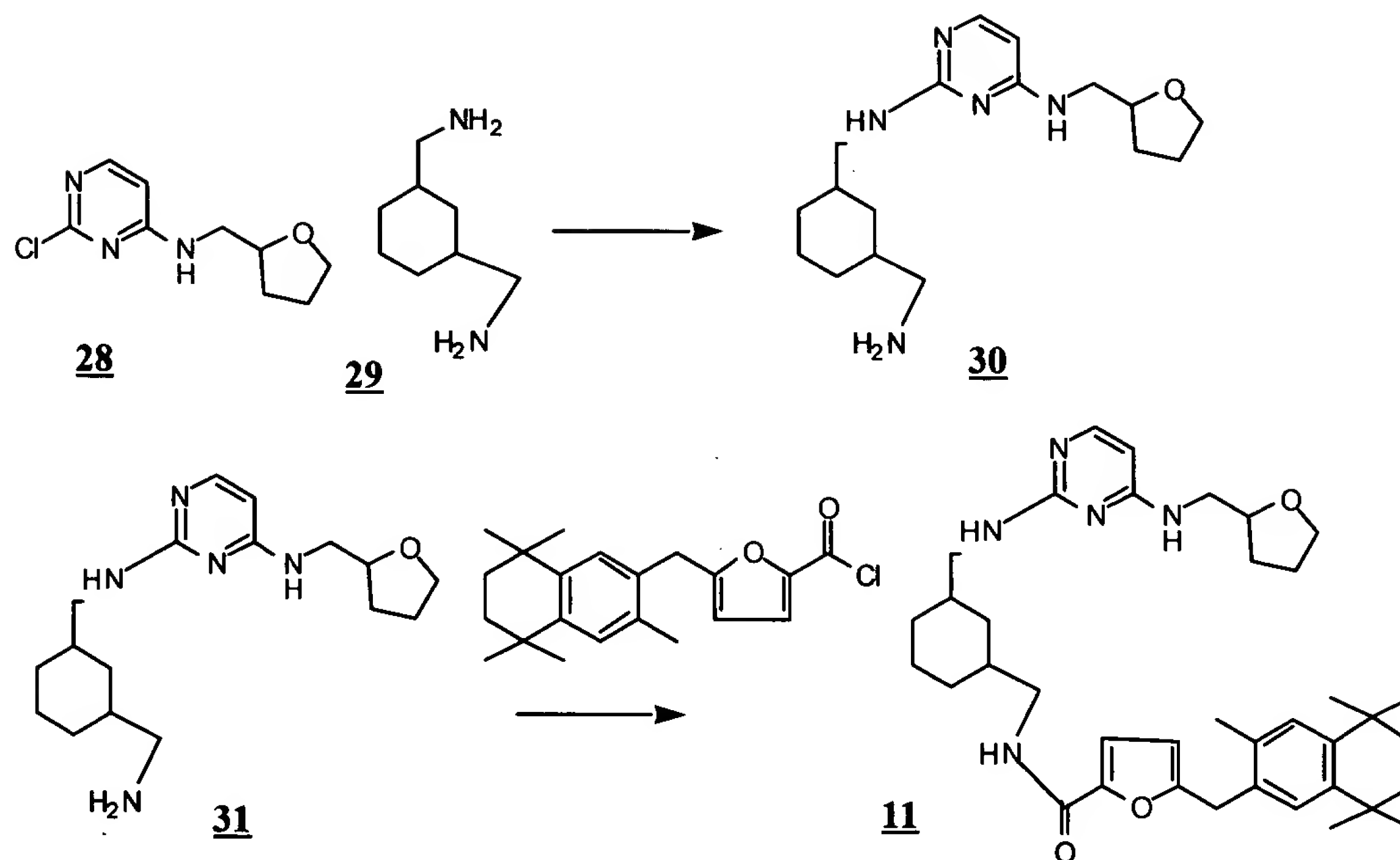


To a solution of *cis/trans*-1,2-bis-aminomethylcyclohexane (7.5 g, 52.8 mmol) in THF (30 mL) is added a solution of 1,3-bis(tert-butoxycarbonyl)-2-methyl-2-thiopseudourea (7.65 g, 26.3 mmol) in THF (40 mL) within 0.5 hour. The solution is stirred at room temperature for 5 hours. The solvent is removed under reduced pressure, and the product is purified by column chromatography on silica gel using a mixture of methylene chloride/methanol as the eluant, to afford 1-(N,N'-diBoc)-guanidinomethyl-2-aminomethylcyclohexane (Compound (k)).

20 Alternatively, Compound (k) may be prepared in a manner analogous to the alternative preparation described above for Compound (e).

D. Pyrimidine Compounds

Pyrimidines can be utilized according to the following procedures:



A general procedure for the preparation of pyrimidine containing compounds is as follows. To a solution of 1,3 diamine **29** in THF is added **28** and the contents
 5 refluxed for 12 hours. The solvents are removed *in vacuo* and the desired adduct purified by column chromatography. Pure **31** is acylated according to the general procedure given above to give **11**.

As skilled artisans will appreciate, a variety of compounds according to the invention may be prepared based on the above teachings. The chemical reactions
 10 described above have general applicability to the preparation of the GnRH agents of the invention. Thus, other GnRH agents may be similarly prepared by suitable modification as will be readily appreciated by those skilled in the art, e.g., by protection of interfering groups, by adapting for use with other conventional reagents, and/or by routine modifications of reaction conditions.

15 IN VITRO PHARMACOLOGY RADIOLIGAND BINDING.

Cell membranes prepared from human embryonic kidney 293 cells stably transfected with cDNA for the human GnRH receptor were suspended in binding assay buffer containing: 50 mM HEPES, 1 mM EDTA, 2.5 mM MgCl₂, and 0.1% bovine serum albumin. Membranes (5 - 50 µg total protein per well containing approximately

10-100 fmol of GnRH receptor) were incubated in duplicate in 96-well plates in 200 μ l total volume with 125 I-GnRH-A (approximately 0.05 nM) and test compounds for one hour at room temperature. All compounds were diluted in 1% DMSO (final assay concentration) in binding assay buffer. Nonspecific binding was determined in the
5 presence of 100 nM GnRH. Reactions were terminated by rapid filtration onto 96-well Packard GF/C filters soaked in 0.1% polyethyleneimine. Filters were washed three times with PBS buffer, dried and counted on a Packard Topcount by liquid scintillation counting.

Assay conditions were identical for assessing compound activities at other
10 species. A similar number of GnRH receptors was utilized for each species assay. For rat GnRH receptor binding, membranes were prepared from rat pituitary and approximately 25-30 μ g/well of total membrane protein were utilized. For bovine GnRH receptor binding, membranes were prepared from bovine pituitary and utilized at 40-50 μ g/well. For mouse GnRH receptor binding, membranes were prepared from
15 293 cells stably expressing mouse GnRH receptors and were utilized at approximately 25-30 μ g/well. IC_{50} values for control peptides and test compounds were calculated utilizing GraphPad Prism™ software. The result of a radioligand binding experiment is shown in Figure 1. Table 1 shows mean values from multiple experiments of the affinities of various peptide and non-peptide compounds at GnRH receptors from four
20 species.

**Figure 1. Effects of Compounds
on ^{125}I -GnRH-A Binding to
hGnRH Receptors in HEK-293
Cell Membranes**

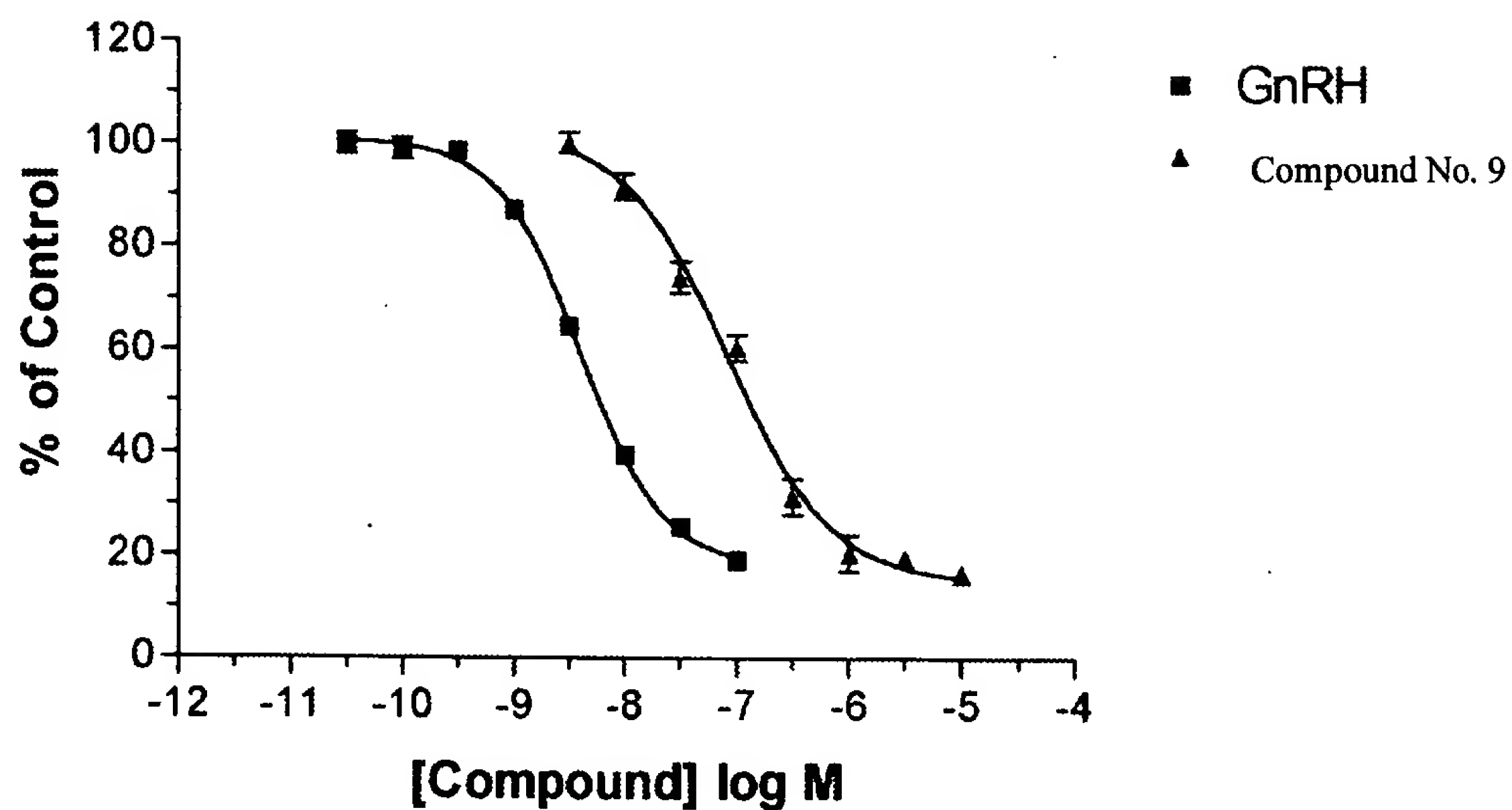


Figure 1. Effects of compounds on ^{125}I -GnRH-A binding to hGnRH receptors expressed in HEK-293 cells. The ability of GnRH (squares) and **9** (triangles) to displace ^{125}I -GnRH-A (approximately 0.05 nM) binding to hGnRH receptors was examined. Values shown are from one representative experiment performed in duplicate.

Various compounds of the Formula I were synthesized according to the general reaction scheme generally described above. Crude compounds were tested using the competitive radioligand binding assay described above. Results of the GnRH competitive binding assay are shown in the table (each compound tested at 1 or 10 μ M).

Table 1.

Compound	Human IC ₅₀ (nM)	Bovine IC ₅₀ (nM)	Rat IC ₅₀ (nM)	Mouse IC ₅₀ (nM)
GnRH	7.2 \pm 1.5	13 \pm 2	33 \pm 1.9	11 \pm 2
GnRH-A	0.34 \pm 0.06	0.3 \pm 0.05	0.49 \pm 0.1	0.22 \pm 0.03
Antide	0.67 \pm 0.09	0.15 \pm 0.02	0.19 \pm 0.04	0.25 \pm 0.05
<u>9</u>	220 \pm 33	3800 \pm 220	680 \pm 120	2300 \pm 460
<u>10</u>	130 \pm 24	1500 \pm 480	390 \pm 10	1400 \pm 440
<u>11</u>	190 \pm 40	320 \pm 10	9.0 \pm 0.3	50 \pm 10
<u>12</u>	230 \pm 37	10400 \pm 3000	3080 \pm 630	7130 \pm 1350
<u>13</u>	110 \pm 20	530 \pm 100	60 \pm 8	120 \pm 20
<u>14</u>	80 \pm 4	1050 \pm 30	60 \pm 15	290 \pm 70
<u>15</u>	100 \pm 17	1000 \pm 240	70 \pm 16	220 \pm 50
<u>16</u>	30 \pm 6	4380 \pm 510	560 \pm 50	1290 \pm 210
<u>17</u>	80 \pm 20	670 \pm 120	30 \pm 4	80 \pm 20
<u>18</u>	55 \pm 11	460 \pm 90	40 \pm 3	115 \pm 25
<u>19</u>	50 \pm 3	ND	ND	ND
<u>20</u>	8.0 \pm 0.9	ND	ND	ND

Values are means \pm SE of at least three experiments performed in duplicate.

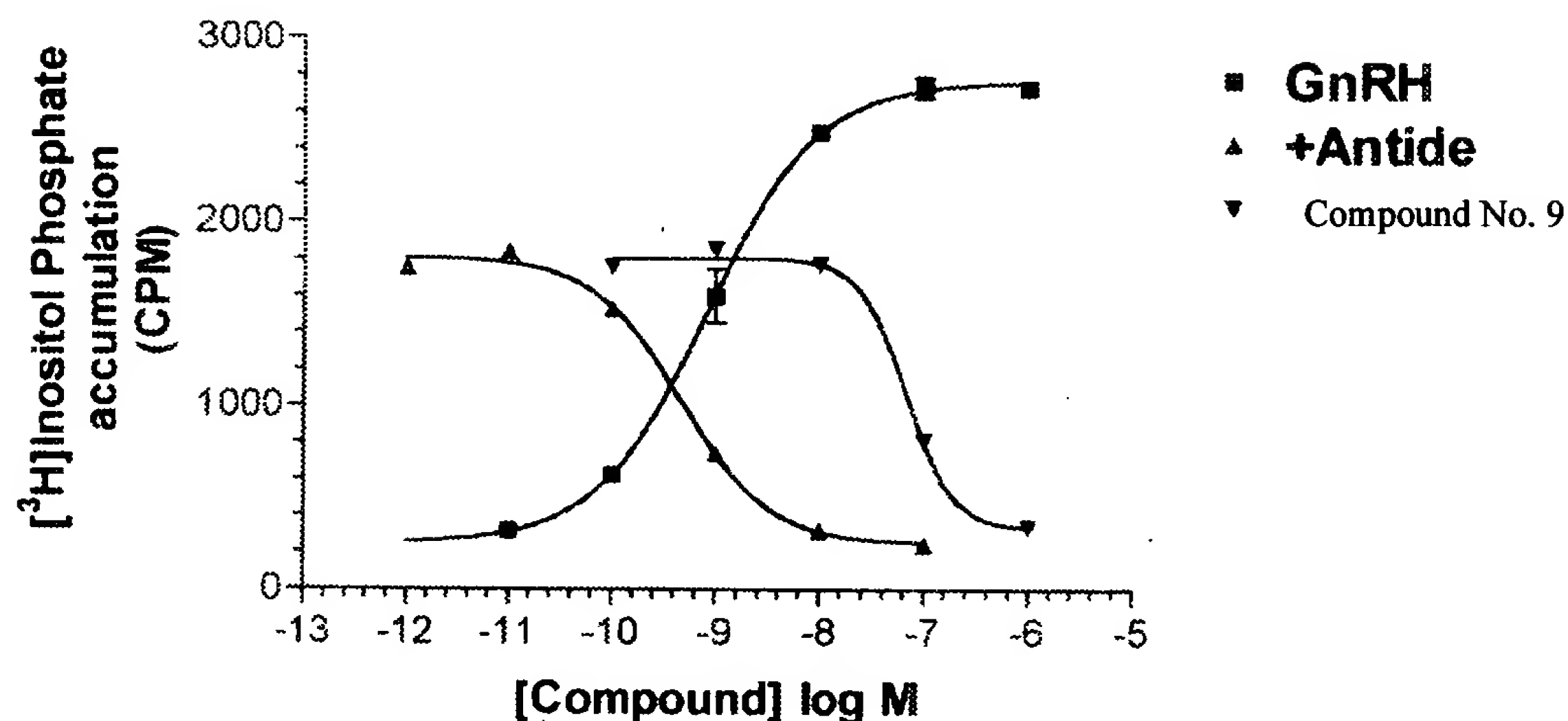
ND = not determined.

TOTAL INOSITOL PHOSPHATES MEASUREMENT.

To assess the activity of the compounds as agonists or antagonists, an assay measuring accumulation of total inositol phosphates was employed. 293 cells containing the hGnRH receptor were plated onto 24-well plates (approximately 200,000 cells/well) using DMEM media. The following day, cells were loaded with

[³H]myoinositol (0.5 Ci/ml) for 16-18 hours in inositol-free medium. The medium was aspirated and the cells rinsed with serum-free DMEM. The medium was aspirated and the cells were then treated with test compounds or vehicle for 30 minutes at 37°C. A half-maximal concentration of GnRH (1 nM) or vehicle was then added to the cells
5 and allowed to equilibrate at 37°C for 45 minutes. The media was replaced with ice-cold 10mM formic acid, which stopped the reaction and also served to extract cellular lipids. Inositol phosphates were separated by ion-exchange chromatography on Dowex columns, which were washed with 2.5 mL of 10 mM myoinositol and 10 mM formic acid. The columns were then washed with 5 mL of 60mM sodium formate and 5 mM
10 borax, and total inositol phosphates were eluted with 5 mL 1M ammonium formate, 0.1 M formic acid. The column eluates were added to liquid scintillation vials containing 15 ml of scintillation cocktail and were counted by liquid scintillation counting. The result of a typical experiment is shown in Figure 2.

Figure 2. Effects of Compounds on GnRH-stimulated (1 nM) total inositol phosphate accumulation in 293 cells expressing the hGnRH receptor



Compound	EC ₅₀ (nM)	K _b (nM)
GnRH	0.83	
Antide		0.2
AXC05885		30

Figure 2. Effects of compounds on GnRH-stimulated total inositol phosphate accumulation in HEK-293 cells expressing the hGnRH receptor. The ability of the peptide antagonist, Antide, and non-peptide compound **9** to block GnRH-stimulated increases in [³H]inositol phosphates was examined. Neither compound alone stimulated an increase in total [³H]inositol phosphates (not shown), but both compounds were able to inhibit the stimulation mediated by a half-maximal concentration of GnRH peptide. GnRH alone dose-dependently increased [³H]inositol phosphate accumulation with an EC₅₀ of approximately 0.8 nM. In the experiment shown, the K_b values of Antide and compound **9** were determined by the method of Cheng and Prusoff (*Biochem. Pharmacol.* 22:3099-3108, 1973). Values shown are from one experiment performed in duplicate.

IN VIVO PHARMACOLOGY ANIMAL EFFICACY STUDIES

Experimental Protocol: Male Sprague-Dawley (225-250 g) rats were castrated and allowed 10 days post-operative recovery. Ten days post castration animals were
5 instrumented with indwelling femoral venous and arterial catheters to facilitate remote infusions and blood sampling. On the day of the experiment, animals were allowed to acclimate to the procedure room while residing in their home cage. Basal blood samples were drawn from all animals. Following basal sampling, either vehicle (10% DMSO, 10 % cremophor/saline), Antide (1.0 µg) or compound 11 (10 mg/kg) was
10 administered intravenously. Blood samples were drawn 10, 60, 90, 120, 180, 240 minutes after injections. Blood was centrifuged, serum collected and stored in -70° freezer until assayed. Serum samples were analyzed using DSL-4600 ACTIVE LH coated-tube immunoradiometric assay kit from Diagnostic Systems Laboratories, Inc.

Results and discussion: Removal of the gonads eliminates the negative
15 feedback of testosterone on the hypothalamus, resulting in elevated GnRH and consequently elevated LH. Figure 3 illustrates the plasma levels of both LH and testosterone in control and castrated rats 10 days after surgery. In these rats, a GnRH antagonist would be expected to reduce GnRH mediated elevations of LH levels. Antide, a peptide GnRH antagonist, reduces LH in the castrated rat model (Figure 4).
20 Compound 11, a small-molecule GnRH antagonist, also suppresses LH in the castrated rat model (Figure 4).

Figure 3

**Characterization of a Castrated Rat Model
used to Evaluate GnRH Antagonists**

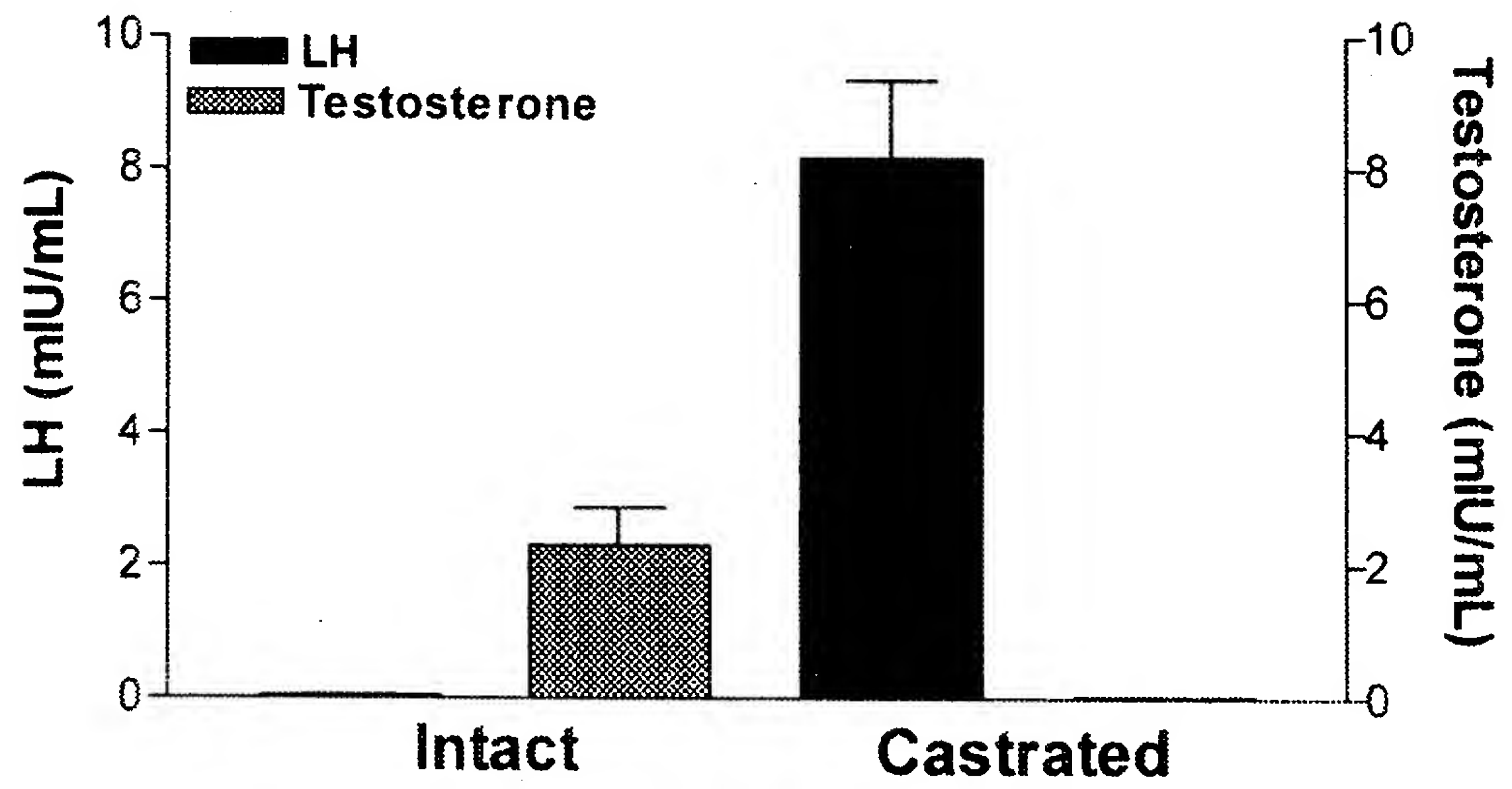
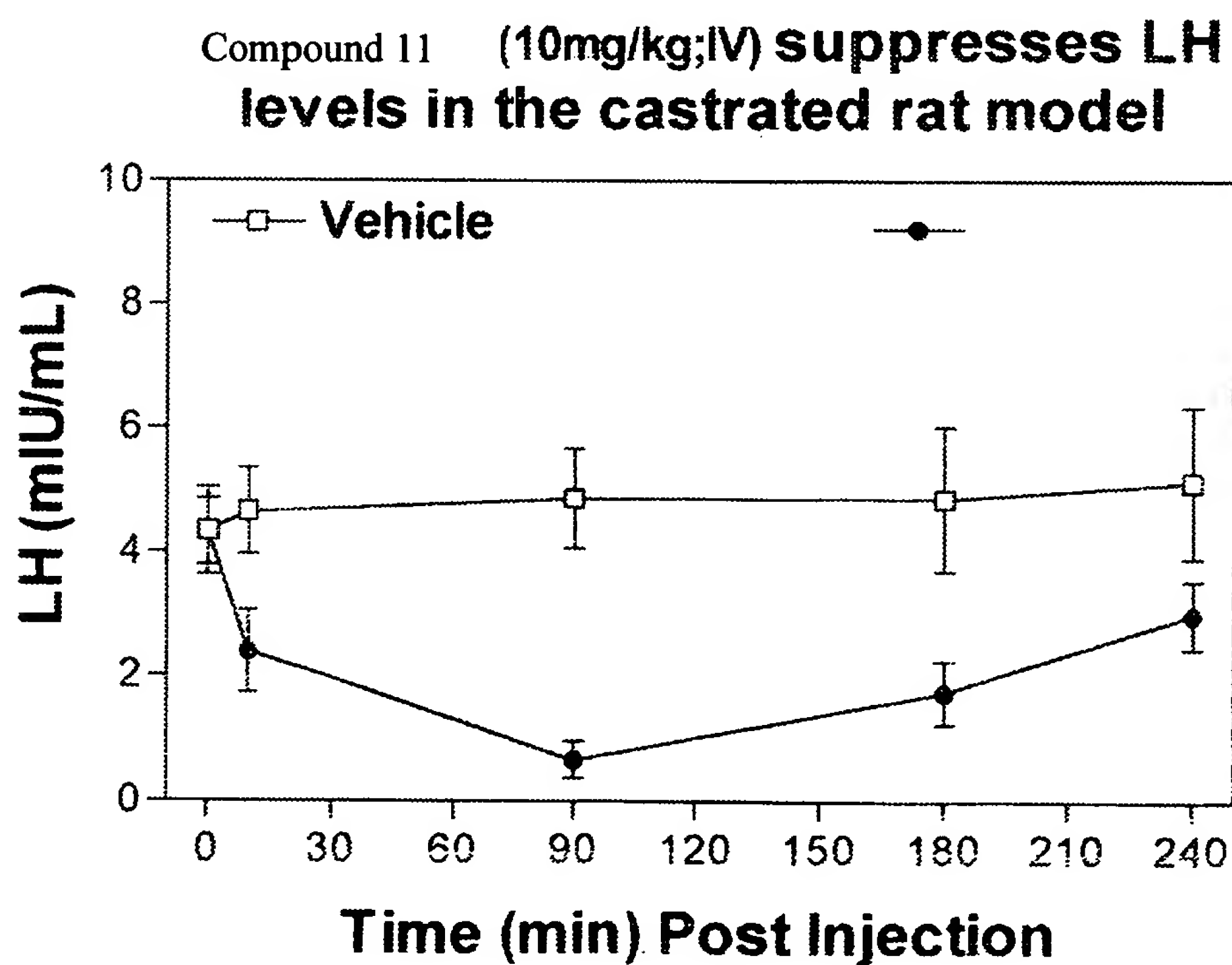
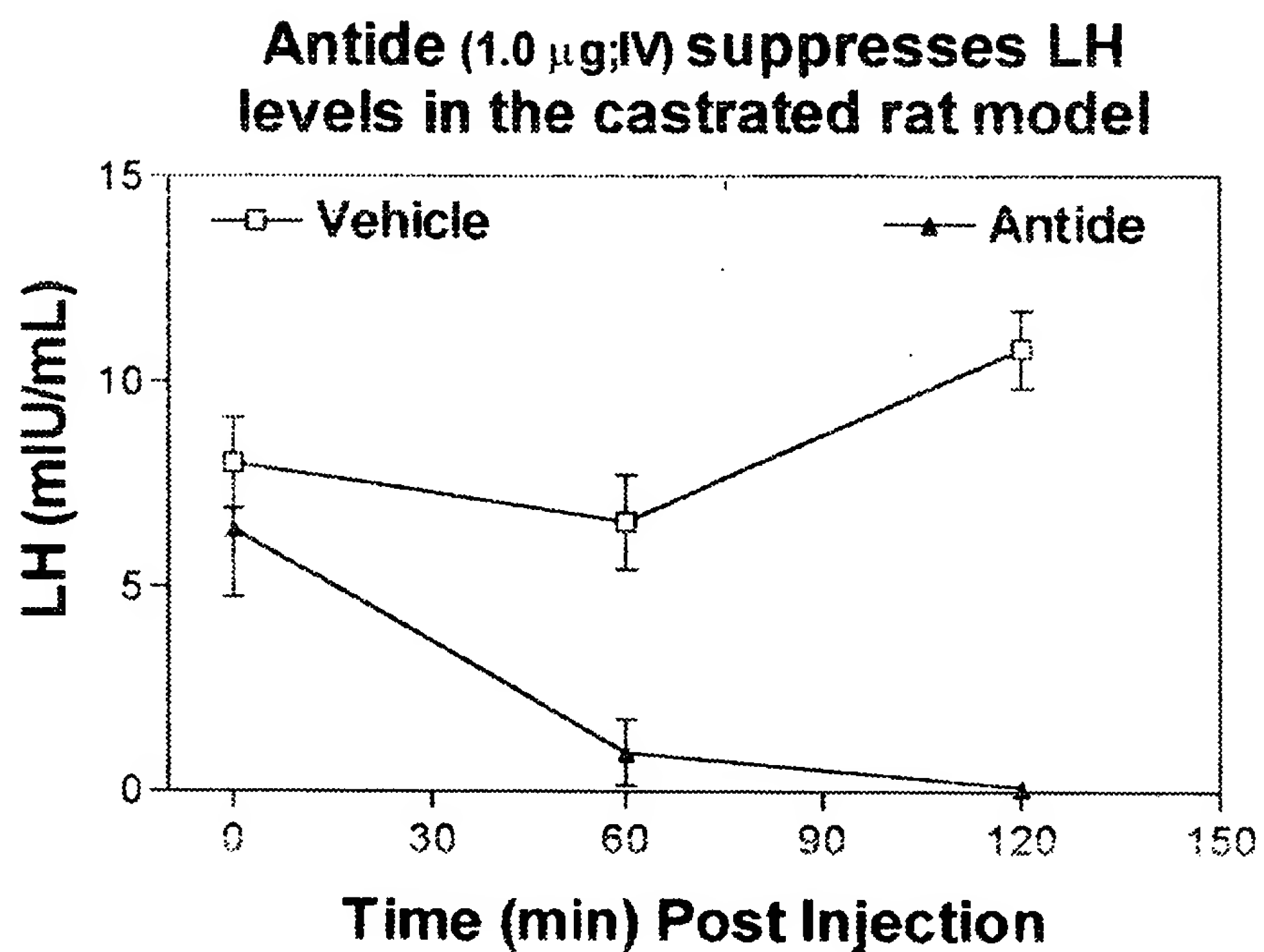


Figure 4



PHARMACOKINETIC STUDIES

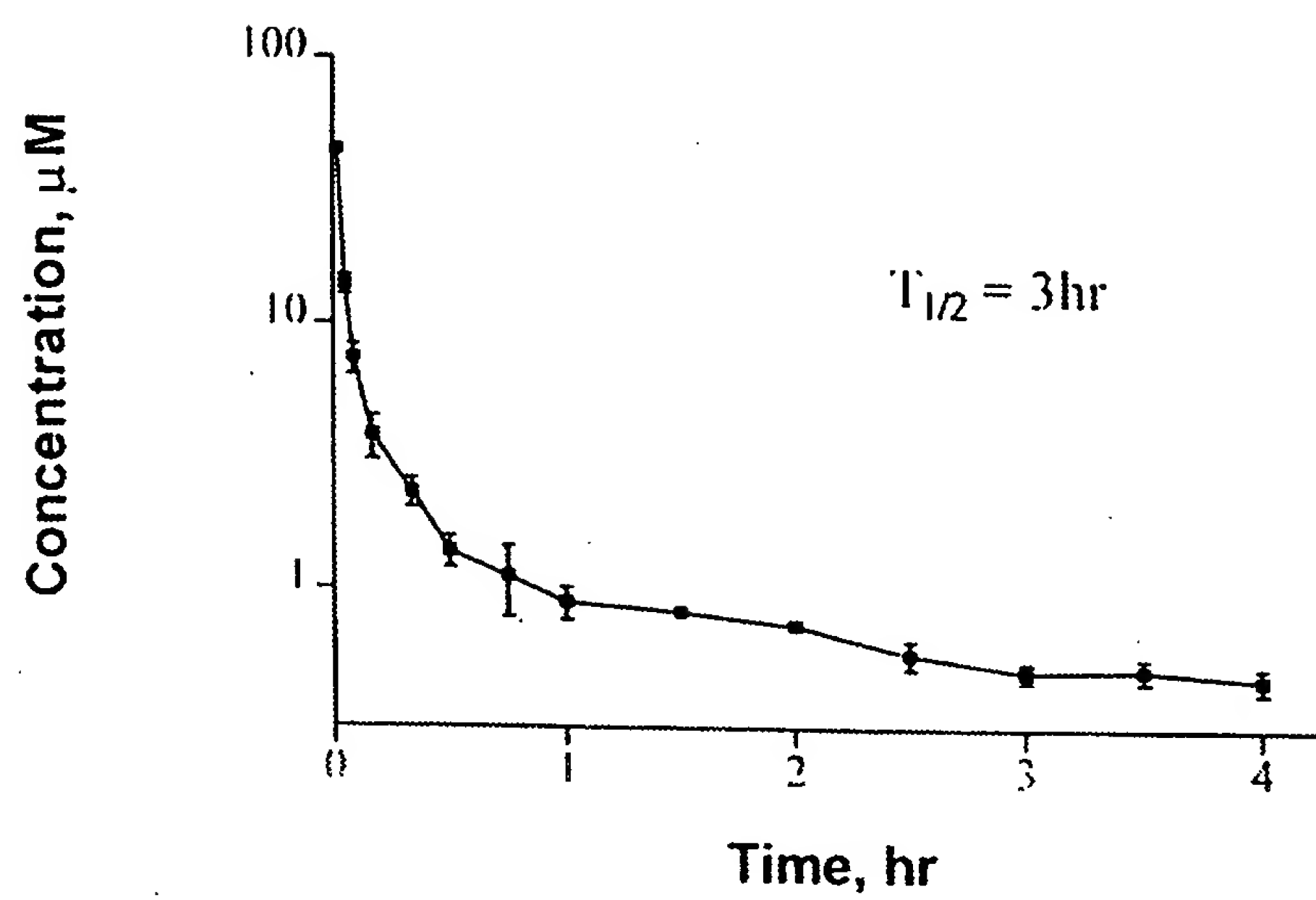
Experimental protocol: Rats were prepared with intravenous catheters inserted in the superior vena cava through the incision in the right external jugular vein and allowed to recover. Drugs were dissolved in a mixture of 10% DMSO, 10% cremaphor, and 80% saline and administered i.v. at a dose of 10 mg/kg. Blood samples were taken at the times indicated, and the compounds were extracted from 0.2 mL of plasma with butyl chloride containing an internal standard. Samples were analyzed by HPLC on a Beta-Basic C18 4x50 mm column using a gradient of 40-80 % acetonitrile in 10 mM ammonium phosphate buffer at a flow rate of 1 ml/min. Sample detection was by UV absorbance at 260 nm.

Results and Discussion: Compound 11, which has excellent affinity at the rat GnRH receptor, had a half life in rat plasma of approximately three hours and had a concentration in plasma of 100-200 nM four hours after i.v. injection (Figure 5).

Figure 5

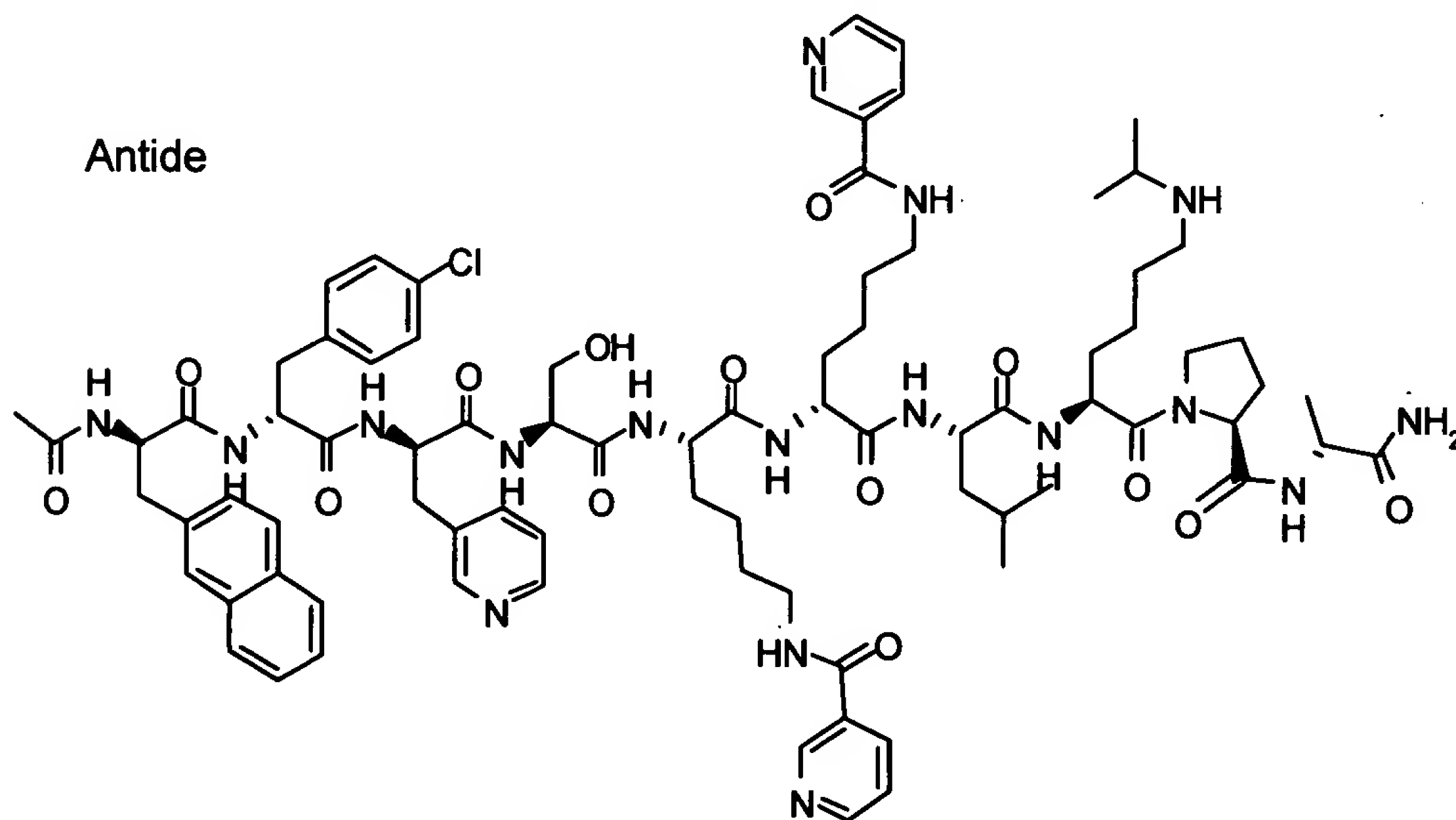
Compound 11

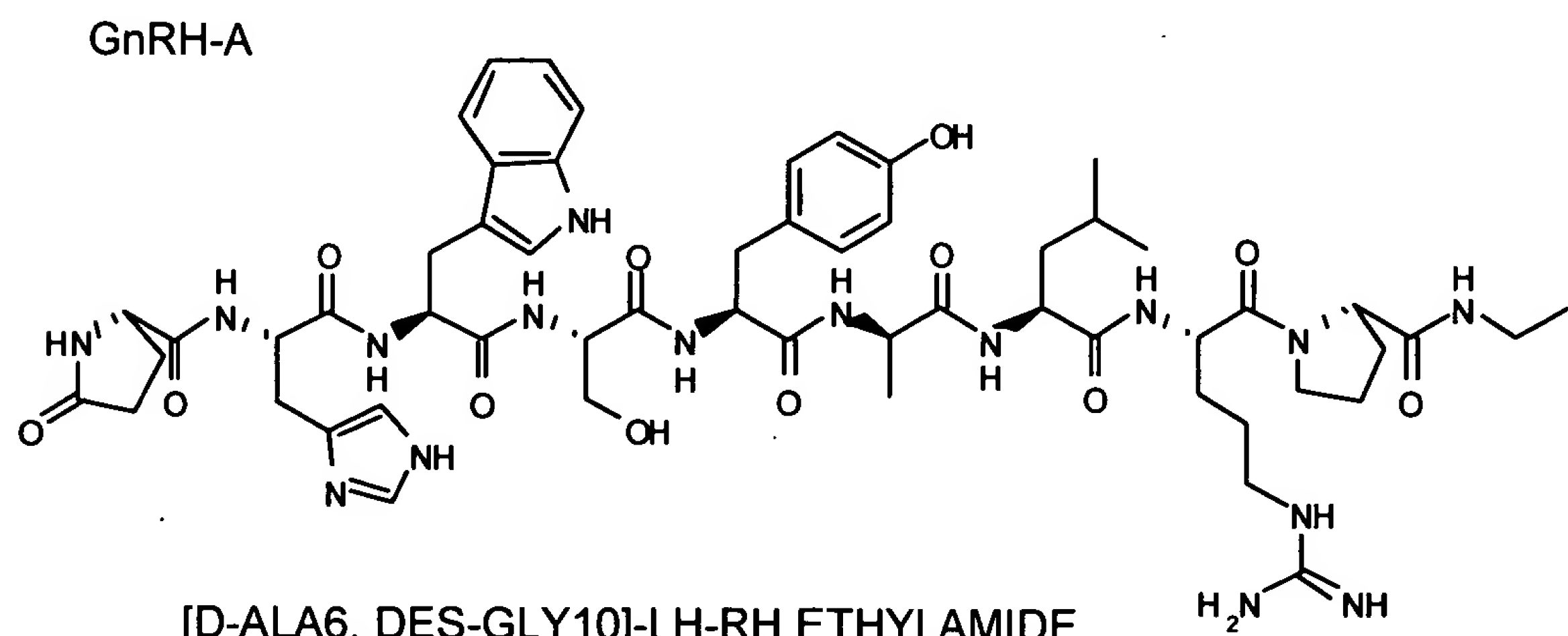
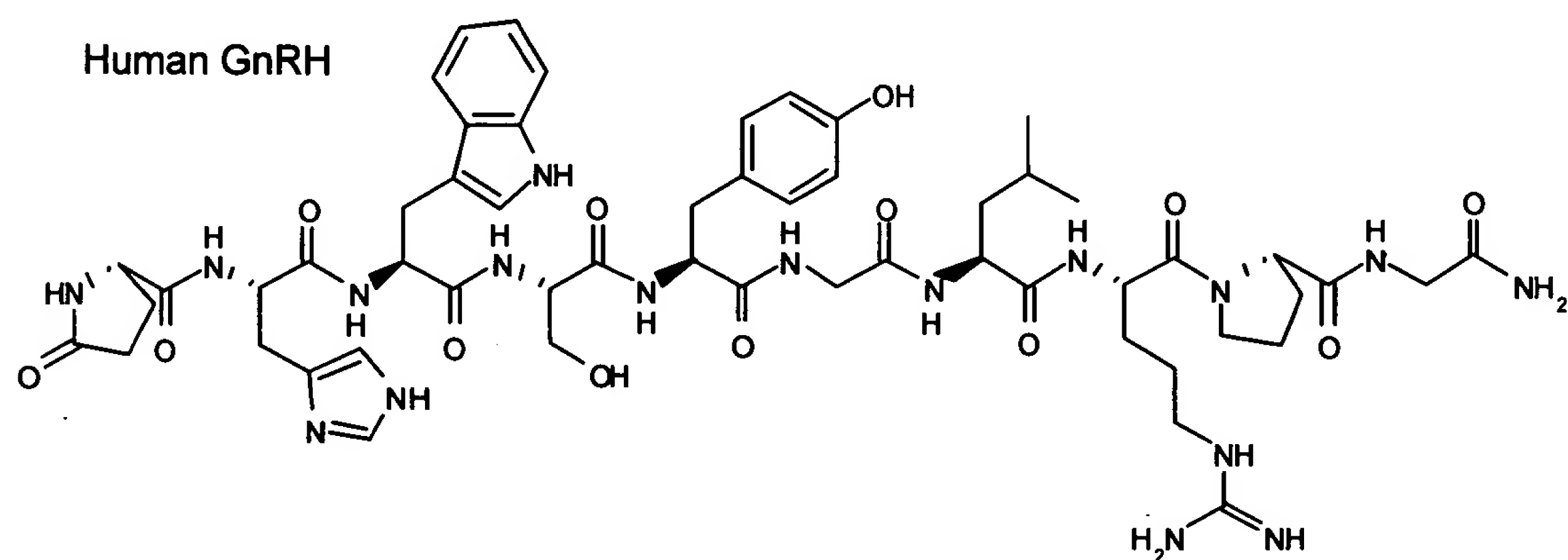
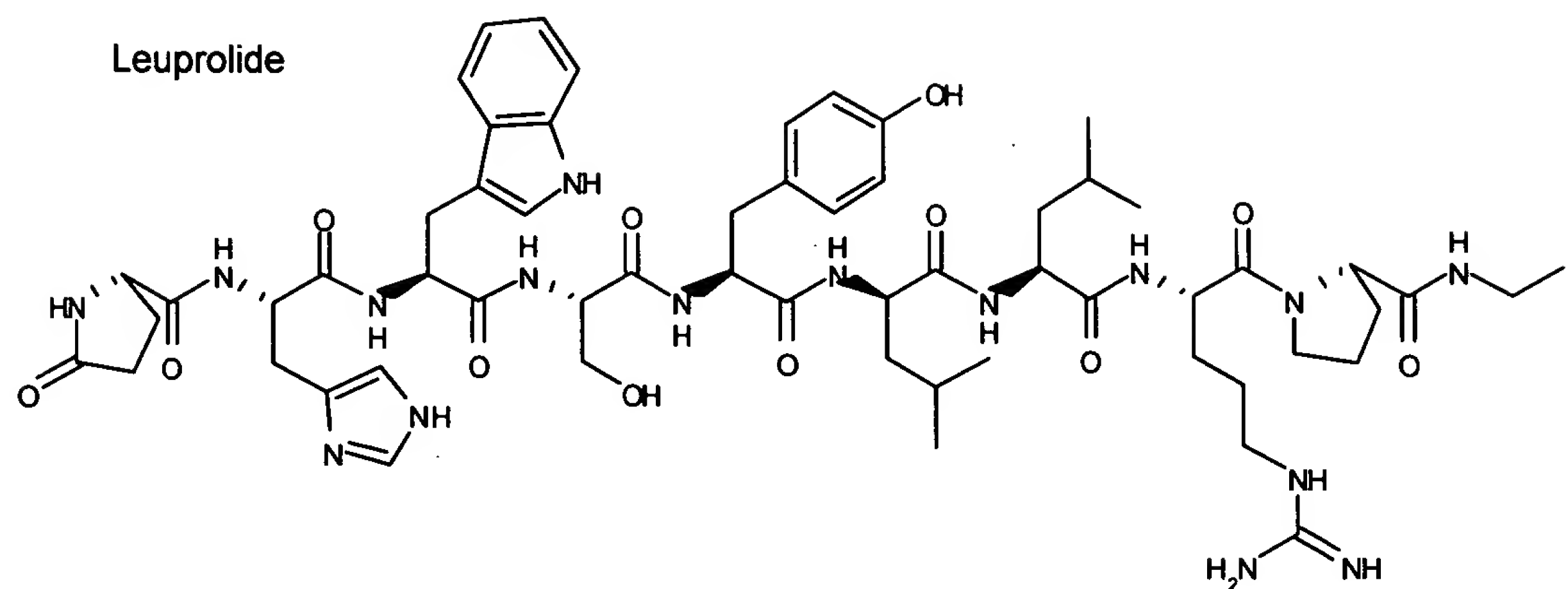
Dose - 10 mg/kg; n=3



Binding of the reference peptides to rat, mouse, bovine and human GnRH receptors are in good agreement with those reported in the literature. Non-peptide compounds of the invention show marked species differences in their binding profile. Several of these compounds exhibit high affinity (<100 nM) at the human GnRH receptor. Functionally, all of these non-peptide compounds assessed for activity in an inositol phosphate assay act as antagonists of GnRH-stimulated total inositol phosphate accumulation in cells containing recombinant human GnRH receptors. Intravenous administration of compound 11 reduced plasma levels of LH in castrated male rats, a model for chronically elevated plasma LH levels. This compound has a half life of three hours, and the plasma concentration correlated with efficacy. Taken together, these data suggest that these non-peptide compounds should have utility as GnRH receptor antagonists.

Peptide Agonists and Antagonists Used as Reference Compounds:





[D-ALA6, DES-GLY10]-LH-RH ETHYLAMIDE

5

Cell Culture

GH₃ cells stably transfected with the rat GnRH receptor (GGH₃) were provided by Dr. William Chin (Harvard Medical School, Boston, MA). These cells have been extensively characterized previously (Kaiser et al., 1997). These cells were grown in

low glucose Dulbecco's modified Eagle's medium (DMEM) containing: 100U/mL penicillin/streptomycin, 0.6 g/L G418 and 10% heat-inactivated fetal bovine serum (FBS).

The cDNA for the human GnRH receptor was cloned into the plasmid expression vector, pcDNA 3 (In Vitrogen), and stably transfected into HEK 293 cells (hGnRH-R/293). This cell line was provided by Dr. Stuart Sealton, Mount Sinai Medical School, New York, NY. These cells were grown in DMEM supplemented with 0.2 g/L G418 100 U/ml penicillin/streptomycin and 10% FBS. Both GGH₃ and hGnRH-R/293 cells were utilized for both for both total inositol phosphate measurement and for microphysiology assessment of compound efficacy.

Radioligand Preparation

The radioiodinated agonist analog of GnRH, [des-Gly¹⁰,D-Ala⁶]GnRH ethylamide (¹²⁵I-GnRH-A) was used as the radioligand. One µg of GnRH-A diluted in 0.1M acetic acid was added to an Iodogen[®]-coated borosilicate glass tube (Pierce) containing 35 µl of 0.05 M phosphate buffer (pH 7.4-7.6) and 1 mCi of Na[¹²⁵I]. The reaction mixture was vortexed and incubated for 1 min at room temperature. After one min, the mixture was vortexed and allowed to incubate for an additional minute. 2 ml of 0.5 M acetic acid/1% BSA was added to the reaction tube and the mixture was added to a C18 Sep-Pak cartridge. The cartridge was washed with subsequent washes of 5 ml H₂O and 5 ml 0.5M acetic acid and then eluted with 5 x 1 ml of 60%CH₃CN/40% 0.5M acetic acid. The eluate was diluted with 3x volume of HPLC buffer A (0.1% TFA in H₂O) and loaded onto a C18 column. The iodinated product was eluted over 20-25 min with a gradient of 25-100% CH₃CN containing 0.1%TFA. The radioactive fractions (750 µl/fraction) are collected into clean polypropylene tubes containing 100 µl of 10% BSA. Fractions were assessed for biological activity by radioligand binding. Specific Activity of the radioligand was approximately 2200 Ci/mmol.

Microphysiometry.

The Cytosensor® Microphysiometer (Molecular Devices, Sunnyvale, CA) is a real-time, noninvasive, nonradioactive semiconductor-based system for monitoring the cellular responses to various stimuli. It is based on a pH-sensitive silicon sensor, the
5 light-addressable potentiometric sensor which forms part of a microvolume flow chamber in which cultured cells are immobilized (Pitchford et al., 1995; Parce et al., 1989; Owicki, et al. 1994).

Additional References:

Owicki, J.C., L.J. Bousse, D.G. Hafeman, G.L. Kirk, J.D. Olson, H.G. Wada, and J.
10 W. Parce. The light-addressable potentiometric sensor: principles and biological applications. *Ann. Rev. Biophys. Biomol. Struc.* 23:87-113, 1994.

Parce, J.W., J.C. Owicki, K.M. Kercso, G.B. Sigal, H.G. Wada, V.C. Muir, L.J. Bousse, K.L. Ross, B.I. Sikic, H.M. McConnell. Detection of cell-affecting
15 agents with a silicon biosensor. *Science* 246:243-247, 1989.

Pitchford, S., K. DeMoor, and B.S. Glaeser. Nerve growth factor stimulates rapid metabolic responses in PC12 cells. *Am. J. Physiol.* 268(*Cell Physiol.*37): C936-C943, 1995.

20

GGH₃ cells were seeded in low-buffered minimal essential media (MEM, Sigma) containing 25 mM NaCl and 0.1% BSA at a density of 500,000 cells/capsule onto the polycarbonate membrane (3 µm porosity) of cell capsule cups (Molecular Devices, Sunnyvale, CA). Capsule cups were transferred to sensor chambers where
25 cells were held in close apposition to a silicon sensor within a sensor chamber, which measures small changes in pH in the microvolume of the sensor chamber. Low-buffered medium was pumped continuously across the cells at a rate of approximately 100 µl/min from one of two fluid reservoirs. A selection valve determined which reservoir from which fluid was perfused onto the cells.

The Cytosensor[®] Microphysiometer generates a voltage signal, which is a linear function of pH, every second. In order to measure acidification rates, flow to the sensor chamber containing the cells was periodically interrupted, allowing for excreted acidic metabolites to build up in the extracellular fluid of the cells. In these experiments, cells were maintained at 37 °C on a two minute flow cycle with cells being perfused with media for 80 seconds followed by 40 seconds in which the flow of media was stopped. During this 40 second interval, acidification rates were measured for a 30 sec interval. In this fashion, a single acidification rate was calculated every two min. The Cytosensor[®] Microphysiometer device contains eight such sensor units, allowing for eight simultaneous experiments to be performed. Each unit was individually programmed utilizing a computer linked to the system.

GGH₃ cells were initially equilibrated in the low-buffered MEM media for a period of 30-60 min in which basal acidification rates (measured as $\mu\text{V}/\text{sec}$), in the absence of any stimuli, were monitored. When the basal rate of acidification changed by less than ten percent over a period of twenty minutes, experiments were initiated. Time course experiments were performed to determine the optimal time for agonist exposure prior to acidification rate measurement and the duration of exposure needed to obtain peak acidification responses to various agonists. From these time course experiments, it was determined that cells should be exposed to GnRH peptide agonists at least one minute prior to collection of acidification rate data. Peak acidification rates usually occurred in the first two-minute exposure cycle. In order to capture the peak response to GnRH, cells were exposed to the agonist for a total of four minutes. Cells were exposed to compounds for 20-60 min prior to a four-minute stimulation with GnRH (1.0 nM – 10 μM) alone or in combination with the various test concentration of each compound. All compounds were tested in a final concentration of 1% DMSO in low-buffered MEM media described above.

Selectivity Profile

In order to determine the specificity of binding of compounds to GnRH receptors, compounds were tested in various binding and functional assays for activity. Table 2 below shows the activity of Compound 20 in other assays.

Table 2. Binding Affinity or Functional Assessment of Compound 20 in Various Assays

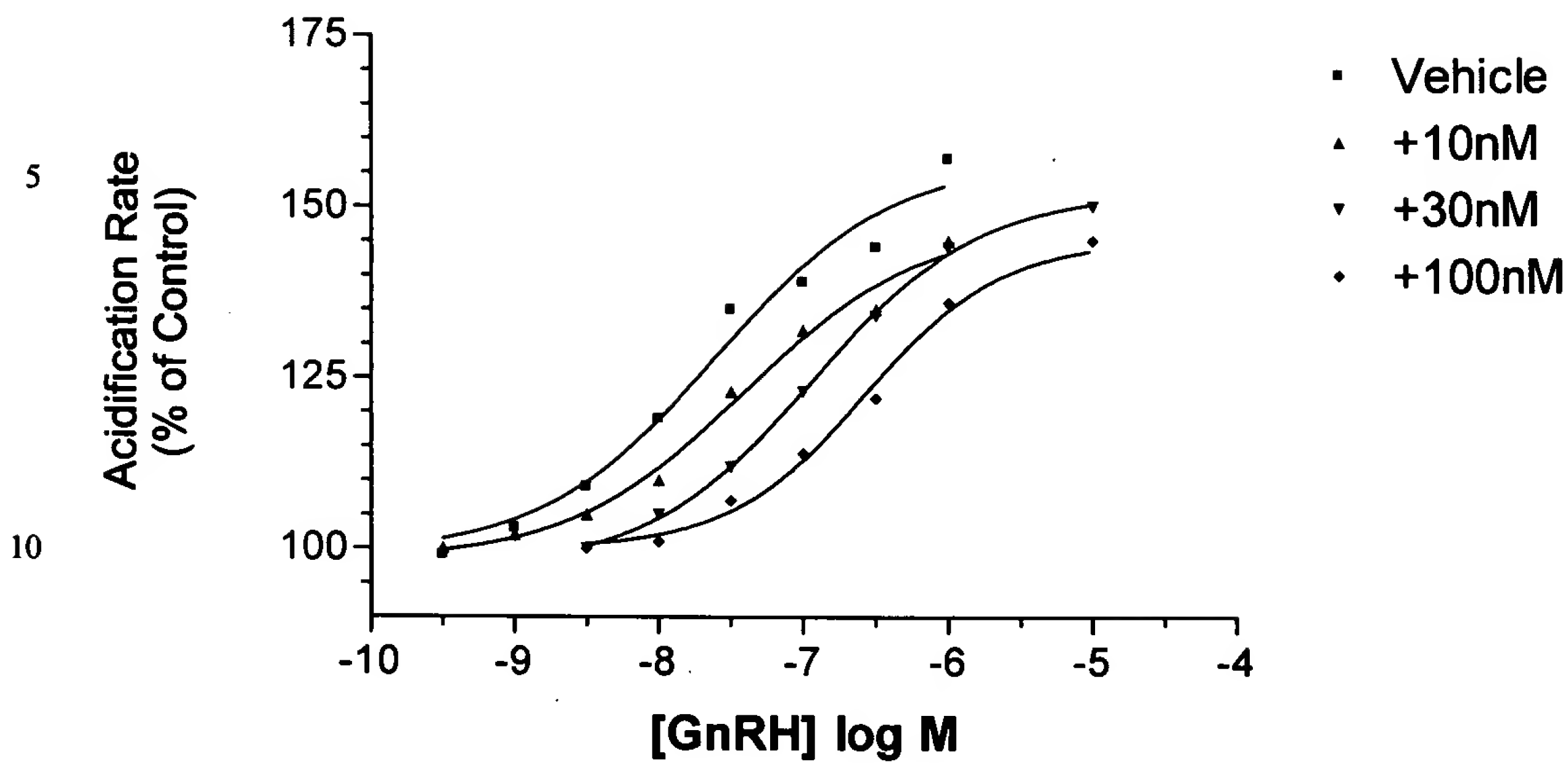
Assay	K _i (nM)*
Adenosine (non-selective)	>1000
Alpha1 adrenergic (non-selective)	>1000
Alpha2 adrenergic (non-selective)	>1000
Beta adrenergic (non-selective)	>1000
Dopamine (non-selective)	>1000
D2 Dopamine	>3200
H1 Histamine	>1000
H2 Histamine	≈1000
H3 Histamine	>1000
M2 Muscarinic	>1000
Muscarinic (non-selective) peripheral	>1000
Opiate (non-selective)	>1000
Serotonin Transporter	>1000
Serotonin (non-selective)	>1000
5-HT _{2a}	2350
5-HT ₇	>4400
Estrogen	>1000
Testosterone	>1000
L-type Ca ²⁺ Channel	>1000
N-type Ca ²⁺ Channel	>1000
ATP-sens. K ⁺ Channel	>1000
Ca ²⁺ -Activated K ⁺ Channel	>1000
Na ⁺ Channel (site 1)	>1000
Na ⁺ Channel (site 2)	>1000
LTB4	>1000
LTD4	>1000
TXA2	>1000
PAF	≈3000-5000
TRH	>1000
Oxytocin	>1000
AT1 Angiotensin	≈3000-5000
Bradykinin 2	>1000
CCKA	>1000
ET-A	>1000
Galanin	>1000
NK1	>1000
NK2	>1000
NK3	≈3000-5000
VIP (non-selective)	>1000
AchE	>1000
Choline Acetyltransferase	>1000
MAO-A	>1000
MAO-B	>1000
IL8RA (CXCR-1)	>10000
GLP-1	>10000
Glucagon	6700
NPY Y1	>10000
CYP3A4 Cytochrome P450 IC50	1700
Basal Histamine Release (rat mast cells) EC50	>10000
RAR γ Retinoid	>10000

RXR α Retinoid	>10000
Vasopressin1	>1000

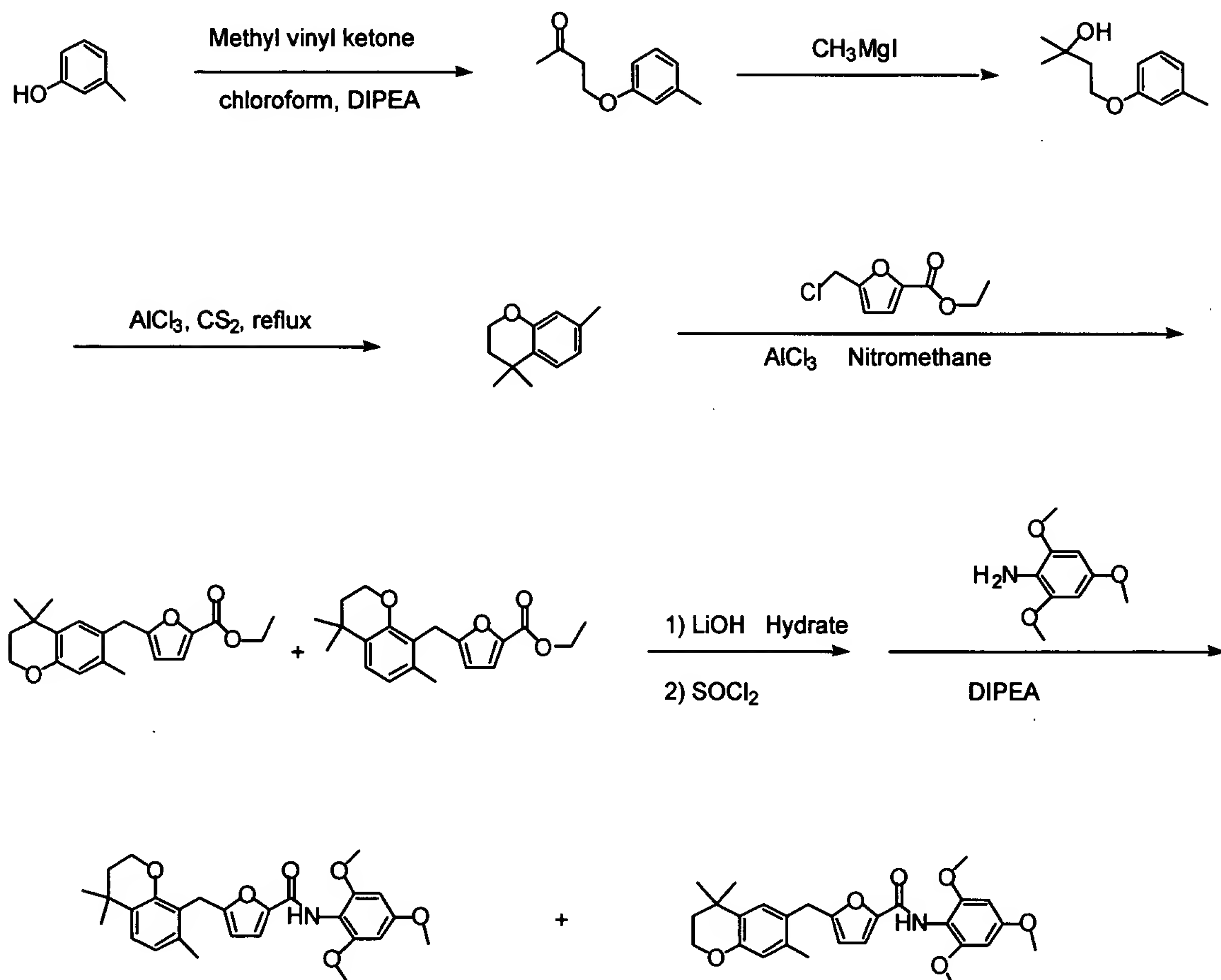
*EC₅₀ for Basal Histamine Release Assay

Figure 6 shows the effect of Compound 136 on GnRH-stimulated increases in extracellular acidification rate in GGH₃ cells. GnRH produced a dose-dependent increase in the extracellular acidification rate of GGH₃ cells. AXC06736 caused a rightward shift in the dose-response curves to GnRH without decreasing the maximum response to GnRH. This suggests that this compound is a competitive receptor antagonist of GnRH at this receptor. Values shown are from one experiment.

Figure 6.

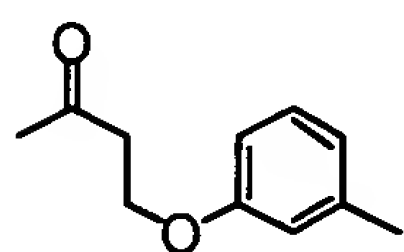


An example of preparation methods for compounds according to the invention is as follows:



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4-(3-methylphenoxy)-2-butanone:

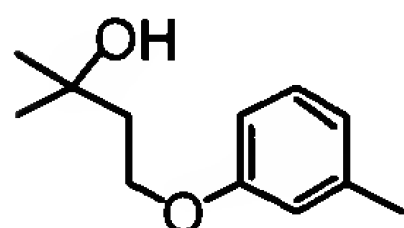


10

To m-Cresol (4.0 g, 37 mmol), methyl vinyl ketone (3.2 mL, 37 mmol) in chloroform (25 mL), added diisopropyl ethyl amine. The mixture was heated at reflux for 16 h, allowed to cool to room temperature and evaporated. The residue has 50% product and 50% starting material, starting material. The starting material was

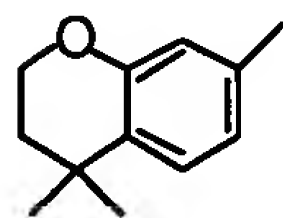
separated as t-butyldimethyl silyl ether. The product was isolated through plug filtration. Yield 4.5 g (68%).

2-Methyl-4(3-methylphenoxy)-2-butanol:



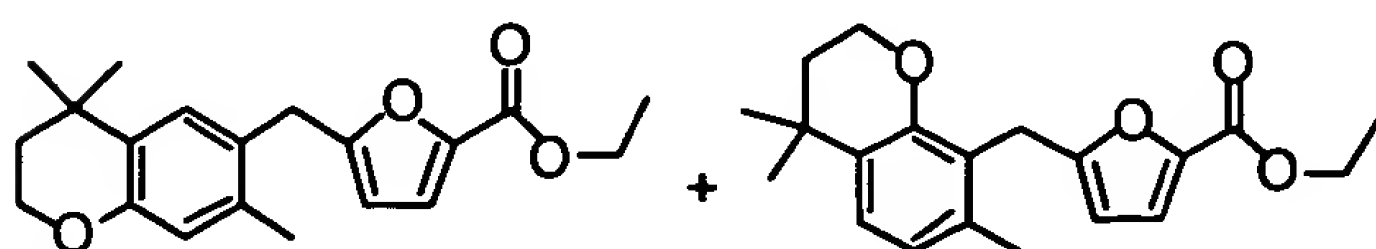
To a solution of methyl magnesium bromide in ether(50 mL), prepared from Mg (572 mg, 23.56 mmol) and MeI(3.34 g, 23.56 mmol), was added 4-(3-methylphenoxy)-2-butanone (2.1 g, 11.78 mmol) in 10 mL ether. The solution was stirred at room temperature for 30 minutes, after which quenched with water and dil. Hydrochloric acid. The organic layer was separated, dried over sodium sulfate, filtered through a silica plug. Colorless syrup 1.91 g (83%).

4,4,7-trimethyl chroman:



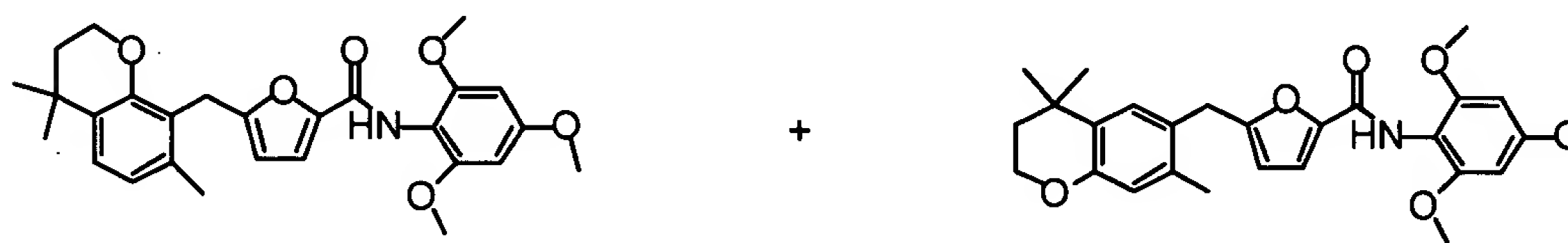
To aluminium chloride (1.3 g, 9.79 mmol) in 40 mL carbon disulfide, added 2-methyl-4(3-methylphenoxy)-2-butanol (1.9 g, 9.79 mmol) in 10 mL carbon disulfide. The mixture was heated at reflux for 2 h. Solvent evaporated, the residue diluted with 50 mL of ethyl acetate, and 10 mL of water. The organic layer separated, dried over sodium sulfate, and purified thorough a quick column. Light yellow syrup 1.5 g (87 %).

Ethyl-5-[(4,4,7-trimethyl-3,4-dihydro-2H-chromen-6-yl)methyl]-2-furoate, and ethyl-5-[(4,4,7-trimethyl-3,4-dihydro-2H-chromen-8-yl)methyl]-2-furoate:



To zinc chloride (950 mg, 6.97 mmol) in nitromethane (20 mL), was added a mixture of 4,4,7-trimethyl chroman (1.23 g, 6.97 mmol) and ethyl 5-chloromethyl-2-furoate (656 mg, 3.48 mmol) in nitromethane (15 mL). The mixture was stirred at room temperature for 16 h. Evaporated to dryness and triturated with ethyl acetate-water(1:1, 100 mL). The organic layer on usual work up, and plug filtration using hexane:ethyl acetate (9:1) gave mixture of these two compounds. 1.34 g (46% based on chroman).

N-(2,4,6-trimethoxyphenyl)-5-[(4,4,7-trimethyl-3,4-dihydro-2H-chromen-6-yl)methyl]-2-furamide and N-(2,4,6-trimethoxyphenyl)-5-[(4,4,7-trimethyl-3,4-dihydro-2H-chromen-8-yl)methyl]-2-furamide:



To a mixture of ethyl-5-[(4,4,7-trimethyl-3,4-dihydro-2H-chromen-6-yl)methyl]-2-furoate, and ethyl-5-[(4,4,7-trimethyl-3,4-dihydro-2H-chromen-8-yl)methyl]-2-furoate (1.34 g, 3.74 mmol) in THF-MeOH-H₂O (7:5:5, 20 mL) was added lithium hydroxide monohydrate (784 mg, 18.7 mmol). The mixture was stirred for 4 h at room temperature. The mixture evaporated to dryness, diluted with 30 mL ethyl acetate and 50 mL of water. After acidification with diluted HCl, ethyl acetate layer separated, dried and evaporated to give mixture of corresponding acids, 1.03 g (quantitative). The acids could not be separated using column chromatography or crystallization.

To the mixture of acids (200 mg, 0.66 mmol) in dichloromethane (30 mL) was added thionyl chloride (392 mg, 3.3 mmol). The mixture was refluxed for 1 h and evaporated. The residue was dissolved in hexane-ethyl acetate (9:1, 20 mL) and filtered through a silica gel plug (0.5 cm X 1.0 cm).

To the residue in 10 mL ethyl acetate, was added 2,4,6-trimethoxyphenyl amine hydrochloride (145 mg, 0.66 mmol) followed by diisopropyl ethyl amine (256 mg, 1.98 mmol). The mixture was stirred at room temperature for 16 h. The reaction was quenched with water (10 mL), ethyl acetate layer separated. The combination and column and HPLC purification gave 15 mg and 21 mg of two components (12 %).

Bioavailability of compounds of the invention are shown in the following Table.

In vivo pharmacology of some compounds of the invention was tested as follows:

In vivo experiments: general

Adult male Sprague-Dawley rats were purchased from Harlan Sprague Dawley (San Diego). Animals were housed two per cage and maintained in a temperature-controlled room (22±2°C) with a photoperiod of 12 hr light/ 12 hr dark (lights on at 0600h). Rat chow (Teklad rat diet) and tap water were provided *ad libitum*.

Animal models to access activity of GnRH antagonists:

Castrated Male Rate Model

Rationale:

Surgical removal of the gonads removes circulating testosterone and eliminates the negative feedback of testosterone on the hypothalamus. As a result GnRH is elevated and consequently elevates LH (Figure #7). A GnRH antagonist would be expected to reduce GnRH mediated elevations of LH levels. Antide, a GnRH peptide antagonist reduces LH levels in castrated rats (Fig. #8). This model seems suitable for evaluating small molecule GnRH antagonists.

Prot c 1:

Male Sprague-Dawley (200-225g) rats were castrated via the scrotal approach under halothane anesthesia. Animals were allowed 14 days post operative recovery prior to study. Thirteen days following castration, animals were anesthetized with halothane and instrumented with indwelling jugular vein cannula. Details of the cannulation procedure have been described previously {Harris and Ojeda, 1974}. On

study day, animals were allowed to acclimate to the procedure room while residing in their home cage. Basal blood samples were withdrawn from all animals. Immediately following basal sampling, vehicle or test compounds were administered by various routes. The routes of administration employed were intravenous (iv), intramuscular (im), intraperitoneal (ip), subcutaneous (sc) and oral (po). Blood samples were drawn into heparin containing tubes at multiple time points post treatment. Blood was centrifuged immediately, plasma collected and stored in -20° freezer until assayed. Plasma samples were analyzed using DSL-4600 ACTIVE LH coated-tube immunoradiometric assay kit from Diagnostic Systems Laboratories, Inc.

Compound Formulations:

Formulation #1 (denoted with superscript 1): 10% DMSO, 10% Cremophor EL and 80% physiological saline.

Formulation #2 (denoted with superscript 2): 10% Cremophor EL and 90% physiological saline.

Results

See Figures 9–11 and Table.

Intact Male Rat

Rationale:

Testosterone is a hormone regulated by the hypothalamic-pituitary-gonadal axis. GnRH is secreted in pulses from the hypothalamus and stimulates the anterior pituitary gland to release gonadotropic hormones LH and FSH. Testosterone is produced when the testes are stimulated by LH. The quantity of testosterone secreted increases approximately in direct proportion to the amount of LH available (Guyton, 1986). A GnRH antagonist is expected to reduce testosterone level by inhibiting LH.

Protocol 1:

Male Sprague-Dawley (250-275g) rats were single-housed and allowed to acclimate for 1 week prior to study. On study day animals were dosed with vehicle or test compound by various routes of administration, including ip, sc, or po. Blood samples were obtained via cardiac puncture under halothane anesthesia from individual

animals at predetermined time points post treatment. Blood samples were drawn into heparin containing tubes. Blood was centrifuged immediately, plasma collected and stored in -20° freezer until assayed. Plasma samples were analyzed using DSL-4000 ACTIVE Testosterone coated-tube radioimmunoassay kit from Diagnostic Systems Laboratories, Inc.

Protocol 2:

Male Sprague-Dawley (250-275g) rats were single-housed and allowed to acclimate for 1 week prior to study. We developed a technique to allow for repeated sampling from the jugular vein by using microrenathane (MRE) catheters implanted 7 days prior to study. Details of the surgical procedure have been described previously {Harris and Ojeda, 1974}. On study day, animals were allowed to acclimate to the procedure room while residing in their home cage. Basal blood samples were drawn from all animals. Immediately following basal sampling, vehicle or test compounds were administered by various routes. The routes of administration employed were intravenous (iv), intramuscular (im), intraperitoneal (ip), subcutaneous (sc) and oral (po). Blood samples were drawn into heparin containing tubes at multiple time points post treatment. Blood was centrifuged immediately, plasma collected and stored in -20° freezer until assayed. Plasma samples were analyzed using DSL-4000 ACTIVE Testosterone coated-tube radioimmunoassay kit from Diagnostic Systems Laboratories, Inc.

Protocol 3: Repeated Dosing Study Compound No. 134

Male Sprague-Dawley (250-275g) rats were double-housed and allowed to acclimate for 1 week prior to study. Daily vehicle treatments were administered either, im, sc, or po between 8:00 and 9:00 am for seven days. On day 8 between 8:00 and 9:00 am, blood samples were drawn via cardiac puncture under halothane anesthesia. The procedure was completed in 45–60 seconds. Over the next 7 days one group of animals continued to receive vehicle treatments while another group of animals

received no treatment. Samples were collected as described above following the 7th day of this treatment regimen. Testosterone levels were not different between vehicle treated animals and untreated animals.

Daily im dosing of Compound 134 (100 mg/kg) or vehicle was performed
5 between 8:00 and 9:00 am for seven days. Samples were collected as described above following the 7th day of treatment.

All blood samples were collected in heparin containing tubes. Blood was centrifuged immediately, plasma collected and stored in -20° freezer until assayed the next day. Plasma samples were analyzed using DSL-4000 ACTIVE Testosterone
10 coated-tube radioimmunoassay kit from Diagnostic Systems Laboratories, Inc.

Results

See Table 2 and Figures 12–14.

Representative Figures

Figure # 7

**Characterization of a Castrated Rat Model
used to Evaluate GnRH Antagonists**

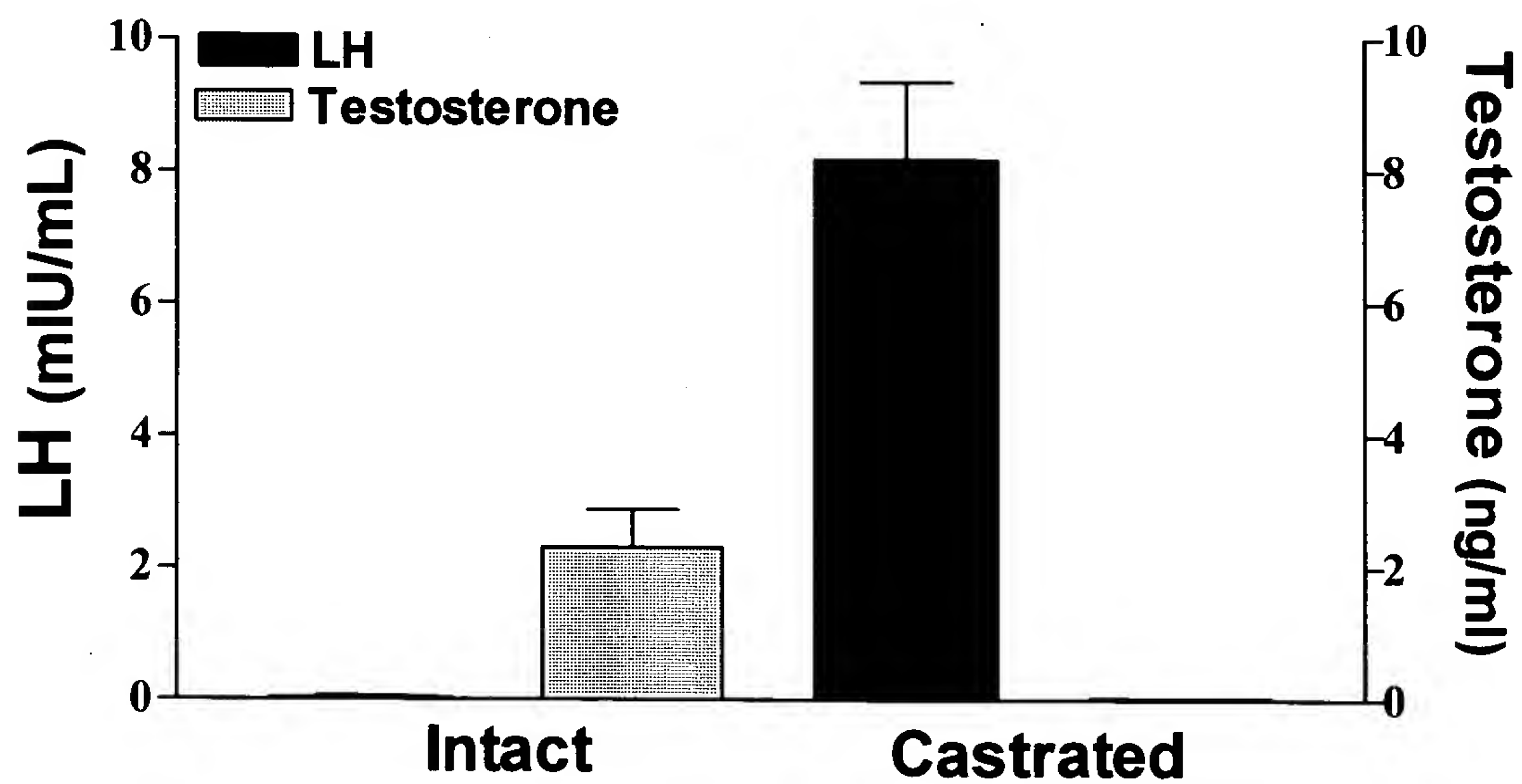


Figure # 8

Antide suppression of LH in castrated rats is dose-dependent

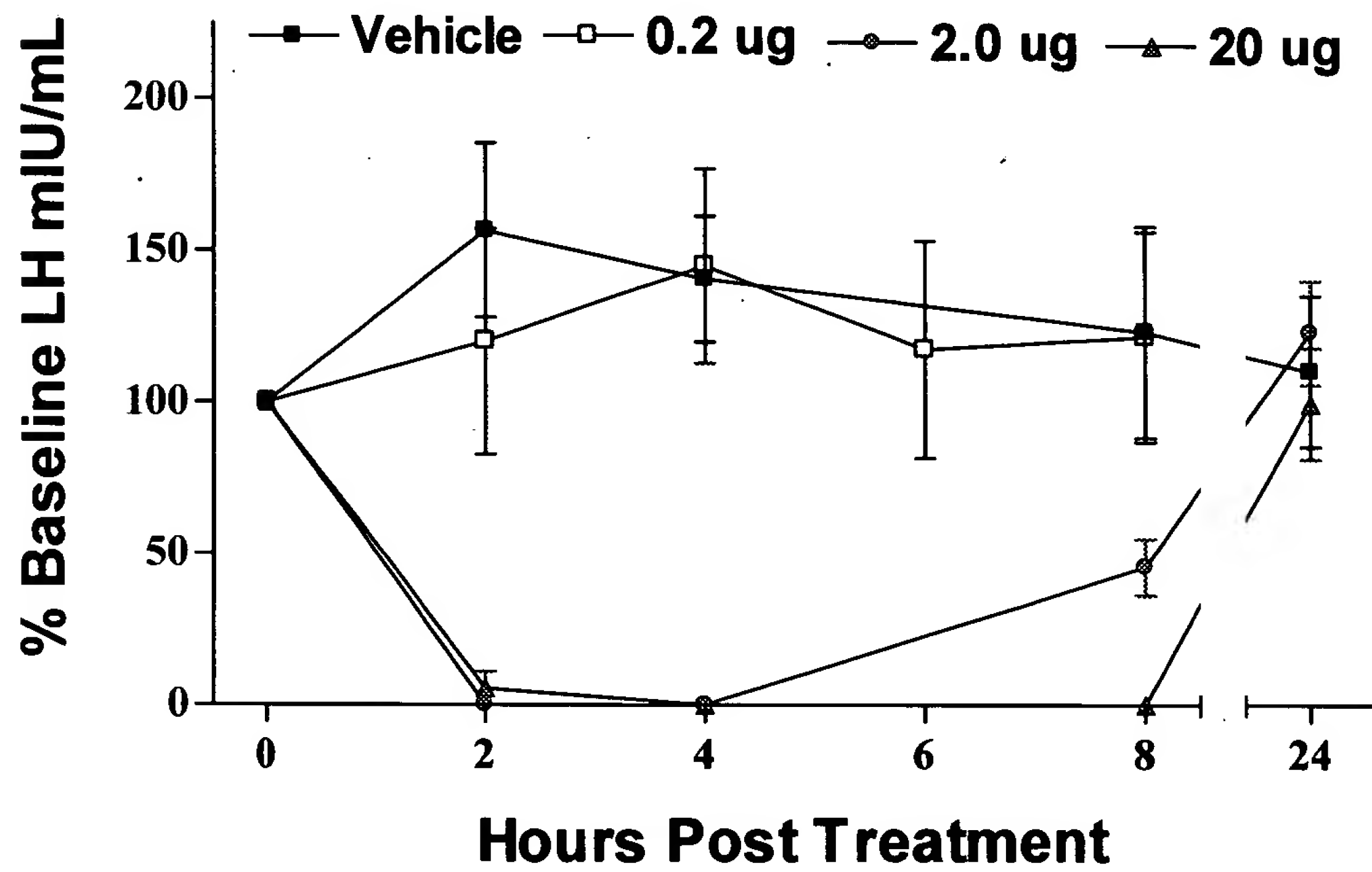


Figure # 9

No. 134 (iv) suppression of LH is dose-dependent

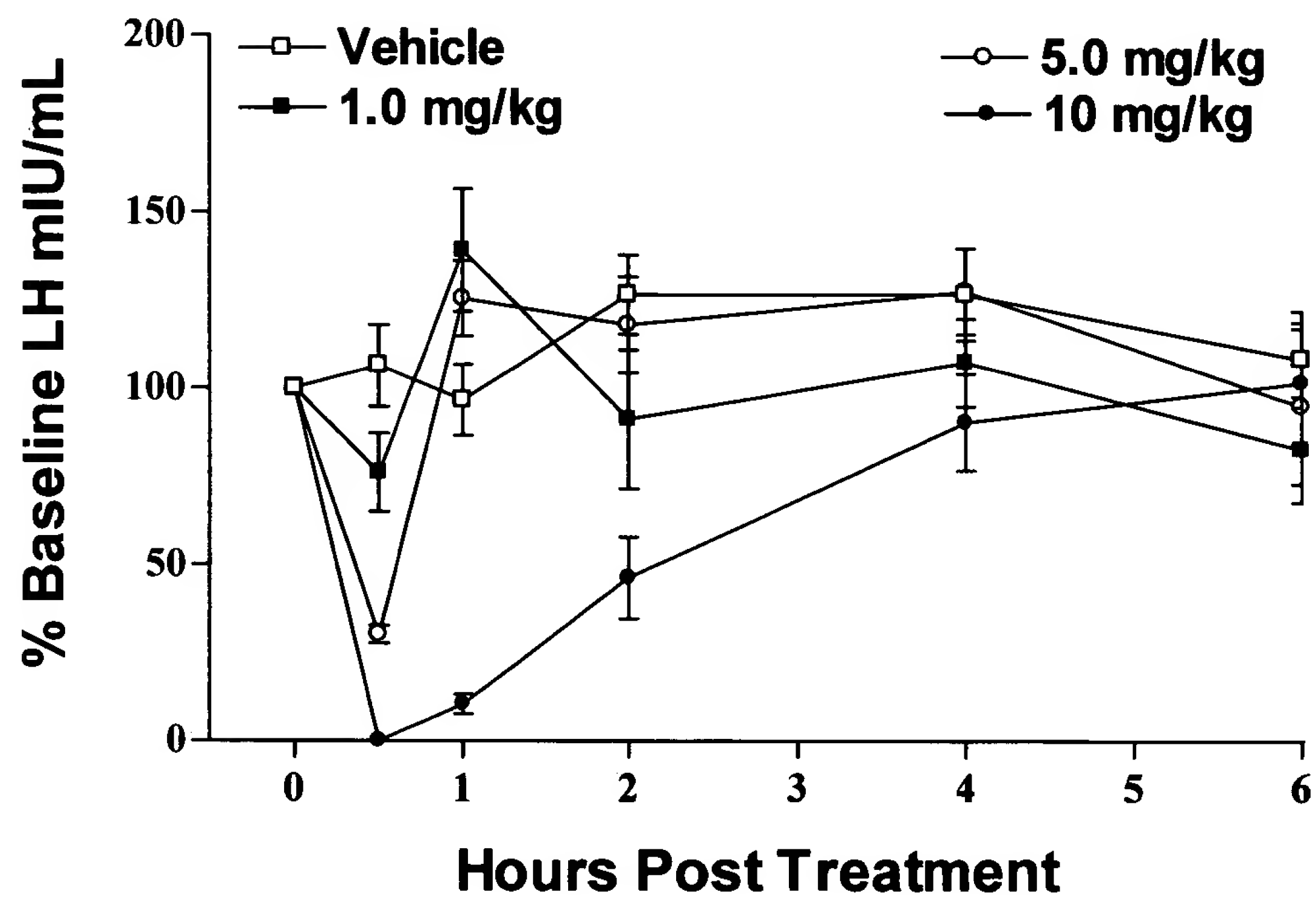


Figure # 10

**Oral delivery of No. 134
suppresses LH in castrated rats**

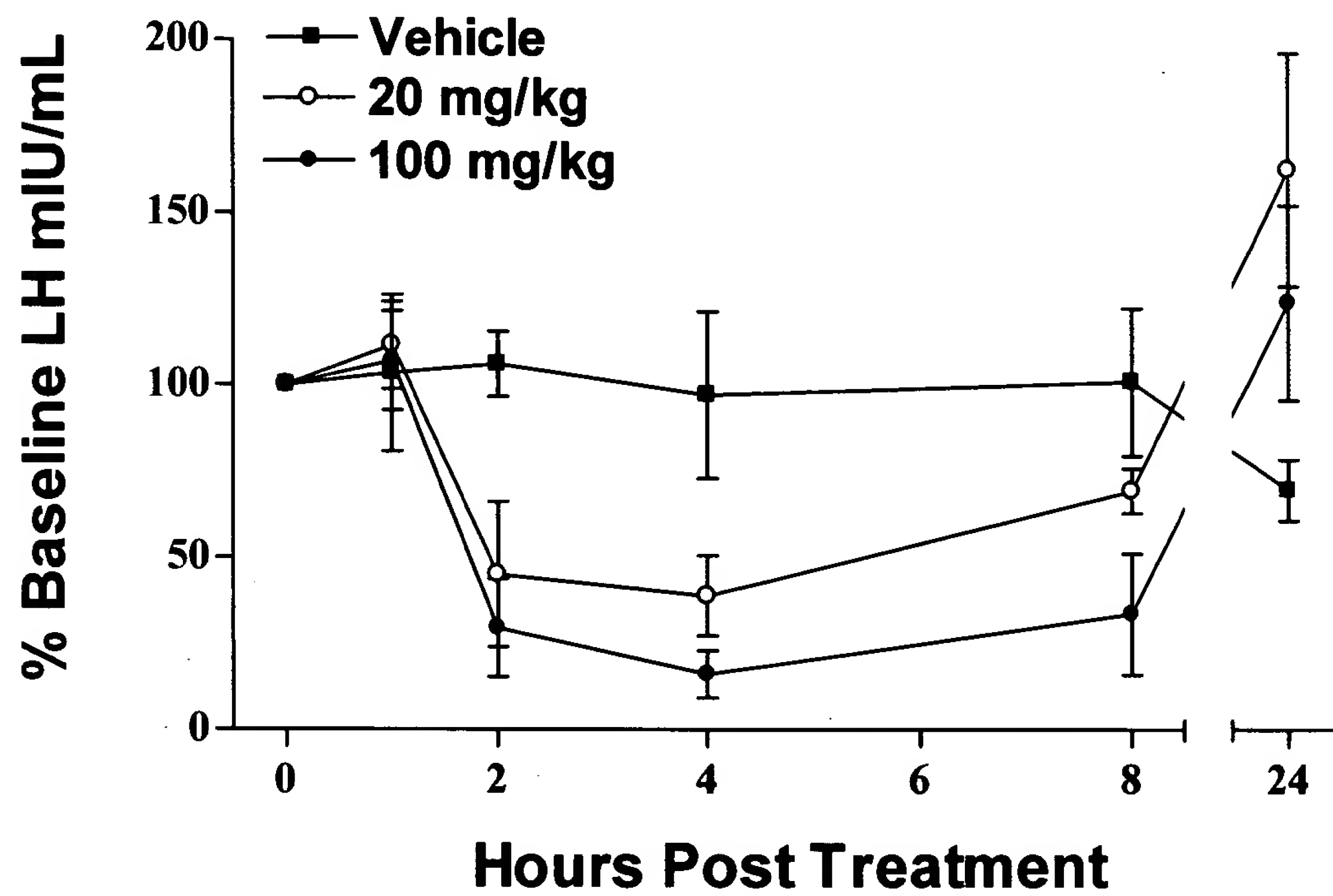


Figure # 11

**IM Delivery of No. 134 (20mg/kg)
suppresses LH in CX rats > 12 hrs.**

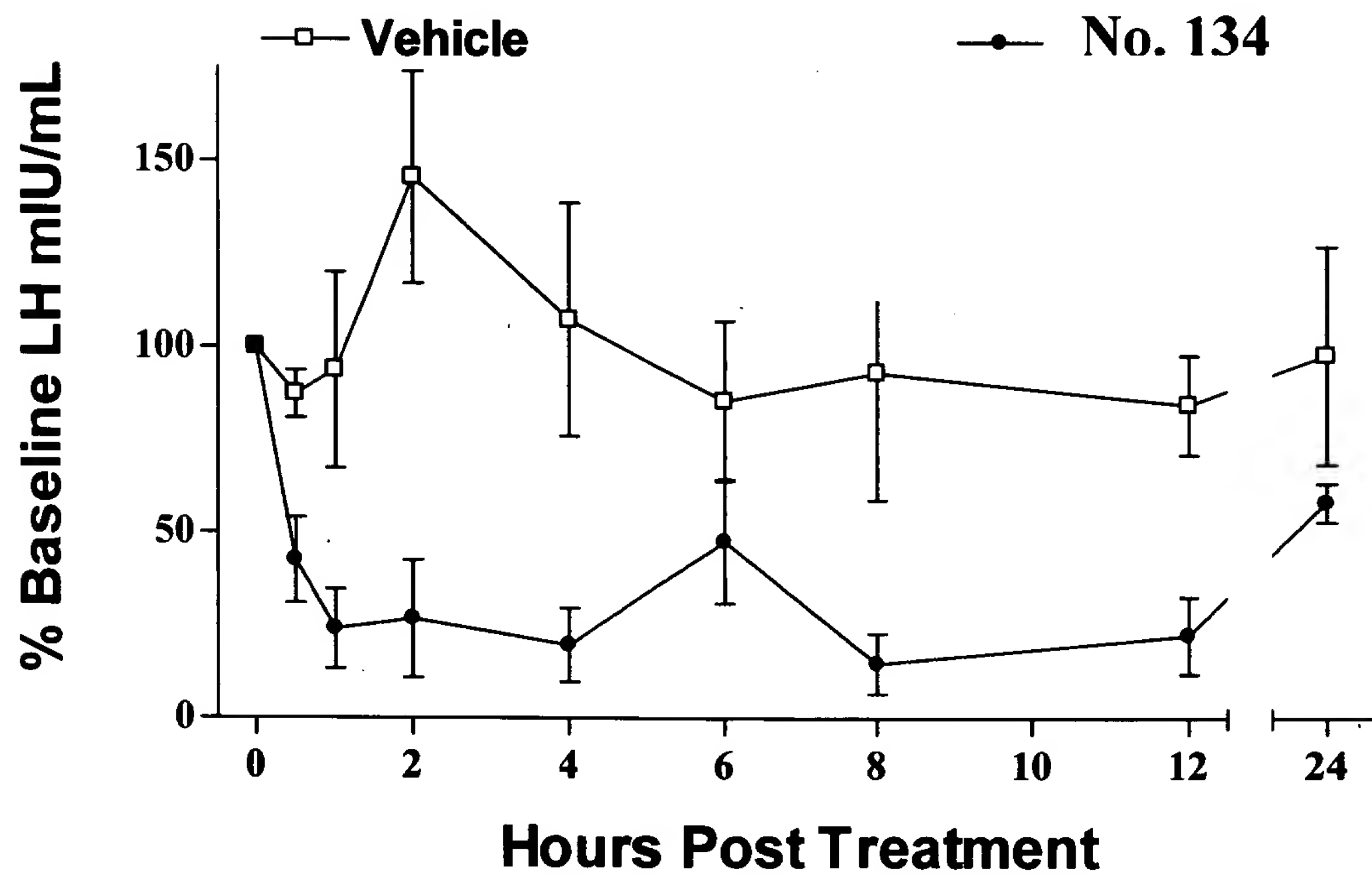


Figure Legends

Figure 7: Bar graph shows basal LH and testosterone levels in castrated (CX) and intact rats. LH is elevated in CX rats. Testosterone is absent in CX rats.

5 Figure 8: Line plot shows LH levels expressed as a percentage of basal LH in vehicle and Antide treated animals. Antide (2.0 and 20 ug;sc) suppresses LH in CX rats.

Figure 9: Line plot shows LH levels expressed as a percentage of basal LH in vehicle and compound treated animals. Compound 134 (1.0, 5.0 & 10 mg/kg;iv) produces
10 dose-dependent suppression of LH in CX rats.

Figure 10: Line plot shows LH levels expressed as a percentage of basal LH in vehicle and compound treated animals. Compound 134 (20 or 100 mg/kg;ip) suppresses LH in CX rats.

15

Figure 11: Line plot shows LH levels expressed as a percentage of basal LH in vehicle and compound treated animals. Compound 134 (20 mg/kg;im) suppresses LH in CX rats.

Table 1 GnRH Compounds in Castrated Rat Model

Compound	Dose mg/kg Route	<u>% Max LH Suppression</u>	<u>≥ 50 % Suppression Duration</u>	<u>Range of Plasma Conc.</u> μM
No. 9	1.0/ iv	40 @ 0.5 hr	NS	ND
No. 11	10/iv 10/ip 20/po	85 @ 0.75 hr 50 @ 0.5 hr NS	3 hr < 1 hr NS	ND
No. 13	20/iv	63 @ 0.5 hr	< 1 hr	ND
No. 48	10/iv	75 @ 0.5 hr	< 1 hr	

	20/iv	51 @ 1 hr	1 hr	35.7-3.49
No. 20	10/iv 20/iv 50/im 50/po 100/po	78 @ 0.5 hr 72 @ 1 hr 65 @ 1 hr 59 @ 1 hr 43 @ 2 hr	0.5 hr 1.0 hr 2 hr 2 hr NS	ND
No. 136	5.0/iv 10/iv 20/iv 20/ip 20/po 40/po	48 @ 1 hr 74 @ 2 hr 98 @ 4 hr NS NS	1.0 hr 6 hr ≥ 6 hr NS NS	3.0-0.3 3.7-0.28 8.9-0.7 0.29 0.72
No. 134	1.0/iv 5.0/iv 10/iv 20/po 100/po 20/ip 100/ip 20/sc 100/sc 20/im 100/im	24 @ 0.5 hr 70 @ 0.5 hr 100 @ 0.5 hr 62 @ 4 hr 84 @ 4 hr 80 @ 2 hr 98 @ 4 hr 53 @ 8 hr 80 @ 6 hr 73 @ 2 hr 98 @ 2-24 hr	 2 hr 4 hr 8 hr 4 hr 8 hr only @ 8 hr 8 hr 8 hr 24 hr	0.53-0.20 3.2-0.11 4.8-0.16 0.94-0.3 1.27-0.7 1.7-0.45 1.7-0.32 0.6-0.3 0.39-.15 1.3-0.12 10.8-0.5
No. 119	5.0/iv 10/iv 20/po 20/ip 20/sc	63 @ 1 hr 61 @ 1 hr 63 @ 2hr 79 @ 2 hr NS	1 hr 1 hr 2 hr 2 hr NS	3.1- 0.8 8.13-0.35 0.1 0.6-0.1 0.2
No. 183	10/iv 50/po 10/im	92 @ 1 hr 60 @ 2 hr 58 @ 2 hr	8 hr 8 hr NS	1.5-0.89 0.65-.13 0.18-.08
<u>No. 206</u>	10/iv 20/po	51 @ 1hr NS	NS NS	0.2-0.06 0.1-0.05

NS=No suppression
ND=Not determined

Representative Figures

Figure # 12

No. 136 (20 mg/kg;IP) suppresses testosterone 6 hours post treatment

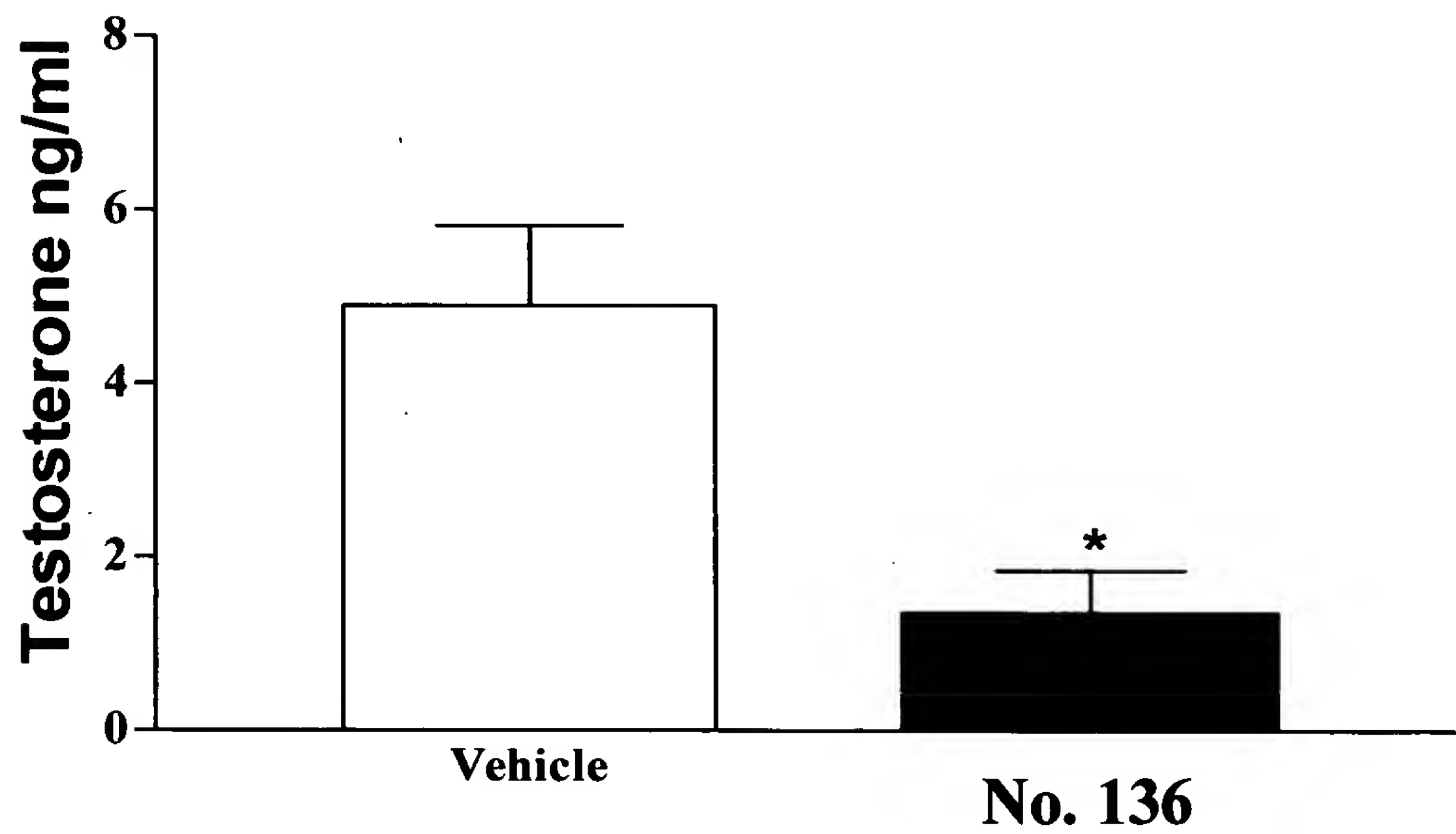


Figure # 13

Effect of oral delivery of No. 134 on testosterone in intact rats

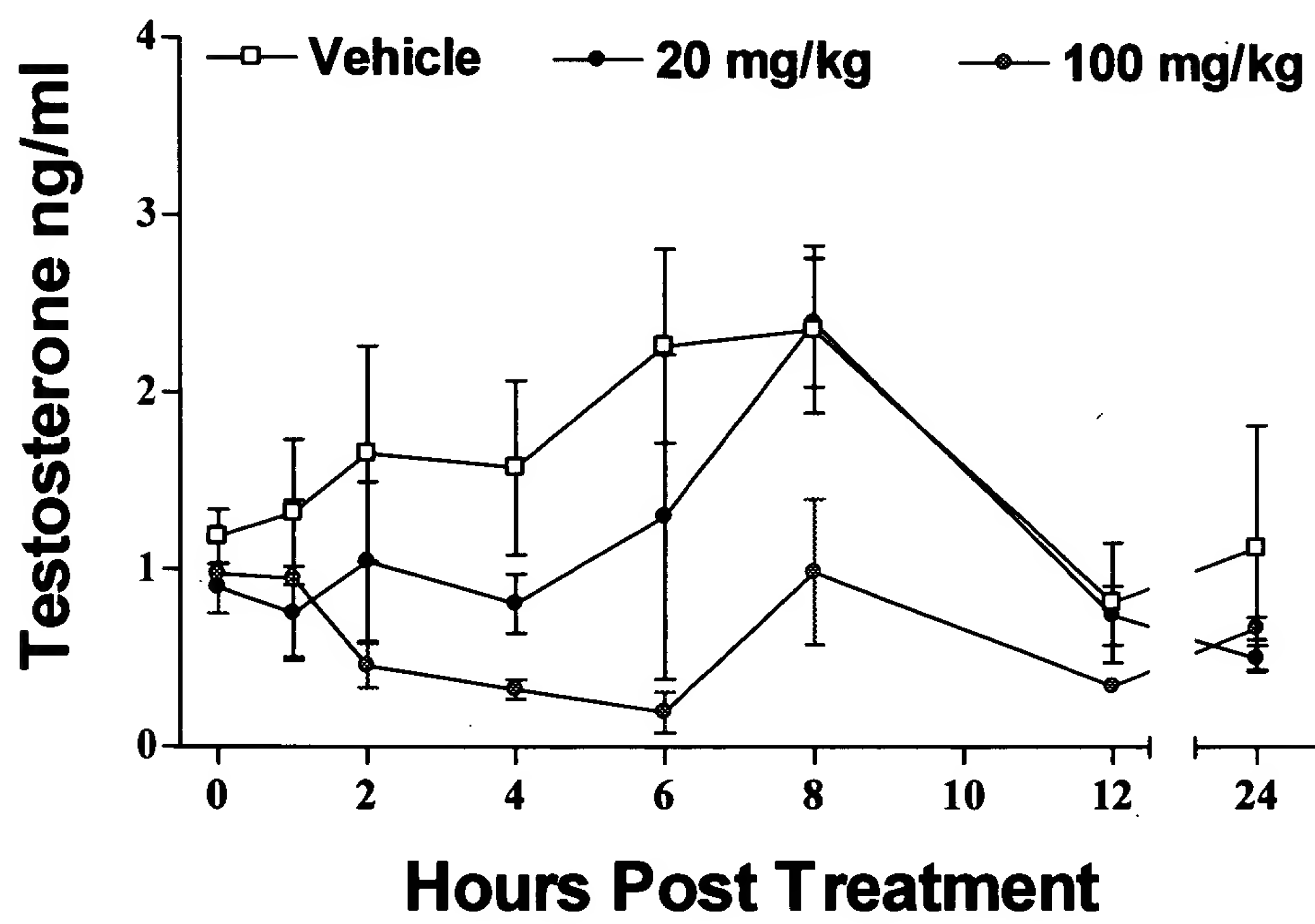


Figure # 14

**Daily dosing of No. 134 (100 mg/kg;im)
suppresses testosterone in rats**

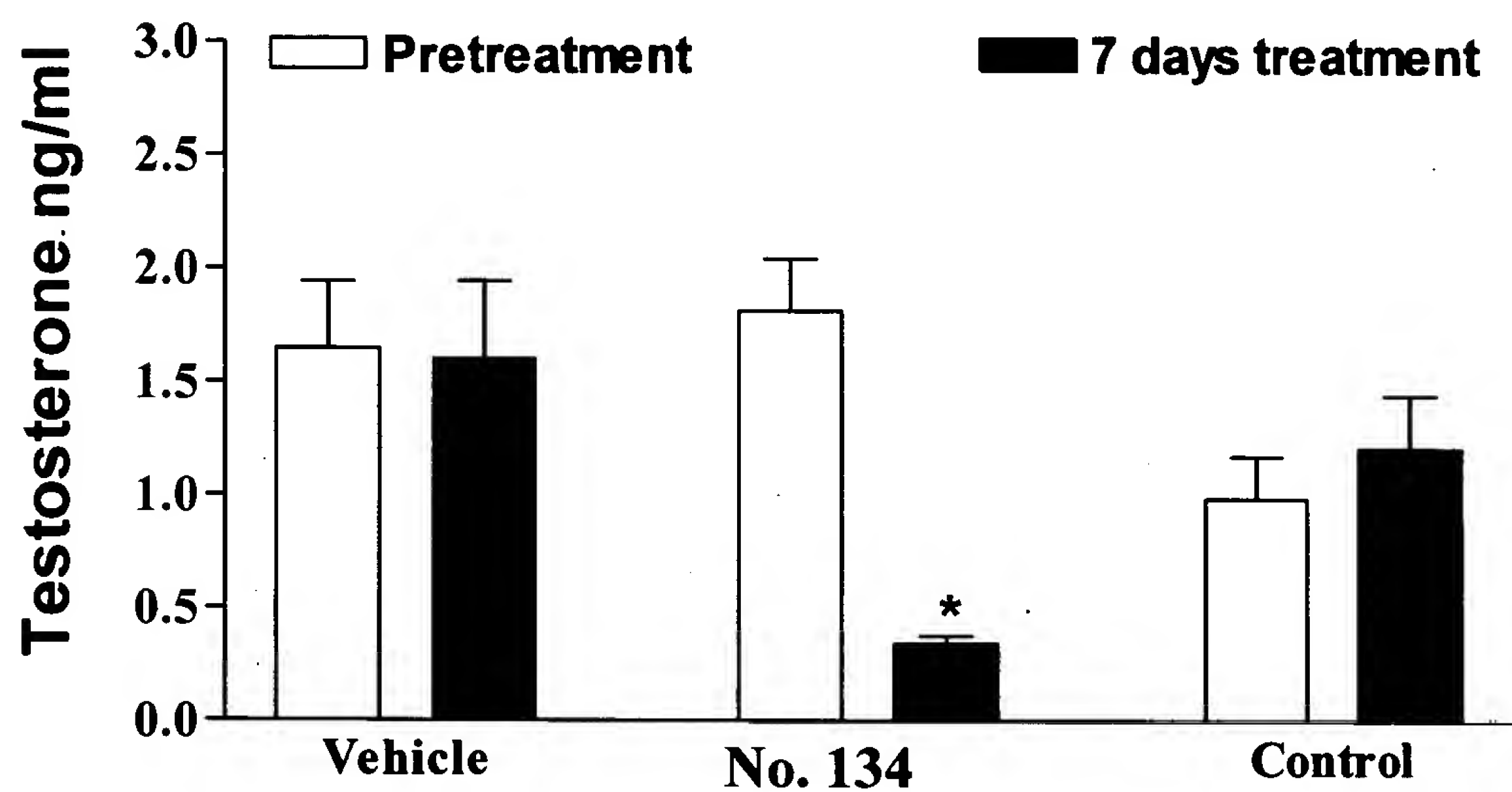


Figure Legends

Representation of Protocol 1:

Figure 12: Bar graph shows testosterone levels in vehicle and compound treated animals 6 hours post ip injection. Compound 136 suppressed testosterone levels compared to vehicle treated animals * =p<0.05, t-test.

Representative of Protocol 2:

Figure 13: Line plot shows testosterone levels over a 12 hour time course and 24 hour time point in vehicle and compound 134 treated rats. Vehicle and compound 134 was delivered by oral gavage. The highest dose of compound 134 suppressed testosterone throughout the course of the study.

Representative of Protocol 3:

Figure 14: Bar graph shows testosterone levels for vehicle, compound 134 and control treated animals. Open bars represent pretreatment testosterone levels and solid bars represent testosterone levels following 7 days of repeated treatment. Compound 134 significantly suppressed testosterone levels compared to pretreatment, vehicle and control-treated animals. * =p<0.05., t-test.

Table 2 GnRH Compounds in Intact Male Rat

<u>Compound</u>	<u>Dose mg/kg</u> <u>Route</u>	<u>% Testosterone</u> <u>suppression</u>	<u>Suppression</u> <u>Duration</u>	<u>Range of Plasma</u> <u>Conc.</u> <u>μM</u>
No. 134	20/ip	36 @ 4 hr	4 hr	0.2
	100/sc	60 @ 24 hr	24 hr	0.13
	20/po	NS @ 2 or 4 hr	NS	0.15 @ 2hr 0.17 @ 4 hr
	100/im * 7days	80 @ 12 hr	12 hr	1.3
	20/po	NS	NS	1.1 @ 1 hr
	50/po	75 @ 4 hr	8 hr	2.5-0.22

	100/po	80 @ 6 hr	12 hr	2.4-0.25
No. 136	20/ip	72 @ 6 hr	6 hr	0.4
	40/po	46 @ 4 hr	4 hr	0.34

NS=No suppression

ND=Not determined

5 **Procedural notes**

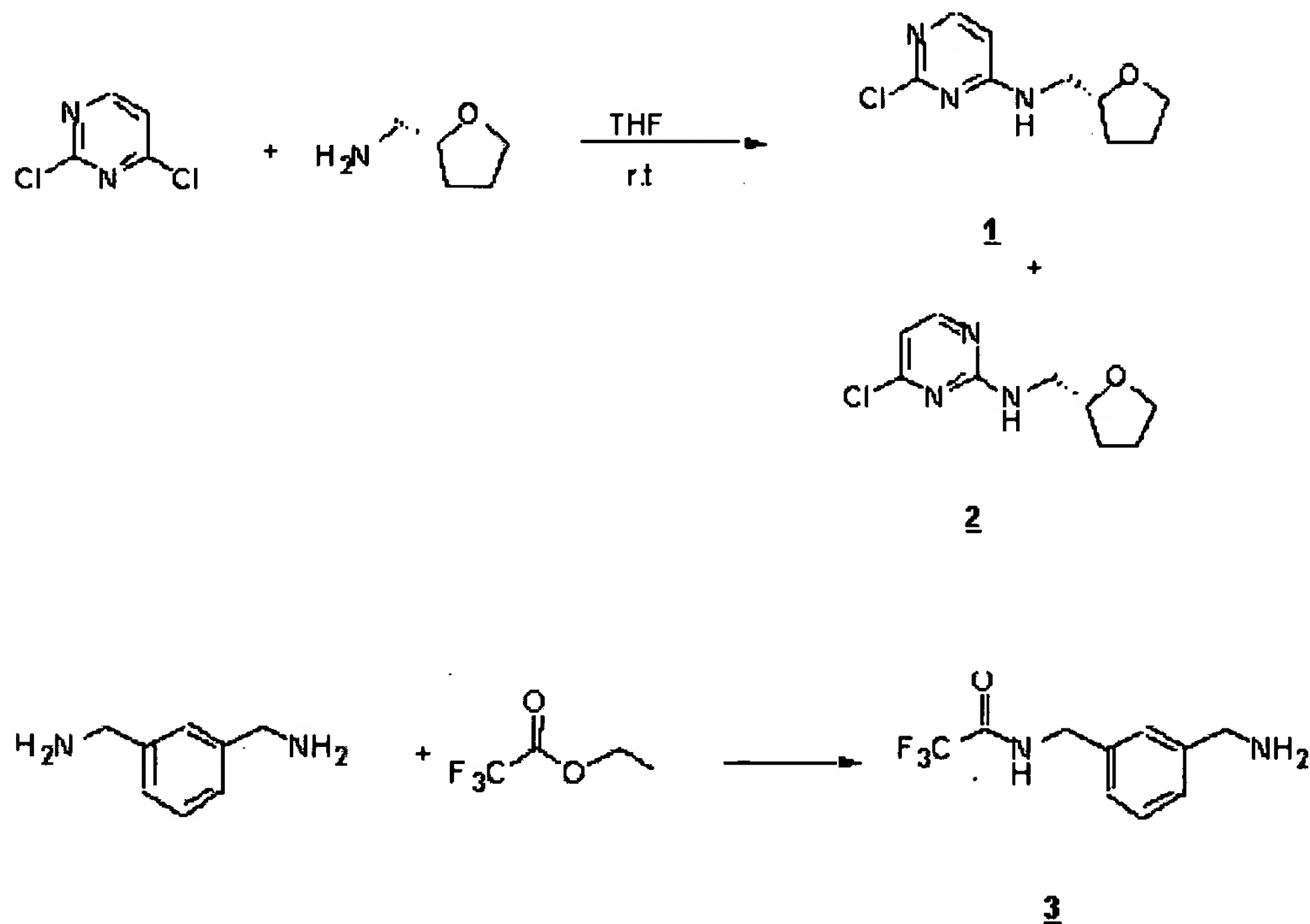
It has been documented that some of the procedures commonly used in endocrine studies on animals, such as anesthesia, fasting, surgery may affect the hormone levels being studied (B.E. Howland, et al., Experientia, 1974.) Luteinizing hormone and testosterone are sensitive to stressors. Numerous reports are conflicting about the effects of stressors on the HPG axis even when the same species and stressors are utilized. For example, male rats that are subjected to restraint or immobilization have been reported to have **low**, (Kruhlich et al., 1974; DuRuisseau et al., 1978), **normal** (Tache et al., 1980; Charpenet et al., 1982; Collu et al., 1984), or **elevated** LH concentrations (Briski et al, 1984). Similarly, plasma testosterone levels have been reported to change following exposure to stressful situations, but again the data appear contradictory and therefore difficult to interpret. For example, during intense physical exercise plasma testosterone levels have been reported to **increase** (Dessypris et al., 1976), **decrease** (Sutton et al., 1973) or remain **unchanged** (Lamb, 1975). The effects of immobilization on testosterone concentrations have been more consistent with most investigations reporting a decline in circulating values (Tache et al., 1980; Charpenet et al., 1982; Collu et al., 1984). It is, however, accepted that stressors do elicit changes in circulating testosterone and the type of stress used, duration and severity cause different stress-induced changes in testosterone concentrations. Considering the susceptibility of LH and testosterone to stress, we have

optimized protocols to evaluate LH and testosterone under conditions which minimize stress.

Compounds according to the invention that have been prepared are shown in the attached Tables.

- 5 Compounds can be prepared with the general experimentals provided above. Specific examples are given below.

Pyrimidine-Containing Compounds



10

Preparation of 2-chloro-N-[(2R)-tetrahydro-2-furanmethyl]-4-pyrimidinamine 1 and 4-chloro-N-[(2R)-tetrahydro-2-furanmethyl]-2-pyrimidinamine 2:

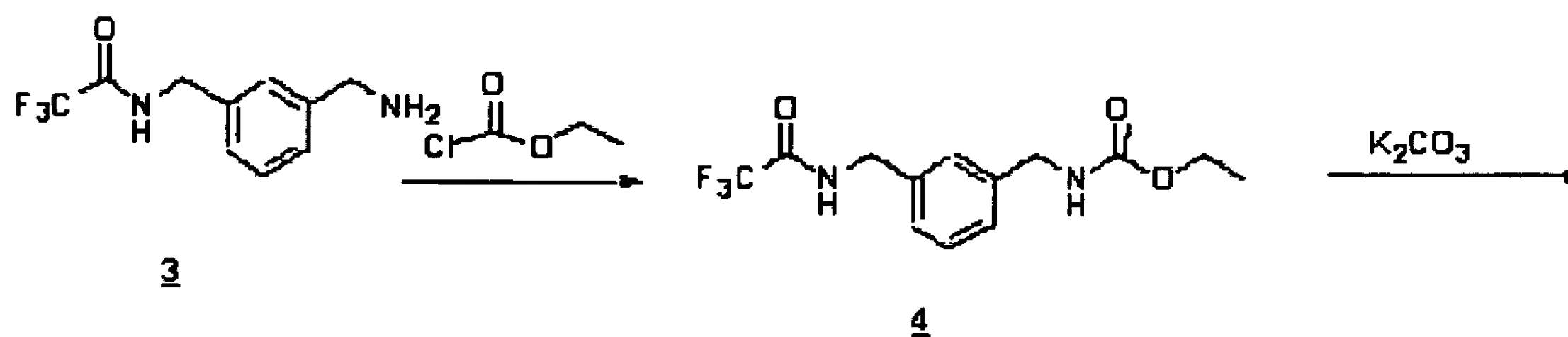
- 15 To a 250 mL round bottom flask was placed 2,4-dichloropyrimidine (5.0g, 33.56 mmol) and 200 mL THF. To this solution was added triethylamine (14.0 mL, 100.68 mmol) and [R]-tetrahydrofurfurylamine. The solution was stirred overnight. The reaction mixture was poured into water and extracted with methylene chloride. The separated organic layer was washed with brine, dried over magnesium sulfate, and

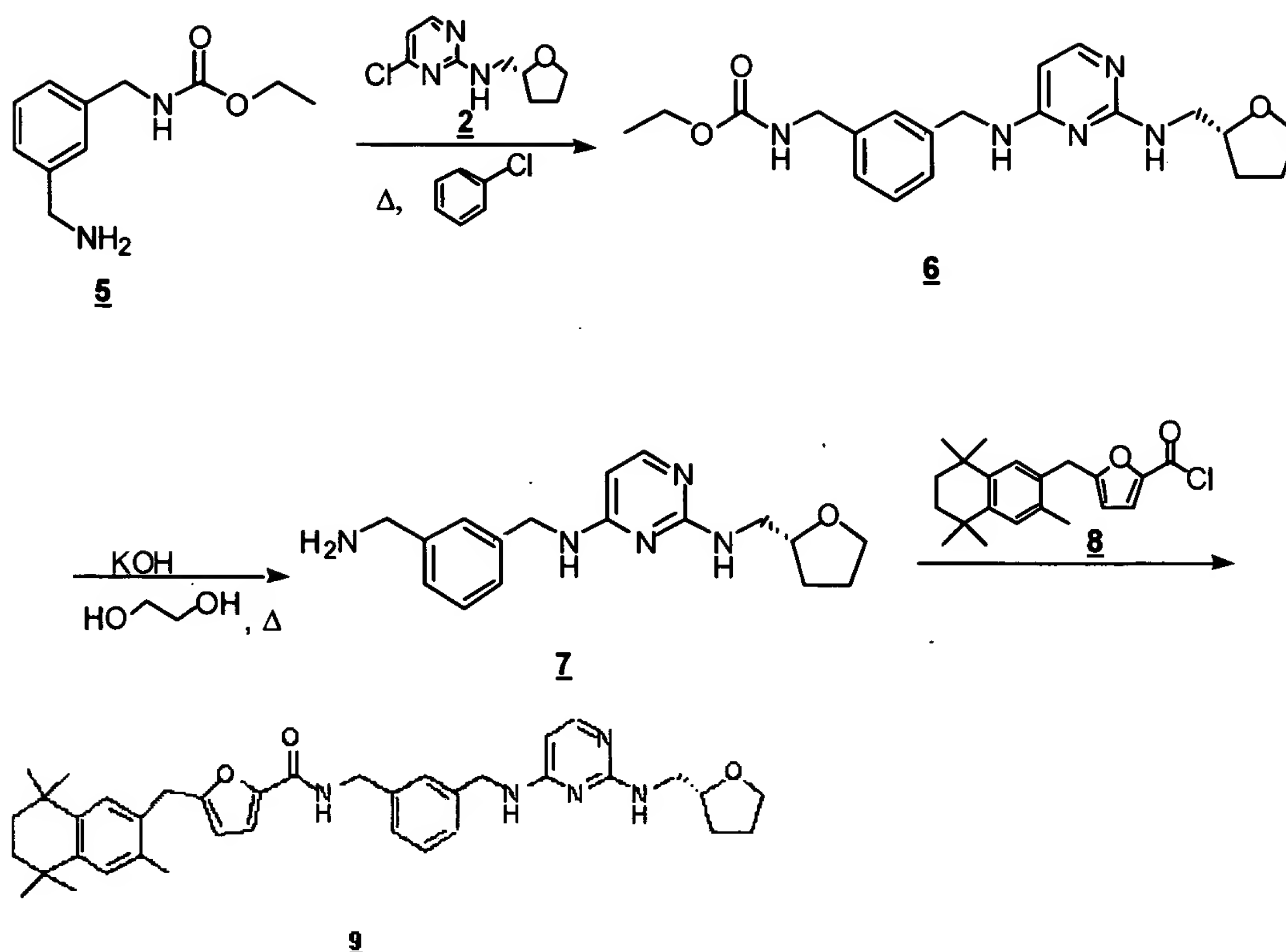
concentrated on a rotary evaporator. The crude compound was purified by silica gel chromatography with hexane/ ethyl acetate (4:1 v/v to 1:1 v/v) to yield 2 (1.3 g) and 1 (3.98 g).

5 **Preparation of N-[3-(aminomethyl)benzyl]-2,2,2-trifluoroacetamide 3.**

To a solution of m-xylene diamine (28.76g, 211.15 mmol) in THF (300 mL, .7M) was added dropwise a solution of ethyl trifluoroacetate (10g, 70.38 mmol) in THF (50 mL, 1.4M). The solution was stirred at room temperature overnight. The reaction was monitored by TLC. The solvent was concentrated and residue was acidified to pH 2 with 4N HCl and dissolved in water and washed with ethyl acetate. The separated aqueous layer was basified to pH 11 using NH₄OH and compound was extracted with dichloromethane. The separated organic layer was wash with water/brine, dried over magnesium sulfate and concentrated to yield 3 (8.71g, 53% yield).

15 **Synthesis of 5-[(3,5,5,8,8-pentamethyl-5,6,7,8-tetrahydro-2-naphthalenyl)methyl]-N-(3-[[2-[[2-[(2R)-tetrahydro-2-furanylmethyl]amino]-4-pyrimidinyl)amino]methyl]benzyl)-2-furamide 9.**





5 Preparation of ethyl 3-(aminomethyl)benzylcarbamate 5.

To a solution of N-[3-(aminomethyl)benzyl]-2,2,2-trifluoroacetamide 3 (10.6g, 43.1 mmol) was added ethyl chloroformate (1eq.) followed by triethylamine.. Reaction was stirred at room temperature for 30 min. Crude product was extracted with methylene chloride and concentrated to give ethyl 3-[[[(trifluoroacetyl)amino]methyl} benzylcarbamate 4. This crude product was dissolved in methanol (100 mL) and 2N K₂CO₃ (100 mL) and stirred overnight. Reaction mixture was basified to pH 14 with 20% NaOH, extracted with methylene chloride, wash with brine and dried over magnesium sulfate to yield 5 (5.2g)

15 Preparation of ethyl 3-[[[(2-[[[(2R)-tetrahydro-2H-pyran-2-yl]amino]-4-pyrimidinyl)amino]methyl}benzylcarbamate 6

To a solution of ethyl 3-(aminomethyl)benzylcarbamate 5 yield 4-chloro-N-[(2R)-tetrahydro-2-furanylmethyl]-2-pyrimidinamine 2 in chlorobenzene was added triethylamine. Reaction mixture was reflux overnight. The solution was cooled to room temperature and loaded on a silica gel column and eluted with hexane/ethyl acetate (1:1 v/v) to yield ethyl 3-[(2-[(2R)-tetrahydro-2-furanylmethyl]amino)-4-pyrimidinayl]amino]methyl}benzylcarbamate 6 (73% yield).

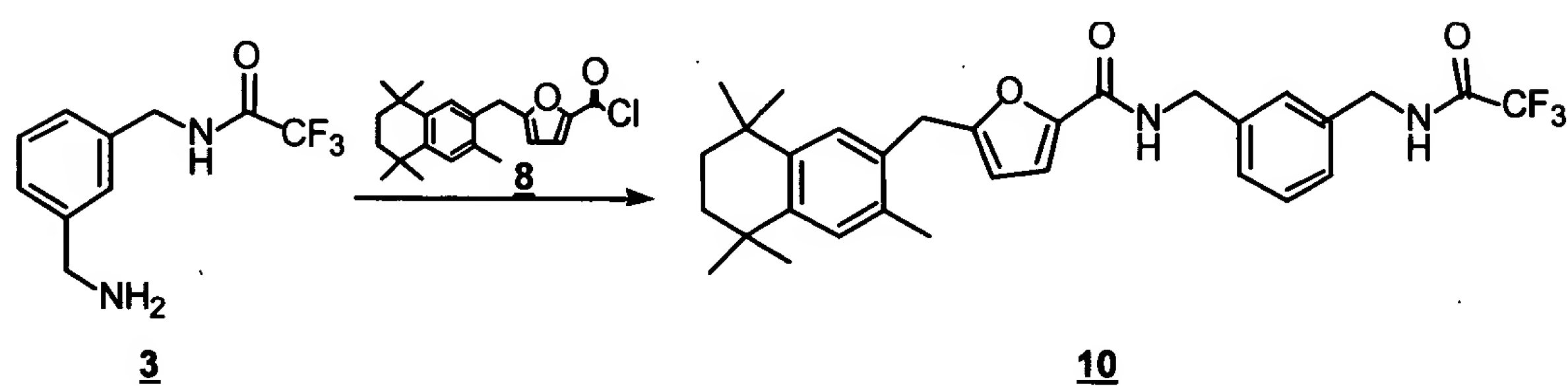
Ethyl 3-[(2-[(2R)-tetrahydro-2-furanylmethyl]amino)-4-pyrimidinayl]amino]methyl}benzylcarbamate 6 was dissolved in ethylene glycol and potassium hydroxide (1:1 v/v). The solution was heated to 100°C overnight. The mixture was cooled to room temperature and extracted with chloroform, washed with brine, and dried over magnesium sulfate to yield N⁴-[3-(aminomethyl)benzyl]-N²-[(2R)-tetrahydro-2-furanylmethyl]-2,4-pyrimidinediamine 7 (82 % yield).

15 Preparation of 5-[(3,5,5,8,8-pentamethyl-5,6,7,8-tetrahydro-2-naphthalenyl)methyl]-N-(3-[(2-[(2R)-tetrahydro-2-furanylmethyl]amino)-4-pyrimidinyl] amino]methyl}benzyl)-2-furamide 9.

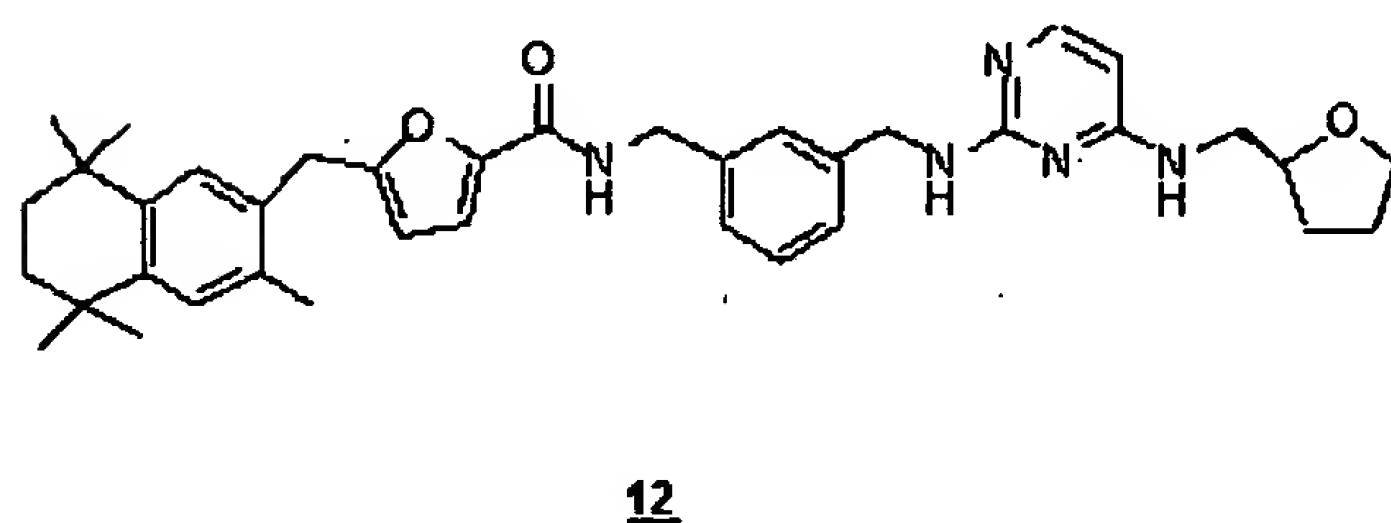
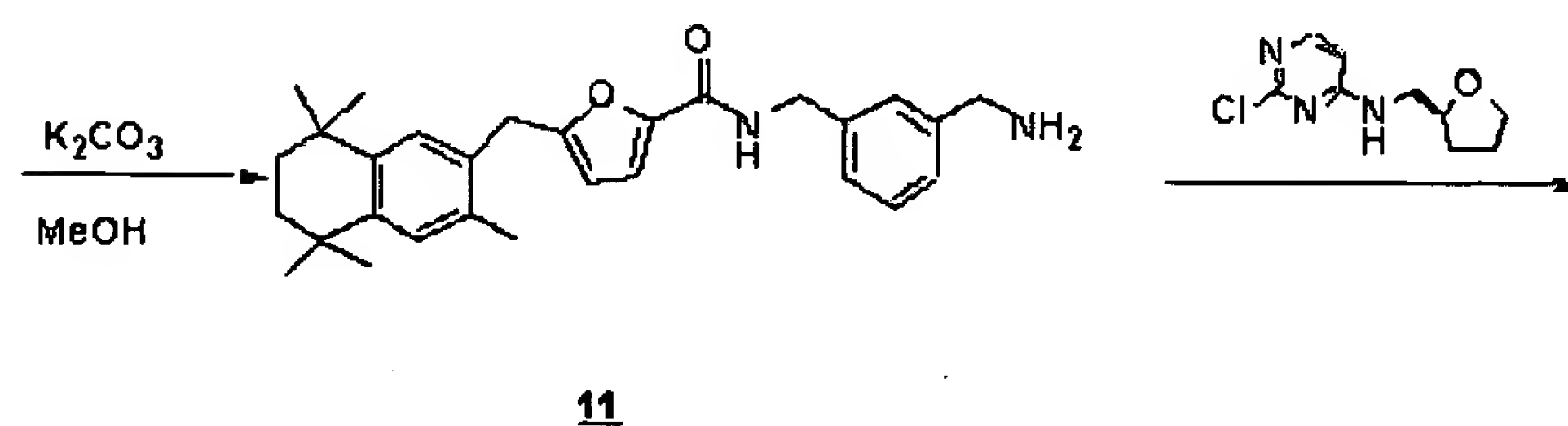
Ethyl 3-[(2-[(2R)-tetrahydro-2-furanylmethyl]amino)-4-pyrimidinayl]amino]methyl}benzylcarbamate 6 was dissolved in ethylene glycol and potassium hydroxide (1:1 v/v). The solution was heated to 100°C overnight. The mixture was cooled to room temperature and extracted with chloroform, washed with brine, and dried over magnesium sulfate to yield N⁴-[3-(aminomethyl)benzyl]-N²-[(2R)-tetrahydro-2-furanylmethyl]-2,4-pyrimidinediamine 7 (82 % yield). This product, 7, (182 mg, .580 mmol) and 2-furoyl chloride reagent 8 was dissolved dichloromethane followed by triethylamine. Reaction was stirred at room temperature overnight. Crude compound was purified on silica gel column and eluted ethyl acetate/hexane (4:1 v/v) to yield 5-[(3,5,5,8,8-pentamethyl-5,6,7,8-tetrahydro-2-naphthalenyl)methyl]-N-(3-[(2-[(2R)-tetrahydro-2-furanylmethyl]amino)-4-pyrimidinyl] amino]methyl}benzyl)-2-furamide 9 (159.1 mg). ¹H NMR (CDCl₃): δ 1.19 (s, 6H), 1.26 (s, 6H), 1.65 (m, 5H), 1.92 (m, 3H), 2.23 (s, 3H), 3.45 (m, 1H), 3.5 (m, 1H), 3.7 (m, 1H), 3.9 (m, 3H),

4.05 (m, 1H), 4.50 (d, 2H), 4.58 (d, 2H), 5.01 (brd, 1H), 5.30 (brd, 1H), 5.71 (d, 1h), 6.03 (d, 1H), 6.61 (t, 1H), 6.99 (s, 1H), 7.06 (s, 1H), 7.07 (s, 1H), 7.28 (m, 4H), 7.80 (d, 1H). MS: 622.4 (M+1).

5 **Synthesis of 5-[(3,5,5,8,8-pentamethyl-5,6,7,8-tetrahydro-2naphthalenyl)methyl]-N-methyl-N-(3-[[4-[(2S)-tetrahydro-2furanylmethyl]amino]-2-pyrimidinyl]amino]methyl}benzyl)-2-furamide 12.**



10



15

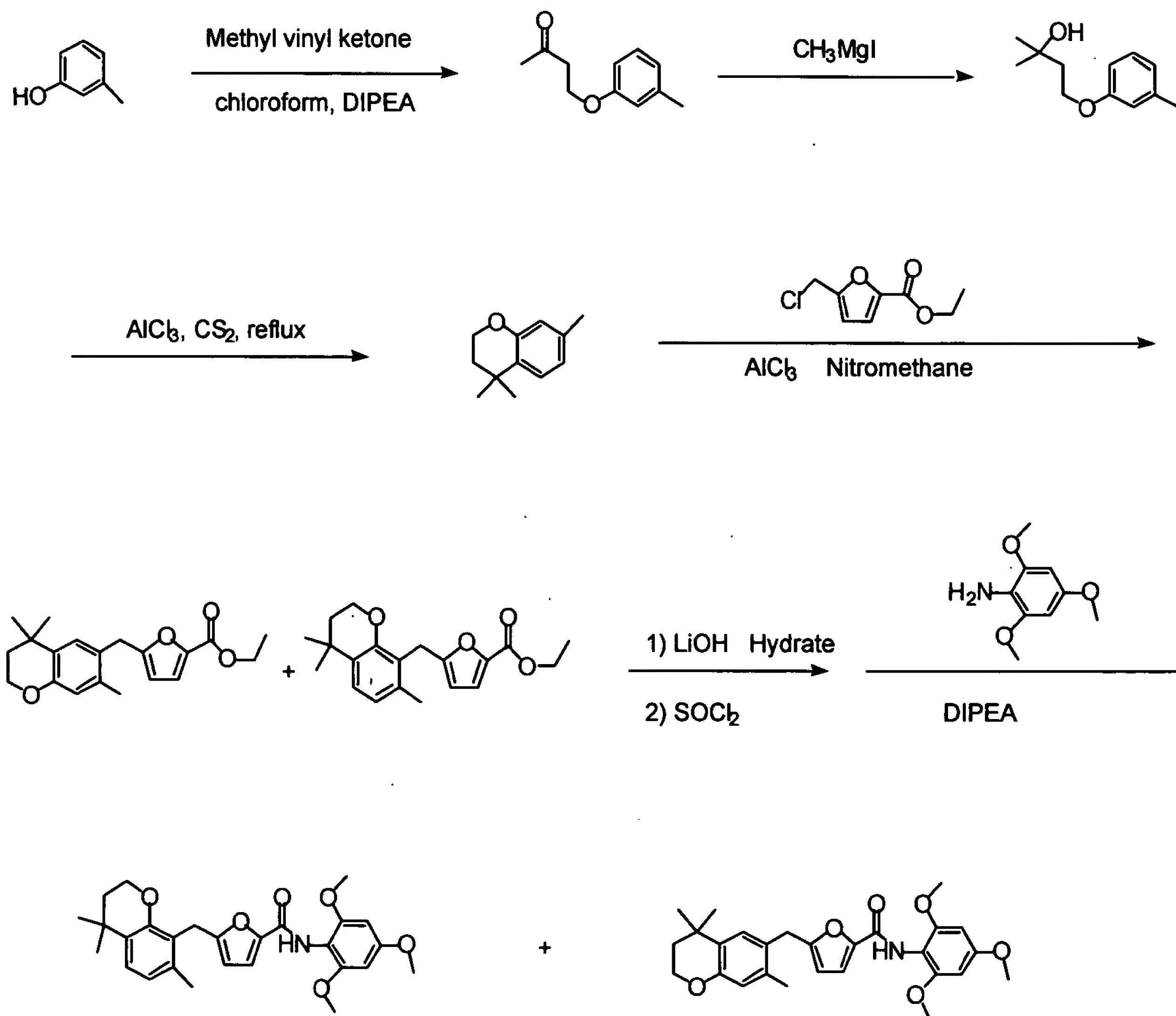
Preparation of N-[3-(aminomethyl) benzyl]-5-[(3,5,5,8,8-pentamethyl-5,6,7,8-tetrahydro-2naphthalenyl)methyl]-2-furamide 11.

To a solution of N-[3-(aminomethyl)benzyl]-2,2,2-trifluoroacetamide **3** and 2-furoyl chloride reagent **8** was added triethylamine. Reaction mixture was stirred at room temperature for 1 hour. Crude mixture was purified by silica gel chromatography eluting with hexane/ethyl acetate (4:1 v/v) to yield 5-[(3,5,5,8,8-pentamethyl-5,6,7,8-tetrahydro-2-naphthalenyl)methyl]-N-(3-[[trifluoroacetyl]amino]methyl)benzyl)-2-furamide **10**. The purified compound was dissolve in methanol (100 mL) and potassium carbonate in water (2M, 100 mL). Reaction was heated to 70°C overnight. The solution was cooled to room temperature, basified with 20% NaOH to pH 14, extract with methylene chloride, washed with brine, and dried over magnesium sulfate to yield N-[3-(aminomethyl) benzyl]-5-[(3,5,5,8,8-pentamethyl-5,6,7,8-tetrahydro-2naphthalenyl)methyl]-2-furamide **11** (4.97g, 85.1 % yield).

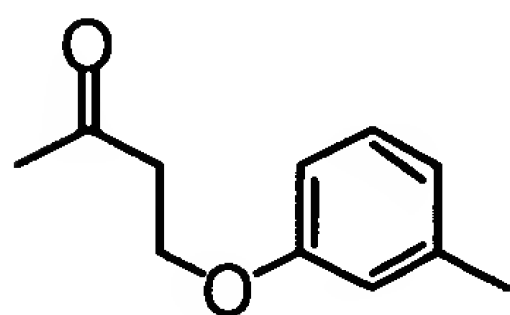
Preparation of 5-[(3,5,5,8,8-pentamethyl-5,6,7,8-tetrahydro-2naphthalenzyl)methyl]-N-(3-[[4-[(2S)-tetrahydro-2furanylmethyl]amino]-2-pyrimidinyl]amino]methyl)benzyl)-2-furamide **12.**

To a solution of N-[3-(aminomethyl) benzyl]-5-[(3,5,5,8,8-pentamethyl-5,6,7,8-tetrahydro-2naphthalenyl)methyl]-2-furamide **11** in chlorobenzene was added and 2-chloro-N-[(2S)-tetrahydro-2-furanmethyl]-4-pyrimidinamine and triethylamine. Reaction mixture was reflux overnight. The cooled mixture was then purified by silica gel chromatography followed by HPLC. To yield of 5-[(3,5,5,8,8-pentamethyl-5,6,7,8-tetrahydro-2naphthalenzyl) methyl]-N-(3-[[4-[(2S)-tetrahydro-2furanylmethyl]amino]-2-pyrimidinyl]amino]methyl)benzyl)-2-furamide **12**. ¹H NMR (CDCl₃): δ 1.01 (s, 6H), 1.25 (s, 6H), 1.43 (m, 1H), 1.56 (s, 4H), 1.70-1.98 (m, 3H), 2.13 (s, 3H), 3.24 (m, 1H), 3.61 (m, 1H), 3.63-3.80 (m, 2H), 3.82 (s, 2H), 3.87-4.06 (m, 1H), 4.37-4.60 4.37 (d, 2H), 4.60 (d, 2H), 5.73 (d, 1H), 5.92 (d, 1H), 6.3 (brd, 1H), 6.75 (brd, 1H), 6.92 (s, 1H), 6.98 (d, 1H), 7.0 (s, 1H), 7.09-7.26 (m, 4H), 7.4 (s, 1H), 9.5 (brd, 1H). MS (APCI): 622.3 (M+1).

Examples of Heterocyclic-Containing Compounds



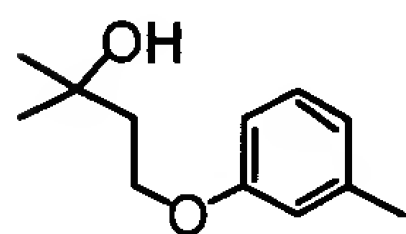
5 4-(3-methylphenoxy)-2-butanone:



To m-Cresol (4.0 g, 37 mmol), methyl vinyl ketone (3.2 mL, 37 mmol) in chloroform
 10 (25 mL), was added diisopropyl ethyl amine. The mixture was heated at reflux for 16 h,

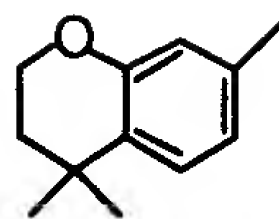
allowed to cool to room temperature and evaporated. The residue has 50% product and 50% starting material. The starting material was separated as t-butyldimethyl silyl ether. The product was isolated via plug filtration using silica gel 50% hexane/ethyl acetate. Yield 4.5 g (68%). As an alternative purification procedure, the crude reaction mixture was evaporated, dissolved in DMF (0.2M) and 0.5 equivalents of imidazole and 0.5 equivalents of tBDMSCl were added. The reaction was stirred for 3 hr at room temperature and then the solvents were removed in vacuo. To the residue was added 75 mL ethyl acetate and 75 mL water (ratio of 1/1). The ethyl acetate layer was separated and dried over Na₂SO₄. The solvents were removed in vacuo. The crude material was placed on a pad of silical gel and the silylated m-cresol was removed with hexanes. The product was obtained by eluting with 5-10% ethyl acetate in hexanes. The solvents were removed in vacuo to give the desired product. ¹H(CDCl₃): 7.15 (t, 1H), 6.65-6.80(m, 2H), 4.25 (t, 2H), 2.75 (t, 2H), 2.25 and 2.35 (2s, 3H each).

2-Methyl-4(3-methylphenoxy)-2-butanol:



To a solution of methyl magnesium bromide in ether (50 mL), prepared from Mg (572 mg, 23.56 mmol) and MeI (3.34 g, 23.56 mmol), was added 4-(3-methylphenoxy)-2-butanone (2.1 g, 11.78 mmol) in 10 mL ether. The solution was stirred at room temperature for 30 minutes, after which it was quenched with water and dilute Hydrochloric acid. The organic layer was separated, dried over sodium sulfate, filtered through a silica plug to give a colorless syrup 1.91 g (83%) mass spectral analysis using APCI +ve 177 (M⁺-OH).

4,4,7-trimethyl chroman:

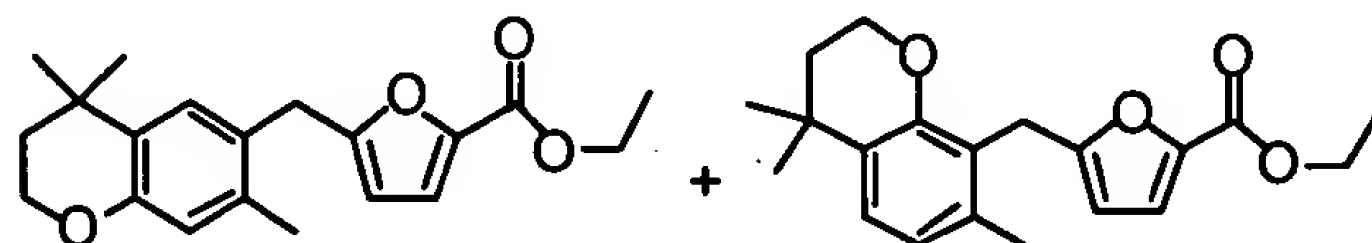


To aluminium chloride (1.3 g, 9.79 mmol) in 40 mL carbon disulfide, was added 2-methyl-4(3-methylphenoxy)-2-butanol (1.9 g, 9.79 mmol) in 10 mL carbon disulfide.

5 The mixture was heated at reflux for 2 h. Solvent evaporated, the residue diluted with 50 mL of ethyl acetate, and 10 mL of water. The organic layer was separated, dried over sodium sulfate, and purified via column chromatography to give a light yellow syrup 1.5 g (87 %). ¹H(CDCl₃): 7.05(br d, 1H), 6.87 (dd, 1H), 6.69 (d, 1H), 4.20 (t, 2H), 2.35 (s, 3H), 1.80 (t, 2H), 1.40 (s, 6H).

10

Ethyl-5-[(4,4,7-trimethyl-3,4-dihydro-2H-chromen-6-yl)methyl]-2-furoate, and ethyl-5-[(4,4,7-trimethyl-3,4-dihydro-2H-chromen-8-yl)methyl]-2-furoate:

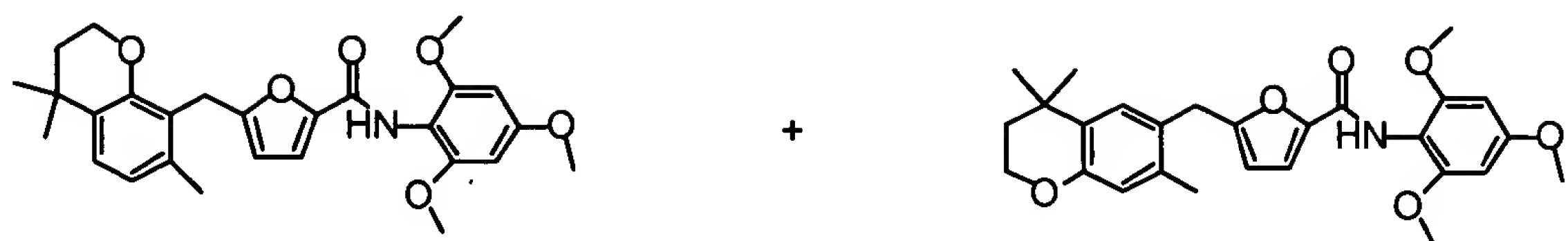


15

To zinc chloride (950 mg, 6.97 mmol) in nitromethane (20 mL), was added a mixture of 4,4,7-trimethyl chroman (1.23 g, 6.97 mmol) and ethyl –5-chloromethyl-2-furoate (656 mg, 3.48 mmol) in nitromethane (15 mL). The mixture was stirred at room temperature for 16 h. The reaction was evaporated to dryness and triturated with ethyl acetate-water(1:1, 100 mL). The organic layer on usual work up, and plug filtration using hexanes in ethyl acetate (9:1) gave mixture of these two compounds. 1.34 g (46% based on chroman).

20

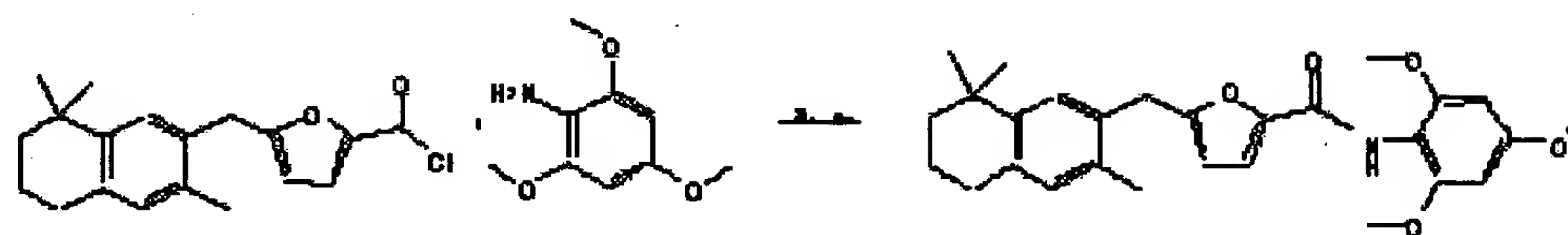
25 **N-(2,4,6-trimethoxyphenyl)-5-[(4,4,7-trimethyl-3,4-dihydro-2H-chromen-6-yl)methyl]-2-furamide and N-(2,4,6-trimethoxyphenyl)-5-[(4,4,7-trimethyl-3,4-dihydro-2H-chromen-8-yl)methyl]-2-furamide:**



To a mixture of ethyl-5-[(4,4,7-trimethyl-3,4-dihydro-2H-chromen-6-yl)methyl]-2-furoate, and ethyl-5-[(4,4,7-trimethyl-3,4-dihydro-2H-chromen-8-yl)methyl]-2-furoate (1.34 g, 3.74 mmol) in THF-MeOH-H₂O (7:5:5, 20 mL) was added lithium hydroxide monohydrate (784 mg, 18.7 mmol). The mixture was stirred for 4 h at room temperature. The mixture evaporated to dryness, diluted with 30 mL ethyl acetate and 50 mL of water. After acidification with diluted HCl, ethyl acetate layer was separated, dried and evaporated in vacuo to give a mixture of corresponding acids, 1.03 g (quantitative). These acids were not separable using typical column chromatography or crystallization. To the mixture of the acids (200 mg, 0.66 mmol) in dichloromethane (30 mL) was added thionyl chloride (392 mg, 3.3 mmol). The mixture was refluxed for 1 h and evaporated. The residue was dissolved in hexane-ethyl acetate (9:1, 20 mL) and filtered through a silica gel plug (0.5 cm X 1.0 cm). To the residue in 10 mL ethyl acetate, was added 2,4,6-trimethoxyphenyl amine hydrochloride (145 mg, 0.66 mmol) followed by diisopropyl ethyl amine (256 mg, 1.98 mmol). The mixture was stirred at room temperature for 16 h. The reaction was quenched with water (10 mL), ethyl acetate layer separated. The combination and column and HPLC purification gave 15 mg and 21 mg of two components (12 %). The isomers were separated using reverse phase HPLC chromatography. Linear isomer: ¹H(CDCl₃): 7.46(br s, 1H), 7.23 (br s, 1H), 7.14 (br s, 1H), 7.02 (s, 2H), 6.63 (s, 1H), 6.15 (s, 2H), 6.0(d, 1H), 4.17 (t, 2H), 3.93 (s, 3H), 3.81, 3.80 (2s, 3H each), 2.21 (s, 3H), 1.81 (t, 2H), 1.29 (s, 6H). M⁺ at 466.2. Angular isomer: AXC07302: 7.25 (br s, 1H), 6.91 (d, 1H), 6.88 (d, hidden, 1H), 6.53 (d, 1H), 5.95 (s, 2H), 5.72 (d, 1H), 3.96 (t, 2H), 3.8 (s, 3H), 3.6 (s, 6H), 2.07 (s, 3H), 1.59 (t, 2H), 1.11 (s, 6H). M⁺ at 466.1

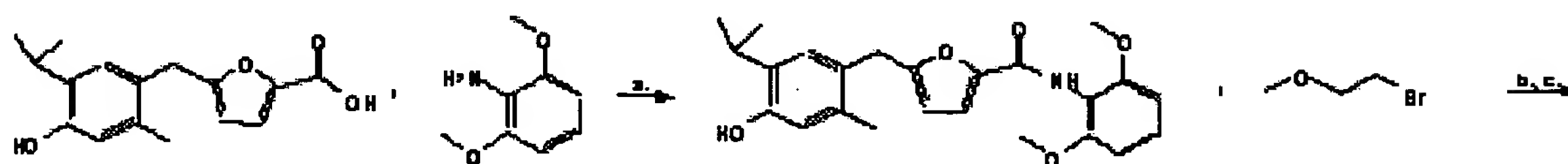
Examples of Aromatic Compounds

Compound A



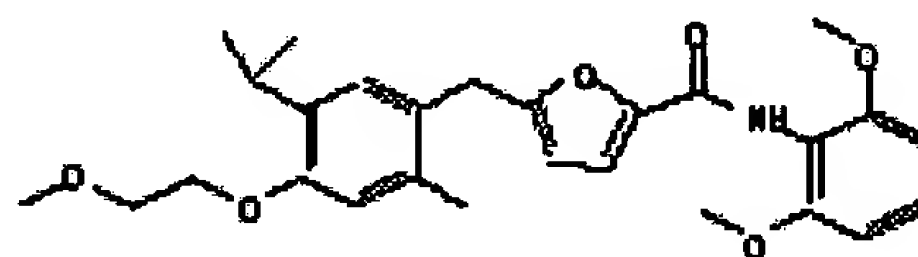
a. Ethyl Acetate, triethyl amine, overnight

Compound B



a. HATU, diisopropyl ethyl amine, DMF, 24 hours

b. potassium t-butoxide in 20% MeOH/DMF, 30 min c. bromide reagent, 20% MeOH/DMF 48 hours



Compound 183

Thymol (1.0 eq, 33.3mmol) and methyl 5-(chloromethyl)-2-furoate (1.0 eq, 33.3mmol) were dissolved in nitromethane (120mL, 0.2M). Aluminum trichloride (1.0 eq, 33.3mmol) dissolved in 25mL nitromethane was added to the above solution under nitrogen and heated to slow reflux over 10 min. The heat was turned off and left under nitrogen overnight. The reaction was quenched with 100mL of water and extracted

with dichloromethane. The crude mixture was evaporated to dryness and loaded onto plug chromatography column (1g crude/100g silica gel ratio). The column was eluted with 7 and 11% ethyl acetate/hexanes to yield the desired product (2.9 g, 30%). The ester was hydrolyzed to acid by lithium hydroxide in THF/MeOH/H₂O (35/25/25).

5

To a solution containing the 5-(4-hydroxy-5-isopropyl-2-methylbenzyl)-2-furoic acid (1.0eq, 3.6 mmol, 0.5M), and 2,6-dimethoxyaniline (1.0eq, 3.6mmol) were dissolved in DMF. To this mixture, HATU (1.0 eq, 3.6 mmol) and di-isopropyl ethyl amine (1.0 eq, 3.6 mmol) were added and stirred overnight. The mixture was heated for 10min at 45°C. The solution was placed into ethyl acetate (3x volume) and washed with water. The organic layer was evaporated to syrup and eluted on plug column chromatography (1:100 g crude/g silicagel) with 30 and 50 % ethyl acetate/hexane to yield: N-(2,6-dimethoxyphenyl)-5-(4-hydroxy-5-isopropyl-2-methylbenzyl)-2furamide (820 mgs, 55% yield). ¹HNMR (CDCl₃) 7.22ppm (1H, t, J=8.68 Hz), 7.08ppm (1H, d, J=3.40 Hz), 6.99ppm (1H,s), 6.64ppm (2H, d, J=8.68 Hz), 6.61ppm (1H, s), 5.97 ppm (1H, d, J=3.40 Hz), 3.95ppm, (2H,s), 3.85ppm, (6H,s), 3.17ppm, (1H,pentet, J=6.8 Hz) 2.23ppm, (3H,s), 1.25 ppm(3H,s), and 1.23ppm (3H, s).

Potassium t-butoxide was dissolved (1.05eq, 0.128mmol) in MeOH (24 μL). To the solution of above furamide (1.0eq, 0.122mmol, 1M) in DMF, t-butoxide solution was added and stirred for 30 min.. The 2-Bromoethyl methyl ether (1.0eq, 0.122 mmol) was added (20%MeOH/DMF, 1M) and stirred at room temperature for 48 hours and purified by reverse phase HPLC (method: 35-75% 90 min acetonitrile in 0.1% aqueous TFA) to yield (8.5 mgs, 15% yield). ¹H NMR (CDCl₃): 7.04 ppm(1H, J=8.31 Hz, t), 6.85 ppm (1H, J=3.40, d), 6.80 ppm (1H, s), 6.53 ppm (1H, s), 6.48 (2H, J=8.31 Hz, d), 5.76 (1H, J=3.40 Hz, d), 3.88 (2H, J=3.40/4.53 Hz, dd), 3.78 (2H, s), 3.58 (6H, s), 3.54 (2H, J=3.40/4.54 Hz, dd), 3.20 (3H, s), 3.07(1H, J=7.2 Hz, pentet), 2.04 (3H, s), 0.97 (3H, s), 0.95(3H,s).

30

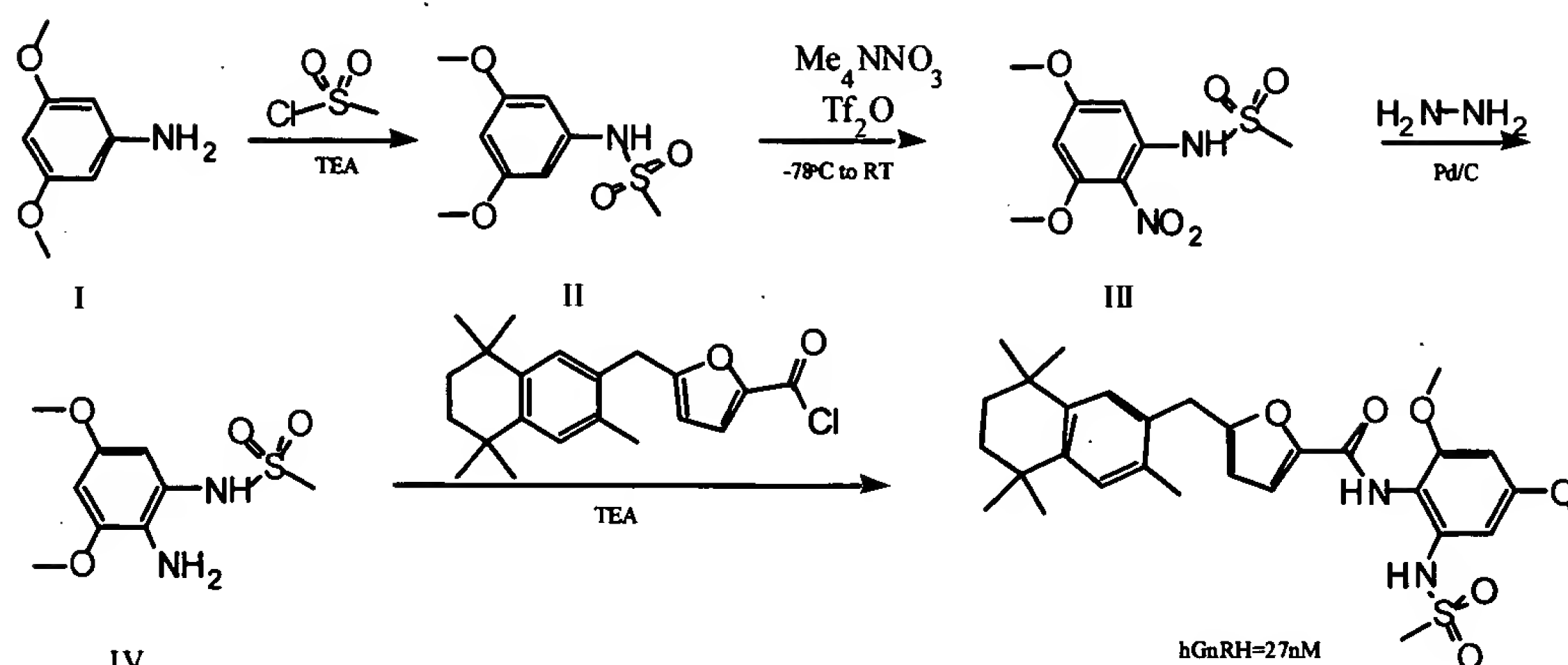
Compound A

1,1,6-trimethyl-1,2,3,4-tetrahydronaphthalene was synthesized from reference : John J. Parlow *Tetrahedron* Vol 49 (13) 2577. It was then connected with methyl 5-(chloromethyl)-2-furoate by Friedles-Crafts reaction as previously stated to yield two
5 major regio-isomers. The desired isomer was separated after hydrolysis using three to five successive recrystallization from 10% acetone/heptane (1g/10mL) system. The acid was then converted to acid chloride with thionyl chloride as previously stated.

To a solution of 5-[(3,8,8-trimethyl-5,6,7,8-tetrahydro-2-naphthalenyl)methyl]-2-furoyl
10 chloride (1.0eq, 0.32 mmol, 0.2M) in 2mL ethyl acetate, the 2,4,6 trimethoxy aniline mono HCl salt (1.0eq, 0.32 mmol) was added. Triethyl amine(excess) was added to this mixture and stirred overnight. The crude product was dried under *vacuo* and purified through plug column chromatography (1:100 crude mass/silicagel ratio) by eluting with 20 and 30 percent ethyl acetate/hexane solution. In some cases, the regio-
15 isomers were separated by recrystallization in twenty five percent ethyl acetate/hexanes (1g/75mL compound/volume) to yield N-(2,4,6-trimethoxyphenyl)-5-[(3,8,8-trimethyl-5,6,7,8-tetrahydro-2-naphthalenyl)methyl]-2-furamide (120mgs, 82% yield).
¹H NMR (CDCl₃) 7.28ppm(1H, broad), 7.12 (1H, s), 7.08 (1H, J=3.40 Hz, d), 6.89 (1H, s), 6.19 (2H, s), 6.00 (1H, J=3.40 Hz, d), 3.97 (2H, s), 3.83 (3H, s), 3.82 (6H, s)
20 2.73 (2H, J=6.05 Hz, t), 2.25 (3H, s), 1.84-1.76(2H, multiplet), 1.69-1.63 (2H, multiplet), 1.59 (3H, s), 1.26(6H,s) elemental: expected C(72.55), H(7.18), N(3.02); actual C(72.67), H(7.22), N(2.98).

Compound 228

25



Compound II To a solution of 3,5-dimethoxyaniline (Compound I, 1.53g, 10mmol) in DCM (20ml) was added methanesulfonyl chloride (0.88mL, 10mmol). TEA

5 (1.40mL, 10mmol) was added dropwise. The reaction mixture was stirred at rt for 15h. The crude product was taken to dryness and purified by flash chromatography (30% ethyl acetate/hexanes), yielding compound II (2.10g, 91%) as a white solid. ^1H NMR δ (300Hz, CDCl_3) 2.96(s, 3H), 3.71 (s, 6H), 6.20 (d, 1H, $J=3\text{Hz}$), 6.34 (d, 2H, $J=3\text{Hz}$), 6.76 (s, 1H). APCI-MS m/z 232 ($\text{M}+\text{H}$) $^+$.

10

Compound III To a solution of $(\text{CH}_3)_4\text{NNO}_3$ (1.12g, 7.89mmol) in DCM (10mL) was added triflic anhydride dropwise. The reaction mixture was stirred at 0°C for 1.5h. In a dropping funnel was placed the compound II (1.75g, 7.51mmol) in 10mL of DCM and the solution was added to the nitronium triflate reaction mixture at -78°C . The reaction

15 mixture was kept in -78°C for 30min and gradually warmed to rt. It was stirred for 15h. The reaction was quenched with 5% NaHCO_3 (50mL), the mixture was stirred for 30 min. The aqueous layer was extracted with DCM (3X20mL). Combined DCM layer was dried over Na_2SO_4 . The crude product was purified by HPLC, yielding compound III (250mg, 11%) as a white solid. ^1H NMR δ (300Hz, CDCl_3) 3.03(s, 3H), 3.87 (s, 3H), 3.90 (s, 3H), 6.32 (d, 1H, $J=3\text{Hz}$), 6.88 (d, 1H, $J=3\text{Hz}$), 8.31 (s, 1H). APCI-MS m/z 275 ($\text{M}-\text{H}$) $^-$.

20

Compound IV To a solution of compound III (106mg, 0.38mmol) in EtOH (2mL) was added 20mg Pd/C and NH₂NH₂ (1mL). The reaction mixture was refluxed for 9h. The reaction mixture passes through a celite pad. The solution was taken to dryness to afford compound IV as a brown solid (90mg, 95%). This compound was used directly
5 in the next step.

Compound 228

To a solution of compound IV (39mg, 0.16mmol), 5-[(3,5,5,8,8-pentamethyl-5,6,7,8-tetrahydro-2-naphthalenyl)-2-furoyl chloride (60mg, 0.174mmol) in DCM
10 (1mL) was added TEA (44μL, 0.31mmol). The reaction mixture was stirred at rt for overnight. Flash chromatography (30% ethyl acetate/hexanes) yields Compound 228 (65mg, 74%) as a white solid. ¹H NMR δ (300Hz, CDCl₃) 1.04 (m, 12H), 1.45 (m, 4H), 2.09 (s, 3H), 2.71(s, 3H), 3.60 (s, 3H), 3.63 (s, 3H), 3.78 (s, 2H), 5.89 (d, 1H, J=3Hz), 6.17 (d, 1H, J=3Hz), 6.58 (d, 1H, J=3Hz), 6.85 (s, 1H), 6.90 (s, 1H), 6.96 (d,
15 1H, J=3Hz), 7.90 (s, 1H), 8.24 (s, 1H). APCI-MS *m/z* 556 (M+H)⁺.

Additional Examples (NMR) of Some Compounds are Shown Below

Compound 20

¹H NMR δ (300Hz, CDCl₃) 1.26 (m, 12H), 1.66 (m, 4H), 2.28 (s, 3H), 3.82 (s, 6H),
20 3.96 (s, 2H), 6.04 (d, 1H, J=6Hz), 6.60 (s, 1H), 6.62 (s, 1H), 7.05 (m, 3H), 7.23 (t, 1H, J=6Hz), 7.44 (s, 1H). APCI-MS *m/z* 462 (M+H)⁺.

Compound 126

¹H NMR δ (300Hz, DMSO) 2.20 (s, 3H), 2.27 (s, 6H), 3.71 (s, 6H), 3.98 (s, 2H), 5.93
25 (d, 1H, J=3Hz), 6.68 (s, 1H), 6.69 (s, 1H), 6.86 (d, 2H, J=3Hz), 7.10 (d, 1H, J=3Hz), 7.23 (t, 1H, J=7Hz), 9.04 (s, 1H). APCI-MS *m/z* 379 (M+H)⁺.

Compound 140

^1H NMR δ (300Hz, DMSO) 2.28 (s, 3H), 2.32 (s, 3H), 2.42 (s, 3H), 3.72 (s, 6H), 4.10 (s, 2H), 6.00 (d, 1H, $J=3\text{Hz}$), 6.60 (s, 1H), 6.68 (s, 1H), 6.71 (s, 1H), 7.10 (m, 2H), 7.24 (t, 1H, $J=6\text{Hz}$), 9.04 (s, 1H). APCI-MS m/z 458 ($\text{M}+\text{H}$) $^+$.

5 Compound 211

^1H NMR δ (300Hz, CDCl_3) 3.76-3.83 (m, 15H), 4.07 (s, 2H), 5.95 (d, 1H, $J=3\text{Hz}$), 6.60 (s, 1H), 6.57-6.62 (m, 3H), 6.78 (d, 1H, $J=9\text{Hz}$), 7.03 (d, 1H, $J=3\text{Hz}$), 7.19 (t, 1H, $J=6\text{Hz}$), 7.46 (s, 1H). APCI-MS m/z 428 ($\text{M}+\text{H}$) $^+$.

10 Compound 220

^1H NMR δ (300Hz, CDCl_3) 3.72-3.77 (m, 15H), 3.90 (s, 2H), 5.78 (d, 1H, $J=3\text{Hz}$), 6.08 (s, 2H), 6.52-6.55 (m, 2H), 6.94 (d, 1H, $J=9\text{Hz}$), 7.12 (t, 1H, $J=9\text{Hz}$), 7.39 (s, 1H). APCI-MS m/z 428 ($\text{M}+\text{H}$) $^+$.

15 Compound 226

^1H NMR δ (300Hz, CD_3OD) 2.33-2.48 (m, 12H), 3.84 (s, 6H), 4.19 (s, 2H), 5.81 (d, 1H, $J=3\text{Hz}$), 6.73 (d, 2H, $J=9\text{Hz}$), 7.06 (d, 1H, $J=3\text{Hz}$), 7.29 (t, 1H, $J=9\text{Hz}$). APCI-MS m/z 472 ($\text{M}+\text{H}$) $^+$.

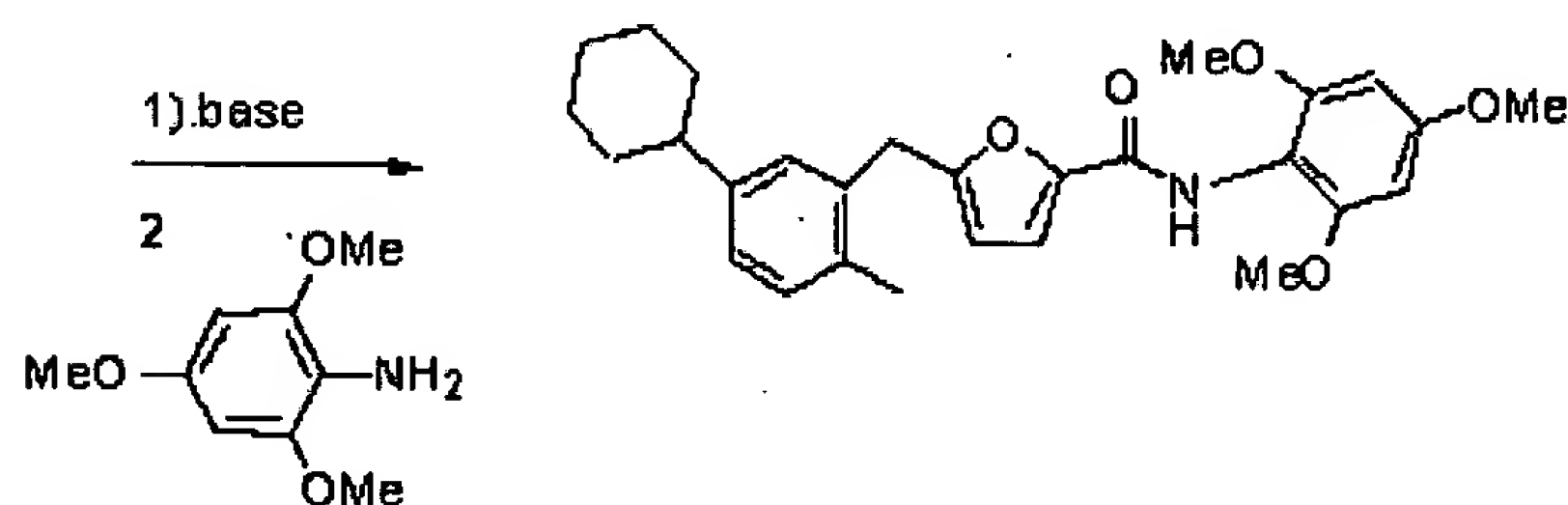
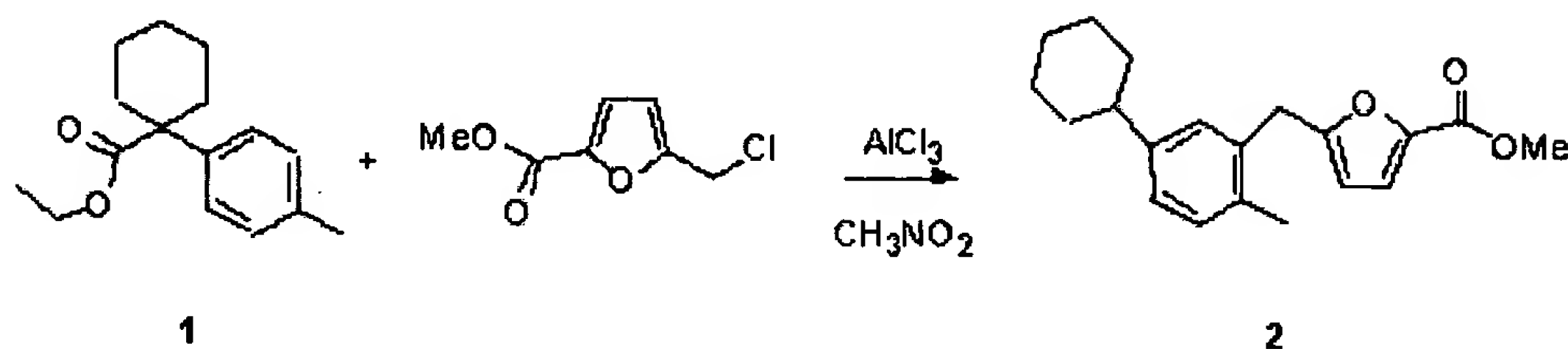
20 Compound 231

^1H NMR δ (300Hz, CDCl_3) 0.67 (t, 3H, $J=6\text{Hz}$), 1.27 (m, 6H), 1.70 (m, 2H), 3.62 (s, 3H), 3.80-3.83 (m, 6H), 4.02 (s, 2H), 6.02 (d, 1H, $J=3\text{Hz}$), 6.60 (d, 2H, $J=9\text{Hz}$), 6.83 (d, 1H, $J=3\text{Hz}$), 7.07-7.19 (m, 4H), 7.43 (s, 1H). APCI-MS m/z 438 ($\text{M}+\text{H}$) $^+$.

25 Compound 232

^1H NMR δ (300Hz, CDCl_3) 0.66 (t, 3H, $J=6\text{Hz}$), 1.23 (m, 6H), 1.54-1.62 (m, 2H), 3.80-3.81 (m, 12H), 4.01 (s, 2H), 6.00 (d, 1H, $J=3\text{Hz}$), 6.81 (d, 2H, $J=9\text{Hz}$), 6.82 (d, 1H, $J=3\text{Hz}$), 7.06-7.28 (m, 4H). APCI-MS m/z 468 ($\text{M}+\text{H}$) $^+$.

Examples of Other Aromatic Containing Compounds



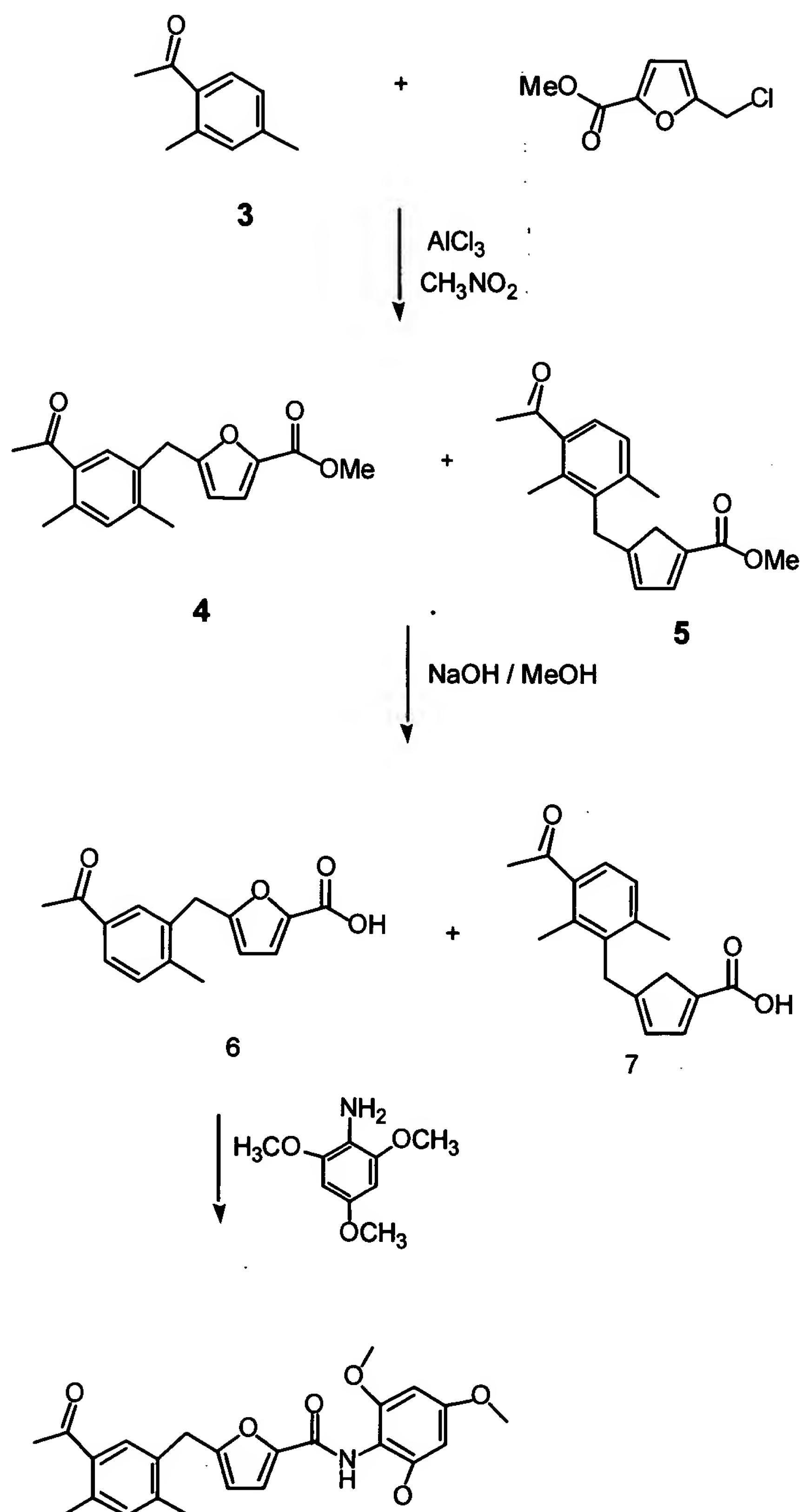
5

Synthesis of 5-(5-cyclohexyl-2-methylbenzyl)-N-(2,4,6-trimethoxyphenyl)-2-furamide.

To a mixture of compounds 1 (5.0g, 20.3 mmol) and the methyl furoate (3.5g, 20.3 mmol) in 100 ml of nitromethane was added a solution of AlCl₃ (5.4g, 40.6 mmol) in

10 CH₃NO₂ (50 ml) at room temperature. The solution was heated to 70 ~ 75⁰C overnight. The dark brown mixture was cooled to room temperature and slowly poured into 300 ml of ice water. The mixture was extracted with ethylacetate. The concentrated organic layer was purified by silica gel chromatography and eluted with hexane/ethyl acetate (15:1 to 9:1 v/v) to yield 920 mg of compound 2, which was then

15 hydrolyzed and coupled with the trimethoxyaniline according to the general procedure to give the compound in a good yield. ¹HNMR (CDCl₃): σ 1.23-1.48 (m, 5H), 1.73-1.86 (m, 5H), 2.31 (s, 3H), 2.51 (m, 1H), 3.83 (s, 1H), 4.02 (s, 2H), 6.03 (s, 1H), 7.44 (s, 1H) MS (APCI): 464.2 (M+1)



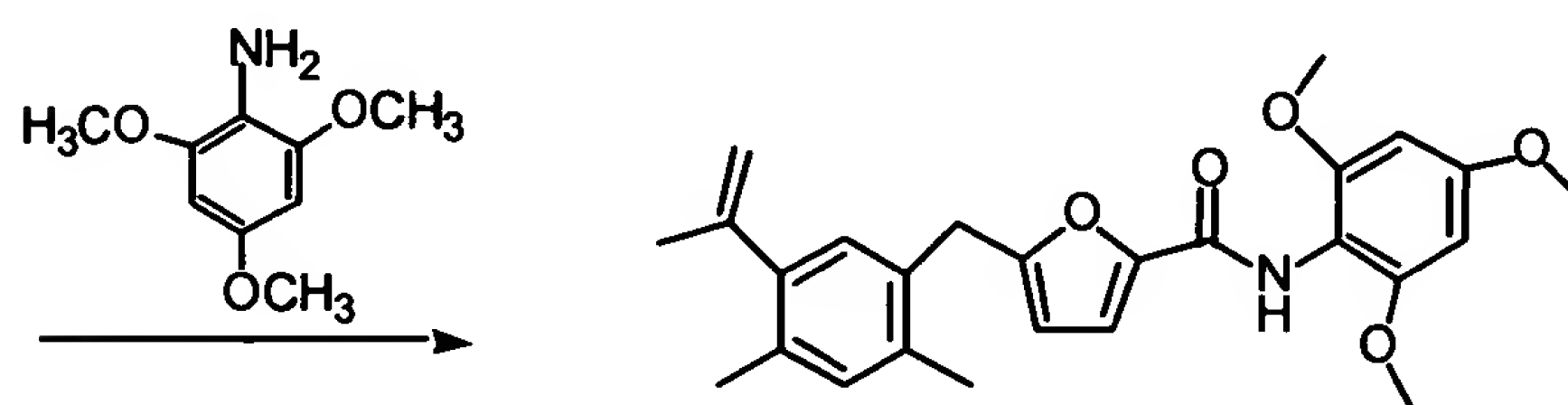
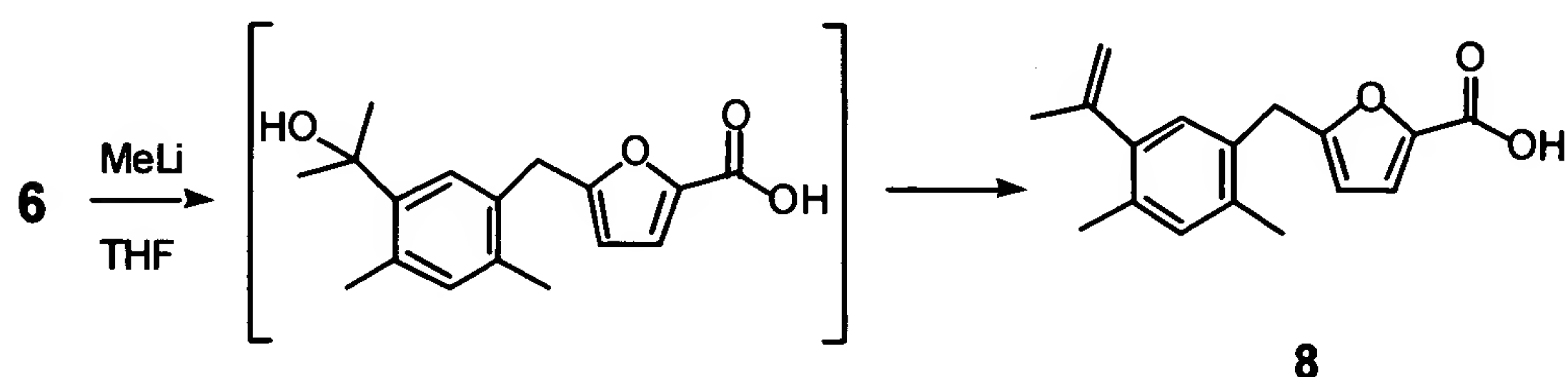
Synthesis of 5-(5-acetyl-2, 4-dimethylbenzyl)-N-(2,4,6-trimethoxyphenyl)-2-furamide.

A mixture of compound 3 (14g, 94.4 mmol), methyl furoate (16.4g, 94.4 mmol) and AlCl_3 (25g, 189 mmol) in 200 mL of nitromethane was stirred and heated to 80°C overnight. The mixture was worked up and purified by silica gel column, eluted with hexane/ethyl acetate (9:1 v/v) to give a mixture of 4 and 5 (3:1, total 16.2g).

The mixture of 4 and 5 (2.0g) was hydrolyzed in 2N NaOH/MeOH (1:1 v/v) at room temperature to give a mixture of 6 and 7, which was recrystallized in acetone and heptane to afford 6 (460 mg).

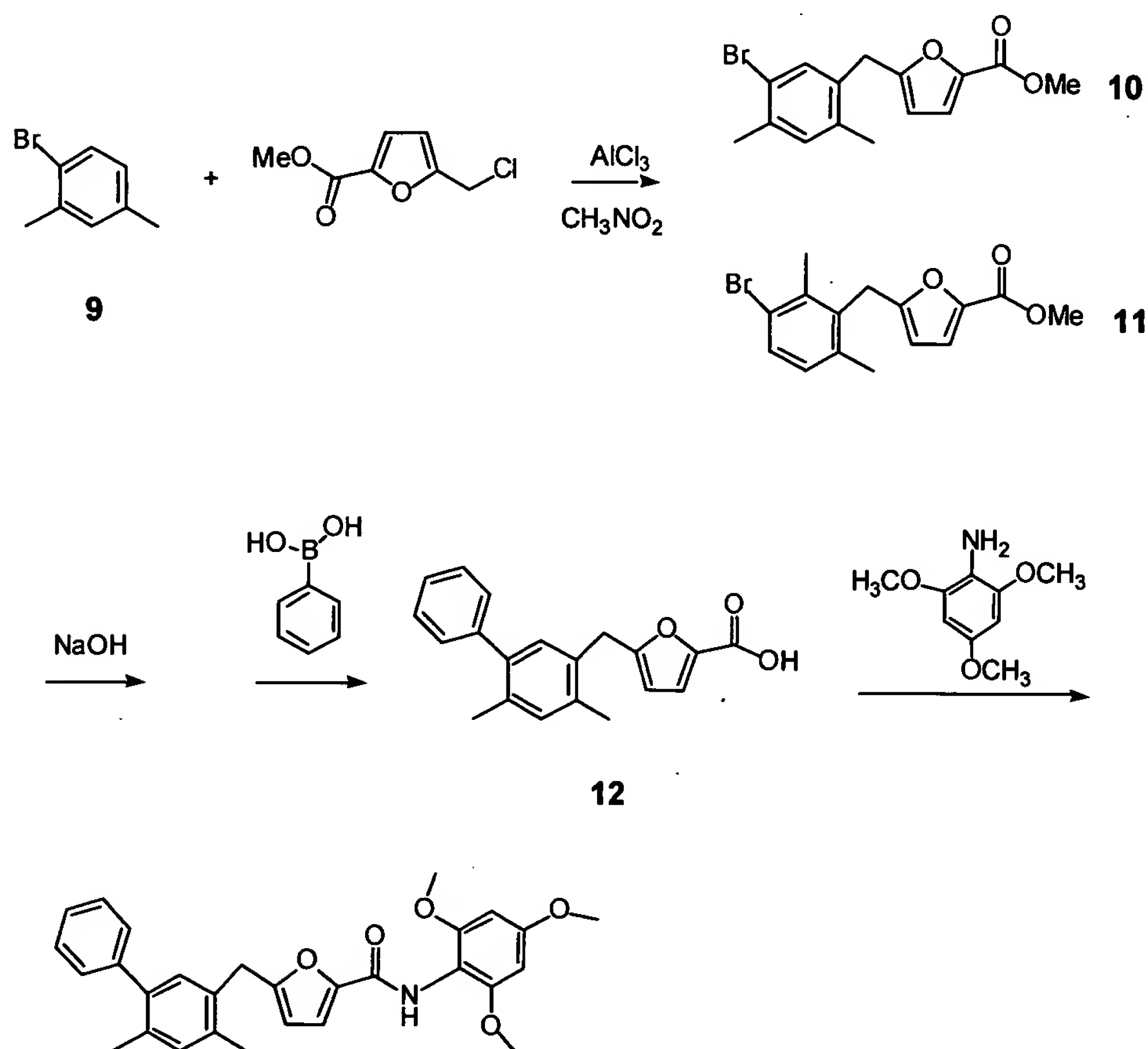
Compound 6 (150 mg, .55 mmol) was treated with thionyl chloride and coupled with trimethoxy aniline to give AXC07485 (124 mg). ^1H NMR (CDCl_3): δ 2.31 (s, 3H), 2.50 (s, 3H), 2.54 (s, 3H), 3.80 (s, 9H), 4.02 (s, 2H), 6.00 (d, 1h), 6.16 (s, 2h), 7.08 (s, 1H), 7.35 (s, 1H), 7.52 (s, 1H), MS (APCI): 438.7 (M+1)

Synthesis of 5-(5-isopropenyl-2, 4-dimethylbenzyl)-N-(2,4,6-trimethoxyphenyl)-2-furamide.



To a solution of compound **6** (250 mg, .92 mmol) in 5 mL dry THF at 0°C under N₂ was added methyl lithium (1.4M in hexanes, 3 eq.). The solution was stirred at 0°C for 3 hours, quench with water and extracted with ethyl acetate. The organic layer was dried over magnesium sulfate and concentrated under reduced pressure. The crude compound was treated with SOCl₂ and coupled with trimethoxy aniline to give AXC 07499 (36 mg). ¹HNMR (CDCl₃): σ 1.80 (s, 3H), 2.06 (s, 3h), 2.11 (s, 3h), 3.59 (s, 9H), 3.75 (s, 2H), 4.60 (d, 1H), 4.95 (d, 1H), 5.81 (d, 1H), 5.95 (s, 2H), 6.71 (s, 1H), 6.79 (s, 1H), 6.89 (d, 1H), 7.18 (s, 1H). MS (APCI): 436.2 (M+1).

10 **Synthesis of 5-[(4,6-dimethyl[1,1'-biphenyl]-3-yl)methyl]-N-(2,4,6-trimethoxyphenyl)-2-furamide.**

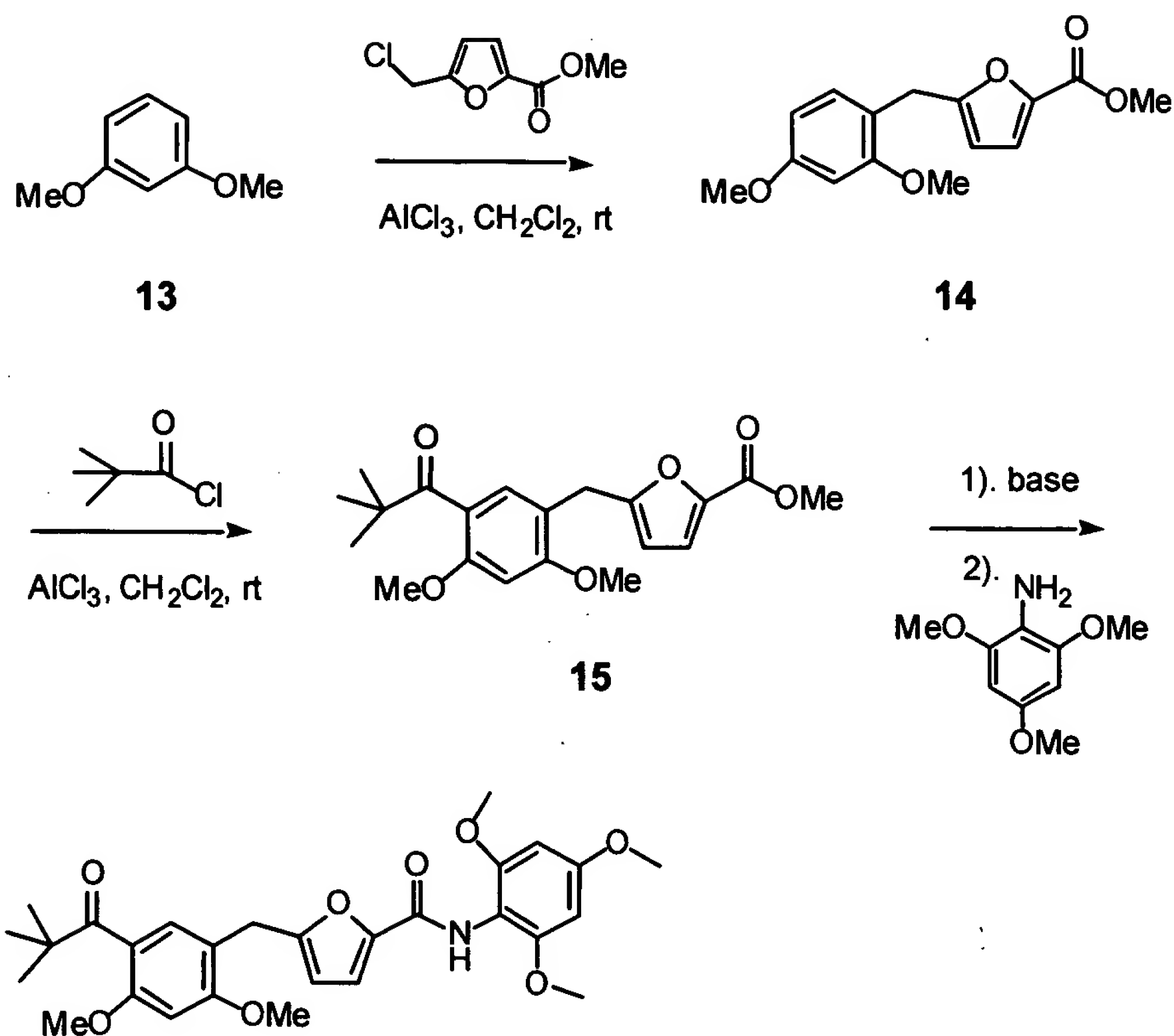


Friedal Crafts reaction of compound **9** (10g, 57.3 mmol), methyl furoate (12.7g, 68.7 mmol) and AlCl₃ (9.1g, 68.7 mmol) was carried out in nitromethane at 80°C for 2 hours. The solution was poured in 200 mL ice water and extracted with ethyl acetate. The organic layer was concentrated and purified by silica gel column eluted with
5 hexane/ethyl acetate (9:1 v/v) to give a mixture of regioisomers **10** and **11** (15.5g) with a ratio of 2:1.

The mixture was hydrolyzed in 2N NaOH/MeOH (1:1 v/v) to give the mixture of acid analogs.

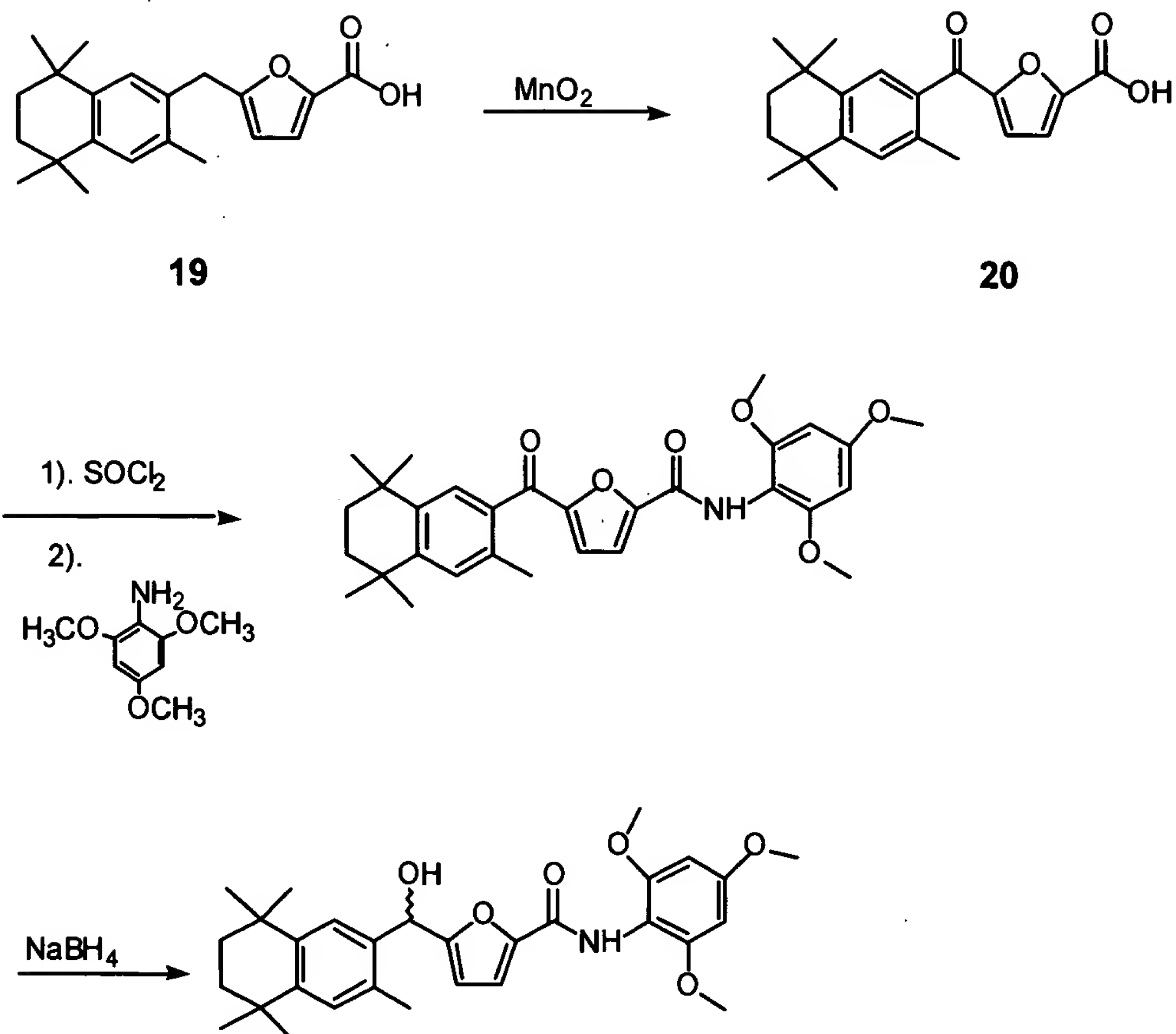
10 The acid mixture (2.3g, 7.4 mmol), benzene boronic acid (1.1g, 8.9 mmol), [P(Ph)₃]₄Pd, and potassium carbonate (2N, 11mL) in DMF (20 mL) was heated to 80°C overnight. After aqueous work up the residue was passed through a silica gel column and eluted with a mixture of solvent hexane/ethyl acetate/acetic acid (7:3:1 v/v/v) to yield a mixture of 2 regioisomers with was recrystallized in hexane and ethyl
15 acetate to give **12** (610 mg). The compound **12** was coupled with trimethoxyaniline through standard procedure to give AXC07468 in good yield. ¹HNMR (CDCl₃): σ 2.24 (d, 3H), 2.33 (d, 3H), 3.82 (s, 6H), 3.84 (s, 3H), 4.02 (s, 2H), 6.03 (d, 1H), 6.18 (s, 2H), 7.06 (s, 2H), 7.12 (s, 1H), 7.29-7.40 (m, 5H). MS (APCI): 472.1 (M+1).

20 **Synthesis of 5-[5-(2,2-dimethylpropanoyl)-2,4-dimethoxybenzyl]-N-(2,4,6-trimethoxyphenyl)-2-furamide**



Compound **15** was prepared in two steps of Friedal Crafts reaction (see general procedure) from compound **13** in moderate yield with good regio selectivity.

- 5 Compound **15** was hydrolyzed and coupled with trimethoxyaniline to give the compound. ¹HNMR (CDCl₃): σ 1.18(s, 9H), 3.79 (s, 3H), 3.81 (s, 6H), 3.85 (s, 6H), 3.93 (s, 2H), 6.03 (d, 1H), 6.15 (s, 2H), 6.45 (s, 1H), 6.86 (s, 1H), 7.11 (s, 1H), 7.40 (s, 1H). Ms (APCI) 512.1 (M+1).
- 10 **Synthesis of 5-[(3,5,5,8,8-pentamethyl-5,6,7,8-tetrahydro-2-naphthalenyl)carbonyl]-N-(2,4,6-trimethoxyphenyl)-2-furamide and 5-[hydroxy(3,5,5,8,8-pentamethyl-5,6,7,8-tetrahydro-2-naphthalenyl)methyl]-N-(2,4,6-trimethoxyphenyl)-2-furamide**



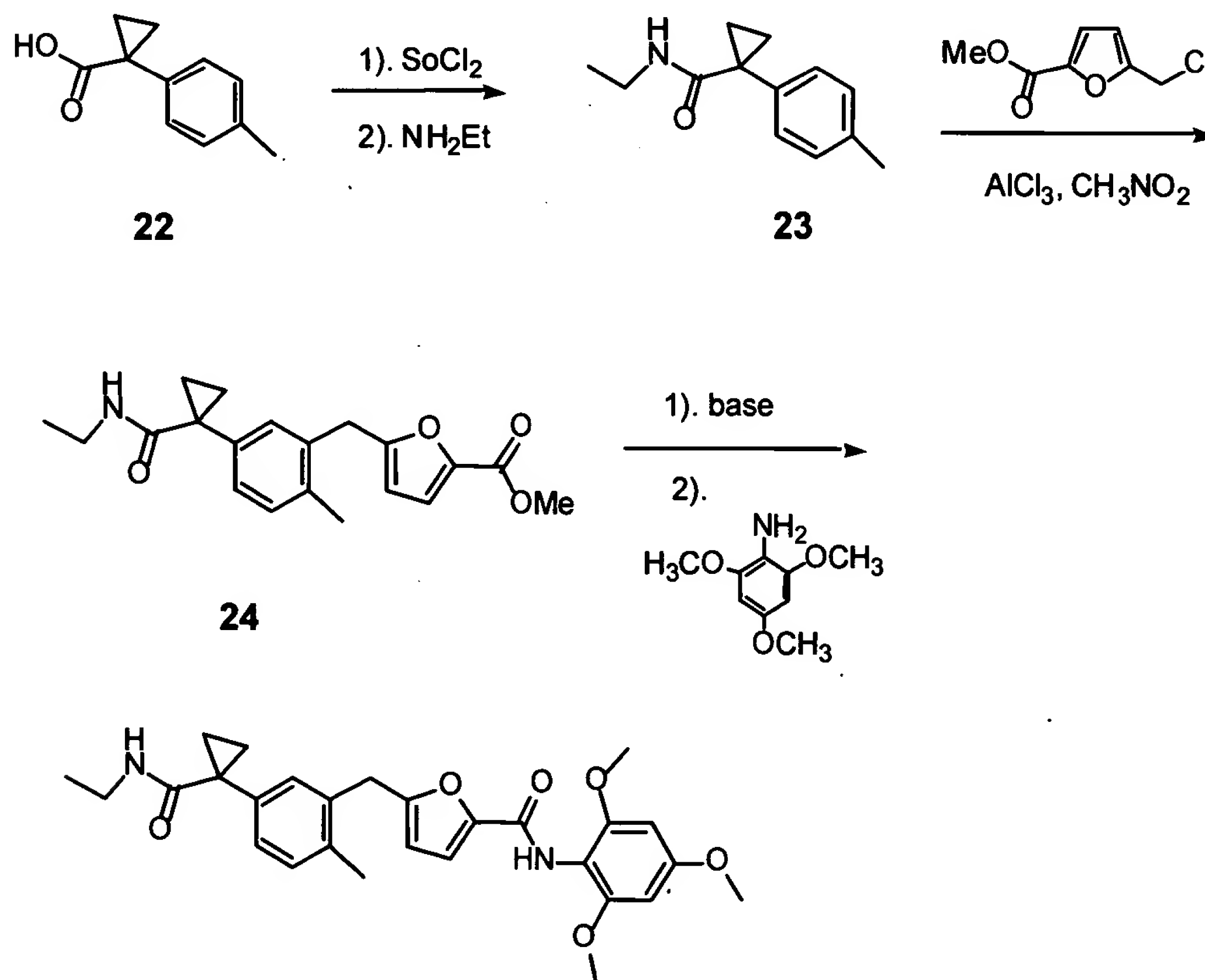
A mixture of compound **19** (9.0g, 27.5 mmol) and MnO_4 (8.2g, 82.7 mmol) in mixture solvent of chloroform and dichloroethane was heated to 70°C overnight. After aqueous workup, compound **20** by silica gel column and eluted with hexane/ethyl acetate/ acetic acid (90:10:1v/v/v). AXC07042 was obtained by the general procedure for amide bond formation. ^1H NMR (CDCl_3): δ 1.26-1.31 (2s, 12H), 1.70 (s, 4H), 3.81 (s, 6H), 3.83 (s, 3H), 6.18 (s, 2H), 7.05 (d, 1H), 7.22 (s, 1H), 7.29 (d, 1H), 7.52 (s, 1H), 7.68 (s, 1H). MS (APCI): 506.2 (M+1).

10

The compound (50mg) was treated with NaBH_4 (1.5eq) in ethanol (2mL) and diethyl ether (0.5mL). The solution was stirred at room temperature for 1.5 hours, quenched with water, and extracted with ethyl acetate. The organic layer was concentrated to

give the compound as a white solid. ^1H NMR (CDCl_3): σ 1.24, 1.28, 1.29 (3s, 12H), 1.66, 1.67 (2s, 4H), 2.29 (s, 3H), 2.55 (d, 1H), 3.80 (s, 6H), 3.82 (s, 3H), 5.98 (d, 1H), 6.17 (s, 2H), 7.08, 7.10 (2s, 2H), 7.38 (s, 1H), 7.41 (s, 1H). MS (APCI): 508.2 (M+1).

5 **Synthesis of 5-(5-{1-[(ethylamino)carbonyl]cyclopropyl}-2-methylbenzyl)-N-(2,4,6-trimethoxyphenyl)-2-furamide**

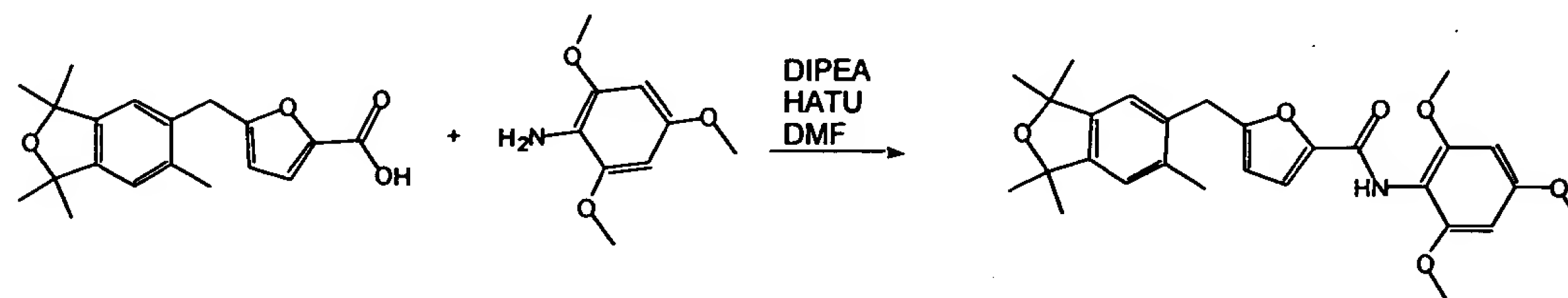
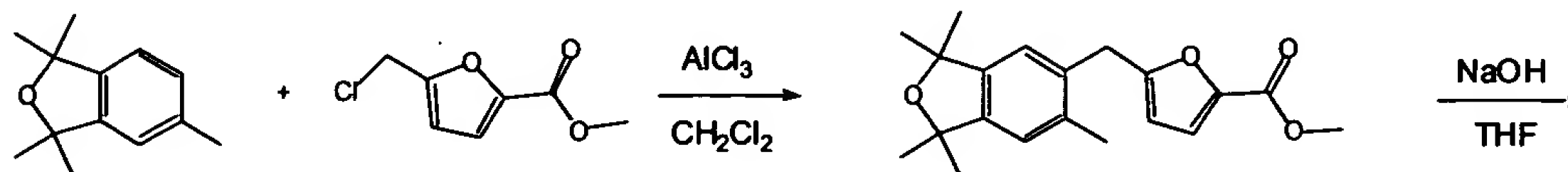
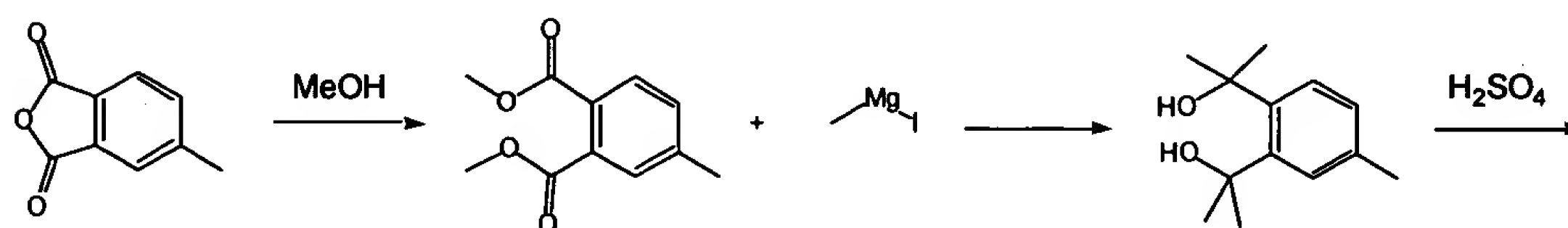
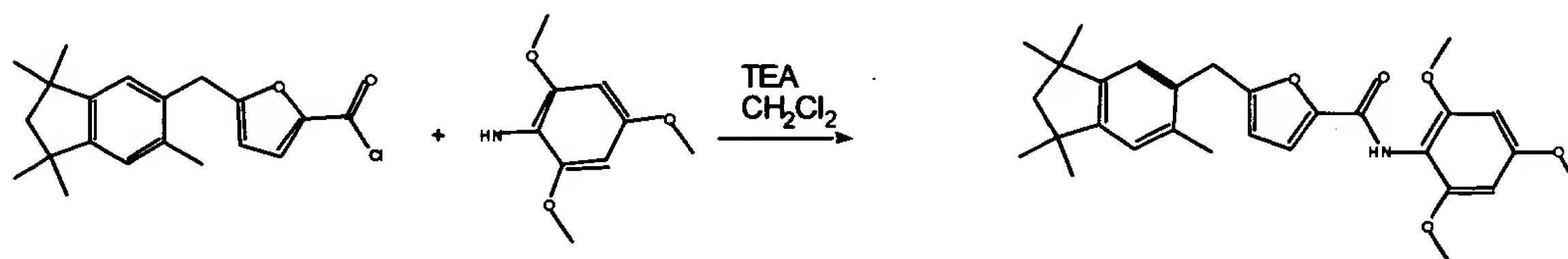
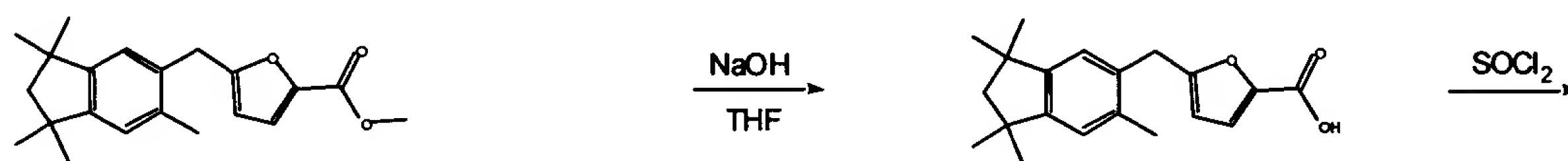
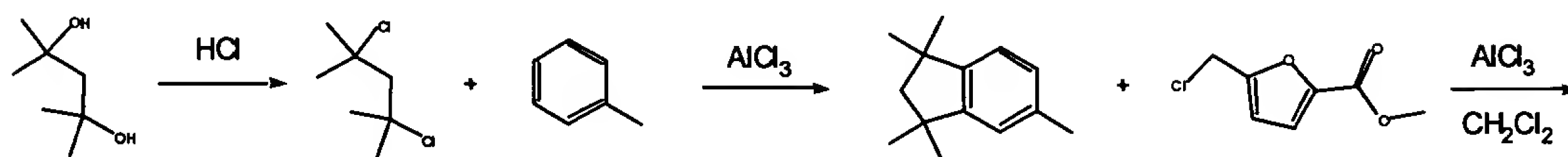


- 10 A solution of **22** (2.0g, 11.3 mmol) in thionyl chloride (8mL) was heated to reflux for 3 minutes. The unreacted thionyl chloride was removed by rotary evaporator. The concentrated residue was dissolved in CH_2Cl_2 . To this solution ethylamine (excess) was added to give compound **23** (1.3g). The compound was converted to AXC07555 by 3 steps (Friedel Crafts reaction, hydrolysis, and amide bond formation) as described
- 15 in general procedures. ^1H NMR (CDCl_3): σ 0.94-1.02 (m, 5H), 1.59 (m, 2H), 2.34 (s, .

3H), 3.17 (q, 2h), 3.80, 3.81 (2s, 9H), 4.01 (s, 2H), 5.39 (brd, 1H), 6.01 (d, 1H), 6.17
9s, 2H), 7.11 (d, 1H), 7.21 (m, 2H), 7.31 9s, 1H). MS (APCI): 493.2 (M+1).

Additional Examples:

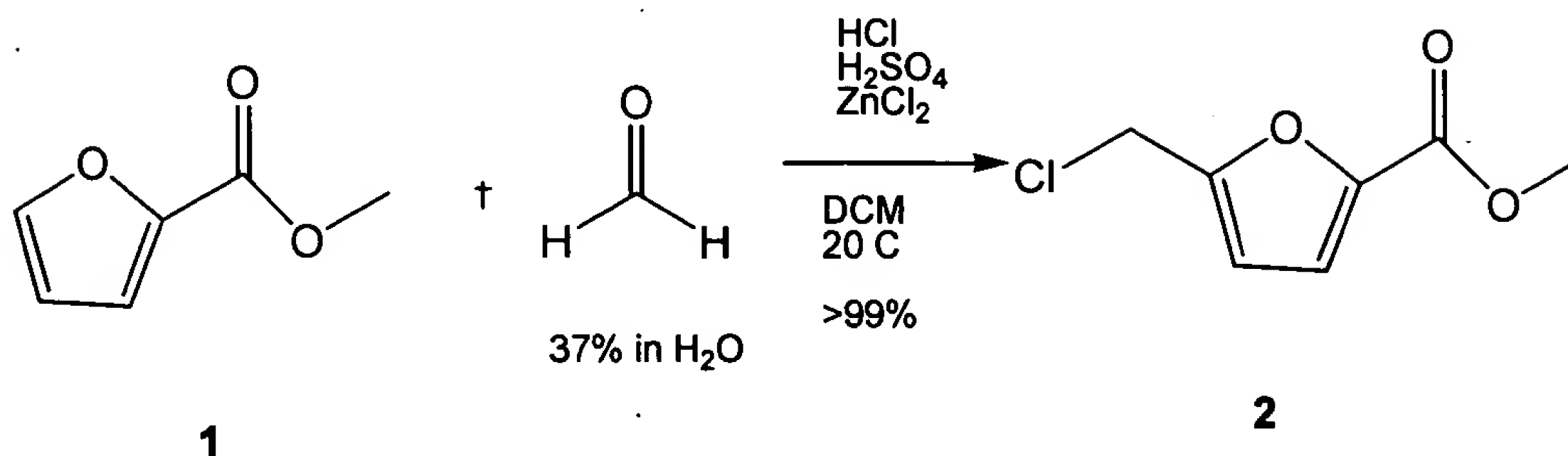
5



10

A useful intermediate can be prepared as follows:

Preparation of methyl-5-(chloromethyl)-2-furoate



5

Required Materials

Materials	Conc.	Molecular Weight	Amount	Number of Moles	Number of Equivalents	Density	CAS
Methyl 2-furoate	97%	126.11	0.371 kg	2.88	1	1.179	[611-13-2]
Formaldehyde (aq)	37%	30.03	0.371 L	4.95	1.72	1.083	[50-00-0]
HCl	37%	36.46	2.2 L	22.3	7.74	1.19	[7647-01-0]
H ₂ SO ₄	95-98%	98.08	1.1 L	11	3.82	1.84	[7664-93-9]
Dichloromethane	Reagent	84.93	2.2 L	-	-	1.325	[75-09-2]
ZnCl ₂	98%	136.28	0.520 kg	3.7	1.3	-	[7646-85-7]
Dichloromethane	Reagent	84.93	*****L	-	-	1.325	[75-09-2]
NaHCO _{3(aq)}	Saturated	-	*****L	-	-	-	[144-55-8]
Silica Gel	-	-	0.600 kg	-	-	-	-
Na ₂ SO ₄	98%	-	as necessary	-	-	-	[7757-82-6]

Itemized Procedure for a 2.88 Mole Scale Batch of Methyl 5-(chloro-methyl)-2-furoate (2)

- A 12L 3-neck round bottom flask (r.b.) is equipped with an addition funnel; over-head electric stirrer; thermocouple; and an ice bath. (a nitrogen blanket is recommended, but may not necessarily be required)
- Charge 2.2 L DCM to r.b. followed by 2.2 L conc. Hcl. (no significant exothermic reaction observed; two layers formed)
- Begin agitation. (ensure adequate mixing of both phases)
- Charge 1.1 L H₂SO₄ to the addition funnel as space permits. Cool reactor to 0-10°C. Begin addition of sulfuric acid drop-wise initially until the exothermic reaction subsides (approximately 1/2 addition), and then increase addition rate to a light stream. (maintain temperature below 20°C for safety purposes)
- Change cooling bath to water at room temperature. (this will act as a heat-sink for the subsequent additions without slowing reaction rates significantly)
- Charge 0.371 kg methyl 2-furoate to r.b. in one portion. (no the exothermic reaction noticed, green/brown solution)
- Charge 0.520 kg ZnCl₂ in many portions. (some bubbling - suspect HCl gas; the exothermic reaction controlled by water bath)
- Charge 0.371 L formaldehyde to rinsed addition funnel. Add to reactor over 2.5 - 3.5 hours. (temperature is maintained at room temperature by water bath; slow addition results in fewer polymerization reactions between 'free' formaldehyde)
- Stir overnight at room temperature.
- When reaction is complete by TLC (see below) drop aqueous layer. Filter organic layer through a 0.550 kg silica plug (dry packed, approx. 10 cm thick). Elute with approximately 4 L DCM until no further product comes off a syndicated by TLC. (ensure this step is performed with proper ventilation, as some acid vapors are still present, even in the filtrate)
- Concentrate to give an oil ranging in color from yellow to brown (approximately 0.5 L)

- Charge an equal amount DCM, about 0.5 L. Wash with 2 x 0.2 L distilled water. Then wash organic layer with 0.05 L sat NaHCO_3 in 0.15 L distilled water. (ensure pH 7-10, no significant product lost in aqueous layer)
- Drop aqueous layer, and dry organic with Na_2SO_4 . Filter through 0.05 kg silica (about 5 cm thick). Elute with DCM until no further product comes off.
- Concentrate by rotovap using house vacuum. Then place oil on high vacuum pump overnight. (yellow to light brown oil)

Yield range: 95-100%

Purity range: 95-98% (HPLC A%)

Further Comments

Visual: The reaction is followed by TLC (254nm) using 30% EtOAc/Hexanes (r.f. starting material - 0.52, r.f. product - 0.40).

HPLC: TFA method (method and spectra attached). Starting material rt = 12.67 min., product rt = 17.37 min.

NMR: ^1H (CDCl_3) (spectra attached) 3.93 (s, 3H), 4.63 (s, 2H), 6.52 (d, 1H), 7.18 (d, 1H).

If the reaction has not gone to completion, 10% of the original volume of formaldehyde can be added every 4 hours. Temperature effects have not been extensively studied, but it has been convenient to maintain pot temperatures in the ranges specified for each step. Acid vapors present in the organic layer even after
5 filtration make handling outside of the fume hood problematic, so care should be taken when transferring material outside of the hood until the pH is neutralized. If the addition rate of formaldehyde is increased, polymer formation will prevent the reaction from going to completion due to the consumption of formaldehyde. Arrangements with the safety department should be made prior to the start of this process so that
10 arrangements to accommodate the large volumes of acid being discharged in the aqueous layer can be made. Neutralization of the aqueous acid layer requires large volumes of base, produces a considerable exotherm, and requires a prolonged addition period, and is therefore not recommended. The final product should be kept cool, as no stability data of this product is available. Keeping the product in a Nalgene container at
15 -20°C causes the product to crystallize, slightly darkening the material, but appears to have no effect on later reactions using this material.

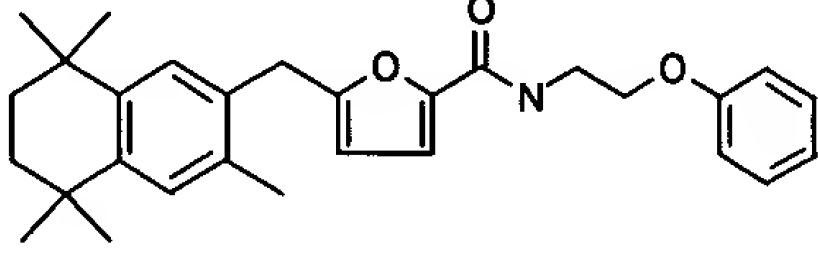
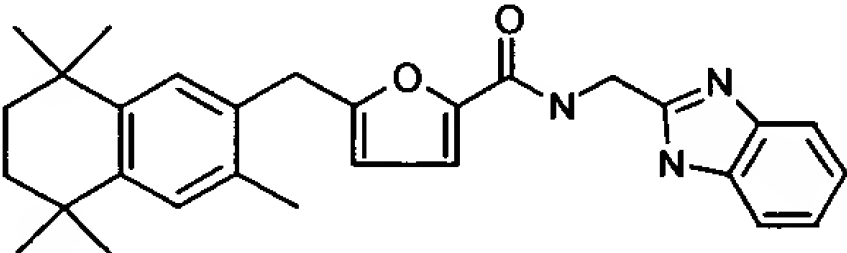
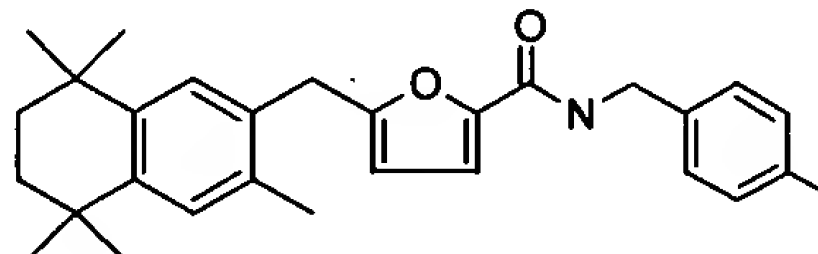
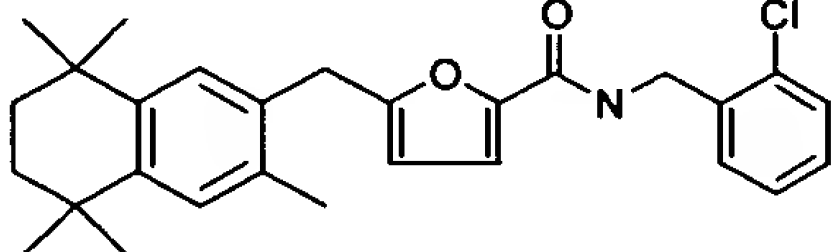
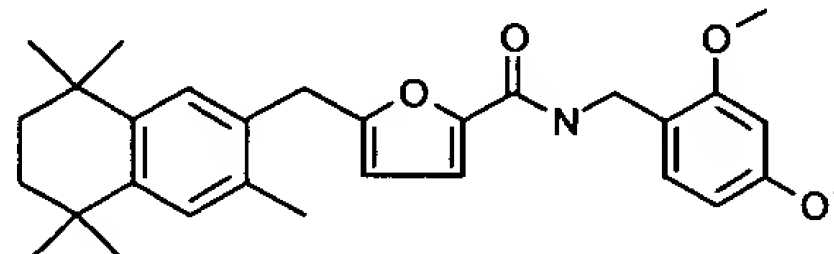
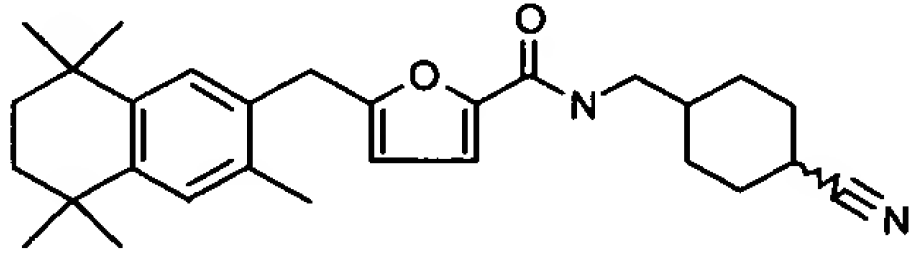
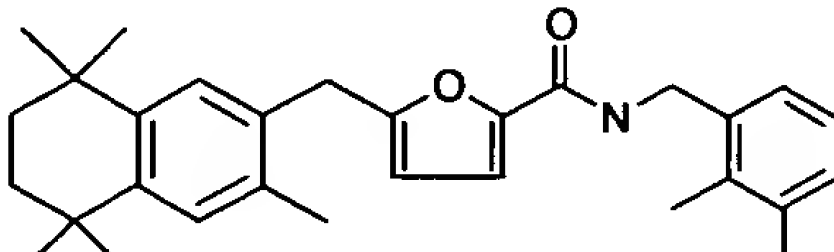
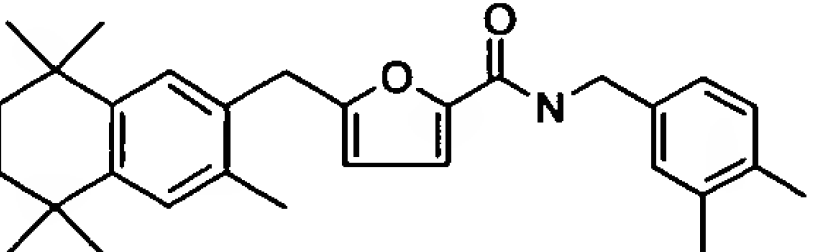
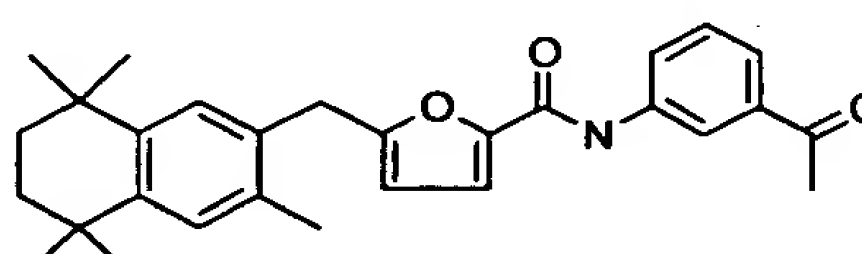
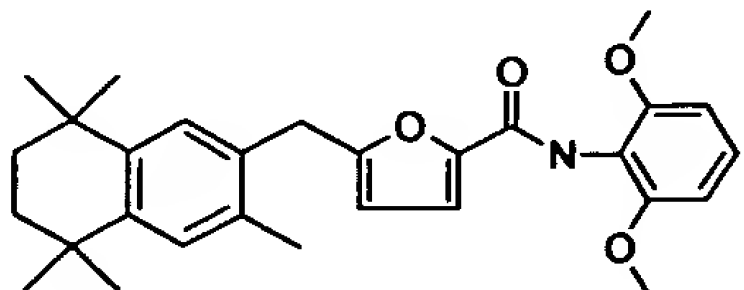
The compounds of the present invention should be useful in treating:

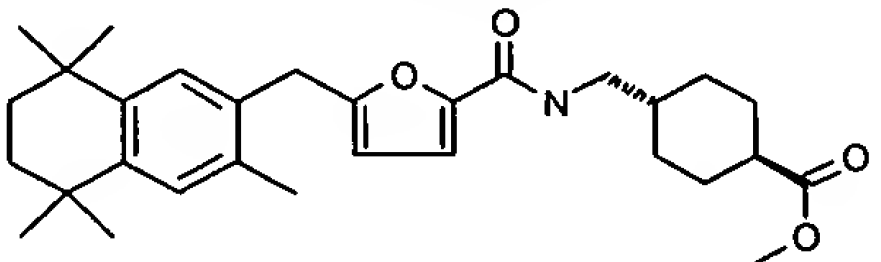
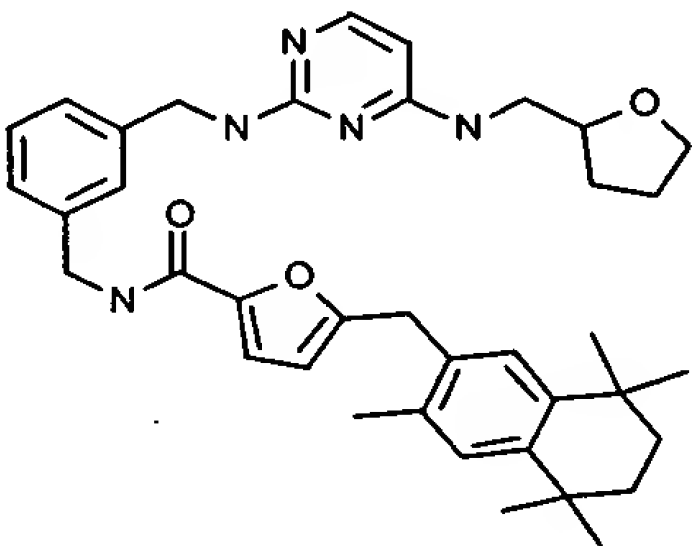
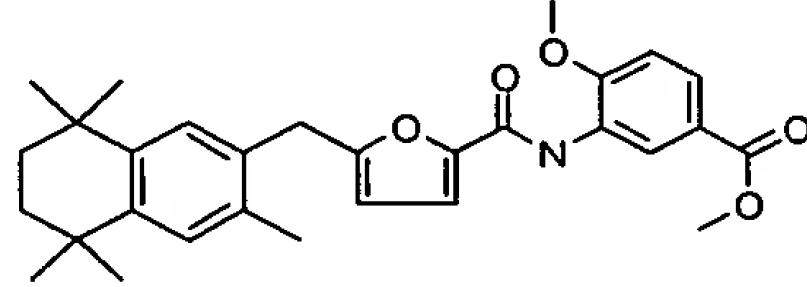
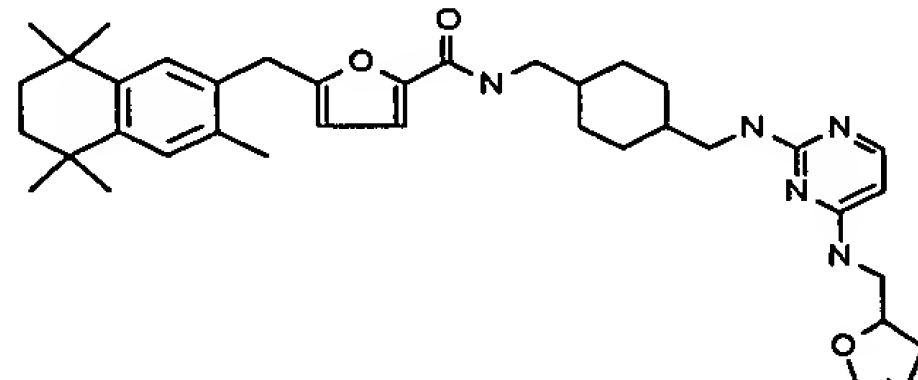
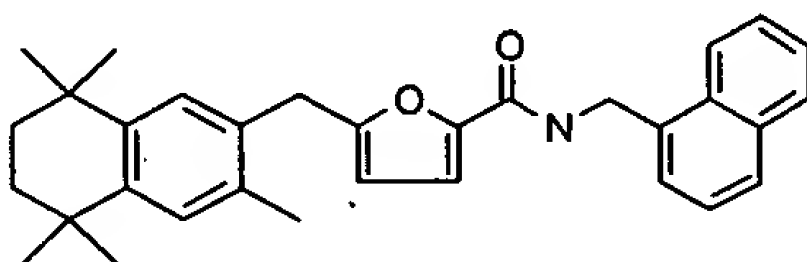
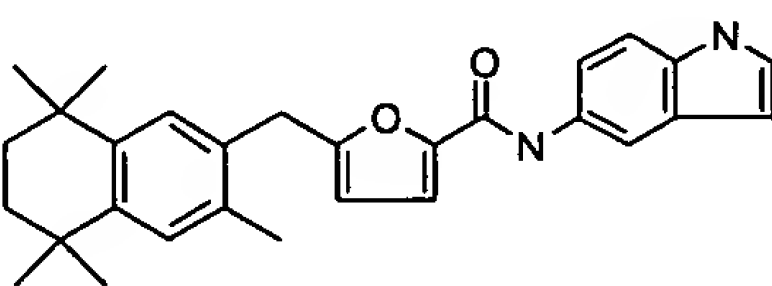
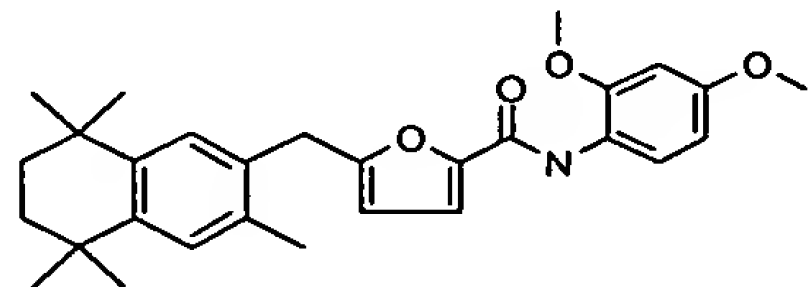
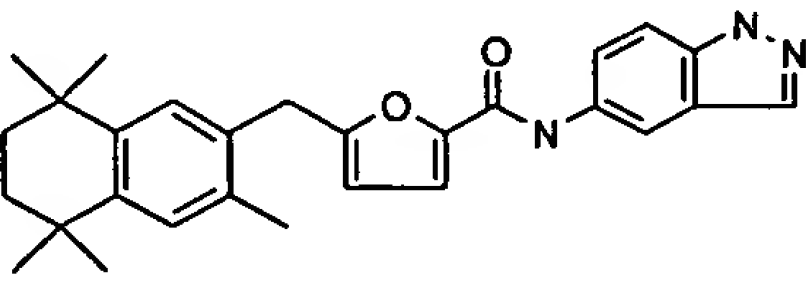
1. hormone dependent cancers/tumors
2. hormone independent cancers/tumors by direct interactions
- 20 3. use in other mechanisms of action

It is believed that Applicants' invention includes many other embodiments which are not herein specifically described, accordingly this disclosure should not be read as being limited to the foregoing examples or preferred embodiments.

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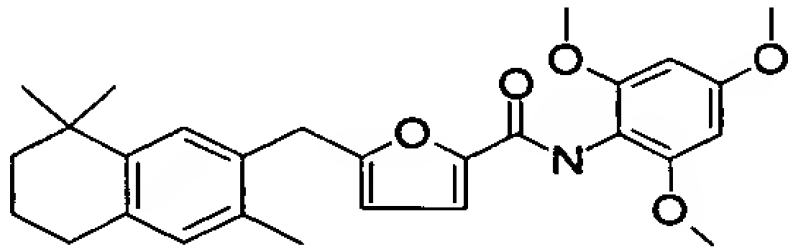
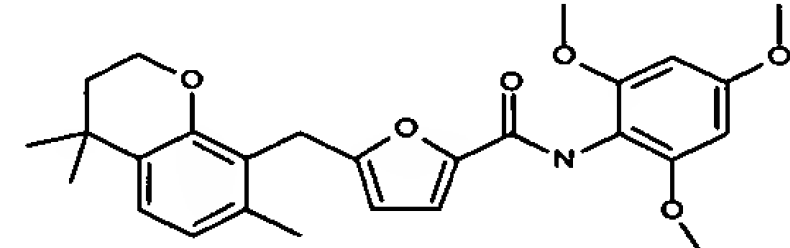
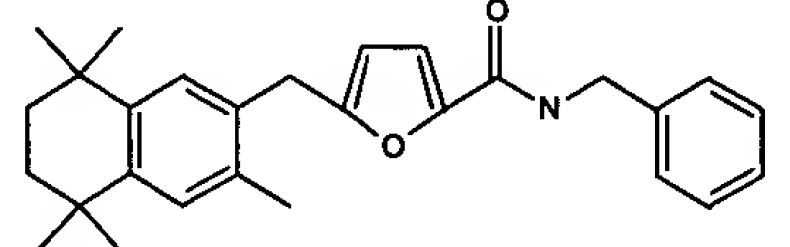
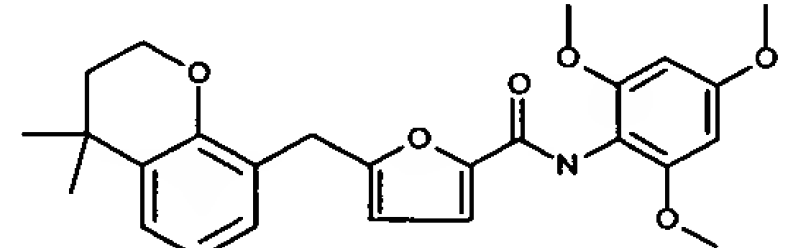
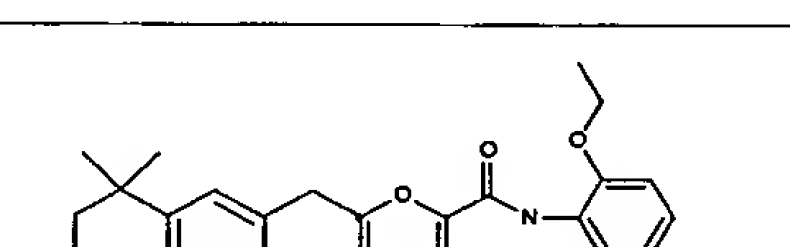
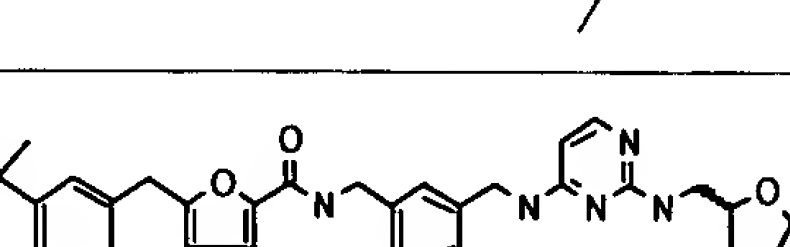
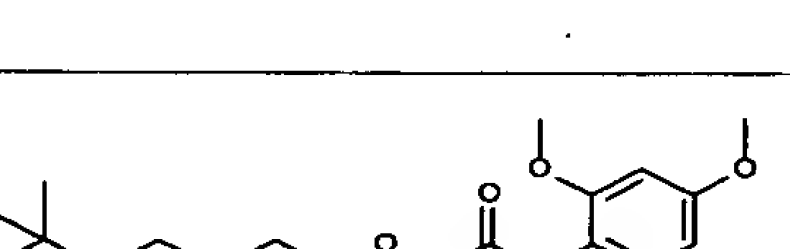
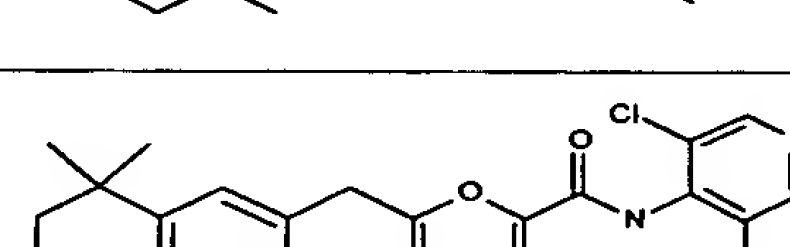
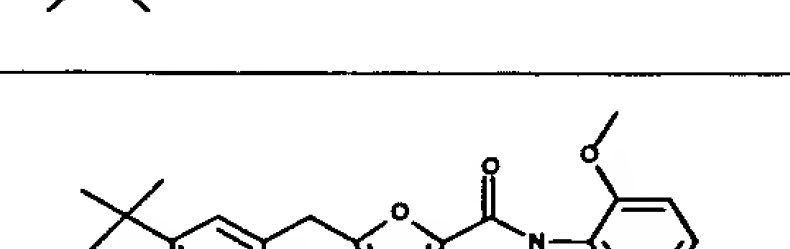
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23		263	141	48 min			
12		418	450	36 min			
13		462	1236	1.5 hr			
38		2776					
41		185	1606				
42		361	1387	58 min			
44		459	1759	55 min			
46		485	1781	55 min			

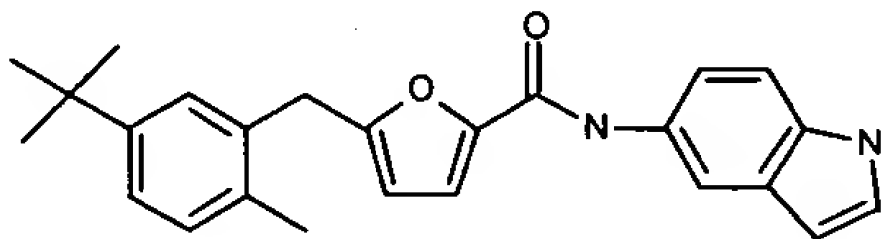
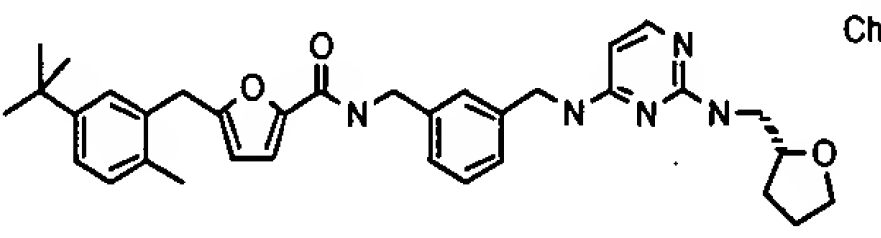
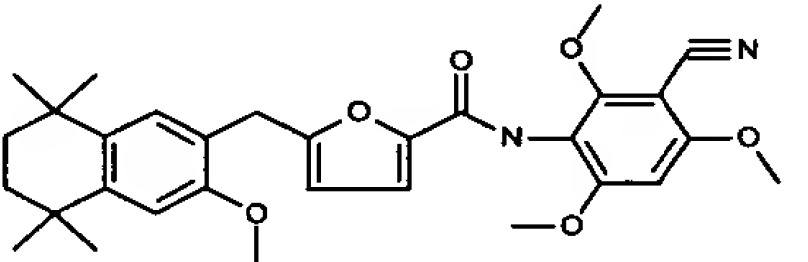
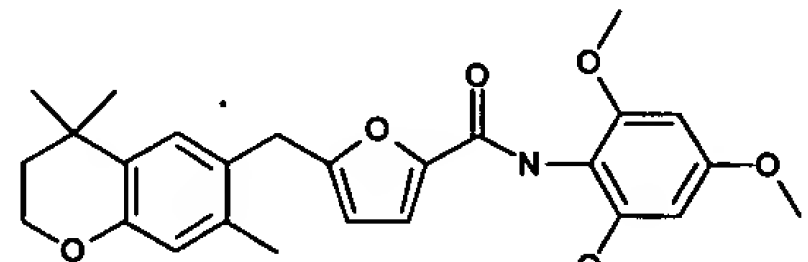
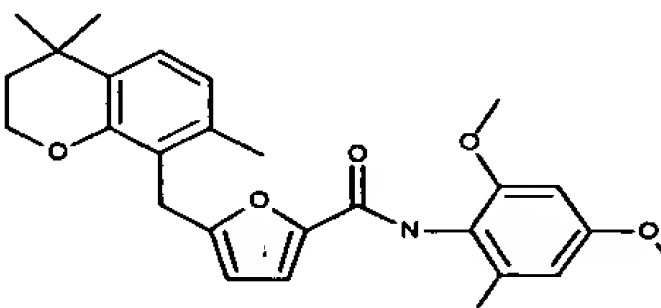
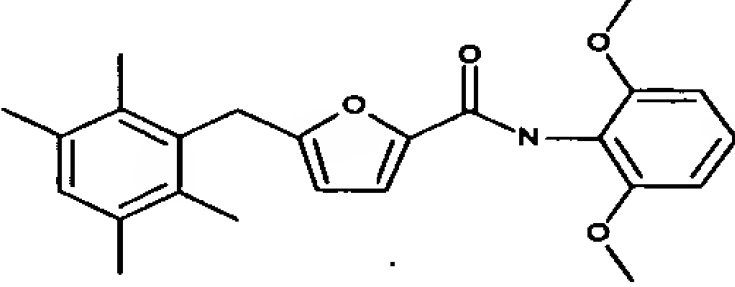
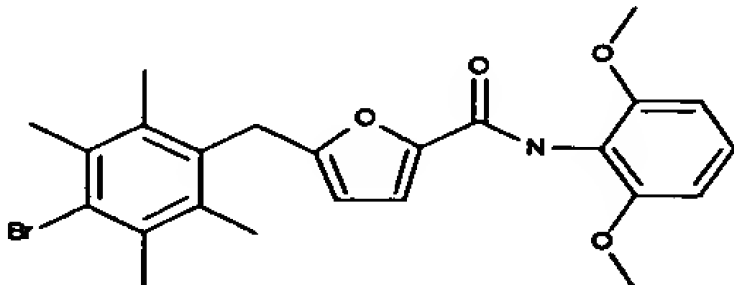
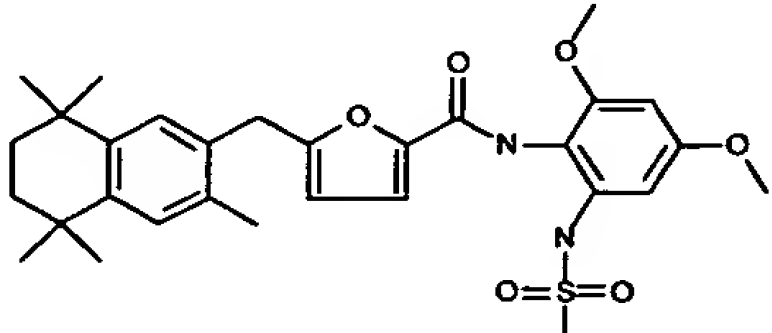
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		1 uM	1 uM		uM		
47		392	2198	1.5 hr			
48		289	2869	1.1 hr	0.33	1.5 hr	4% (20 mg/kg) ¹
14		329	1626	1.4 hr	1.6	1 hr	(10 mg/kg) ⁸
51		392	1877				
56		392	3412	2.6 hr			
16		691	2602	40 min			
17		699	2282	2.2 hr			
18		320	1648	55 min			
19		457	4510	43 min			
20		620	3908	52 min	0.2	1 hr	3% (20 mg/kg) ¹³

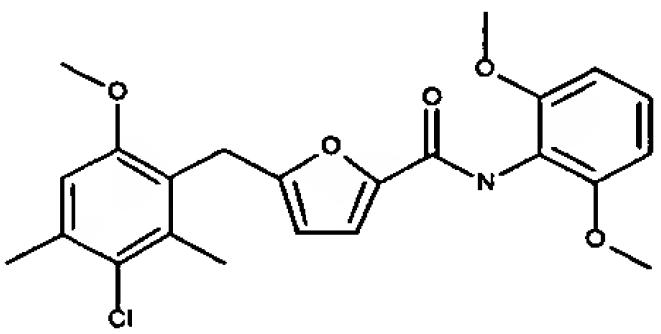
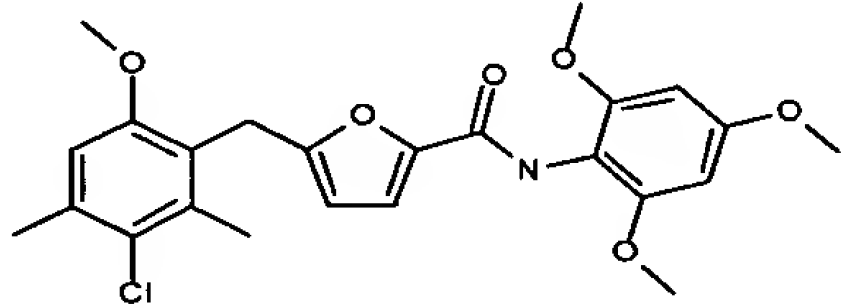
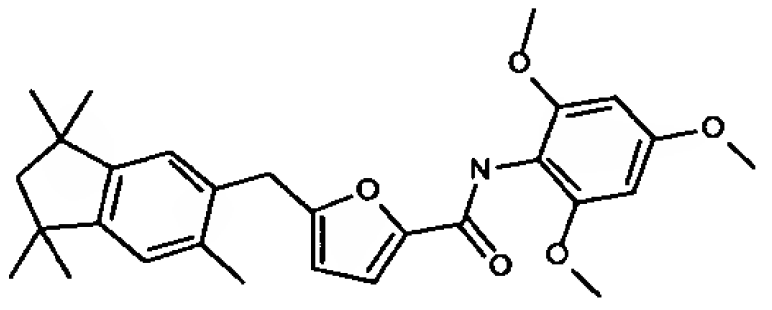
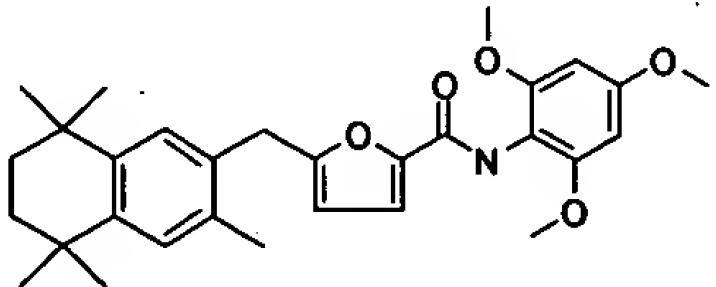
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		1 uM	1 uM		uM	
68		2716	8125			
70		311	2393	58 min		
72		278	4226			
92		241	1282			
95		283	3356			
96		182	1692			
99		353	2549			
109		130	1055			

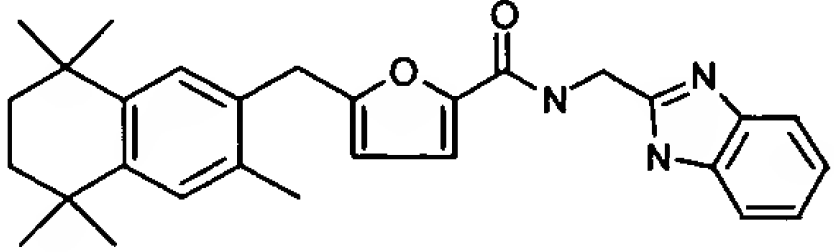
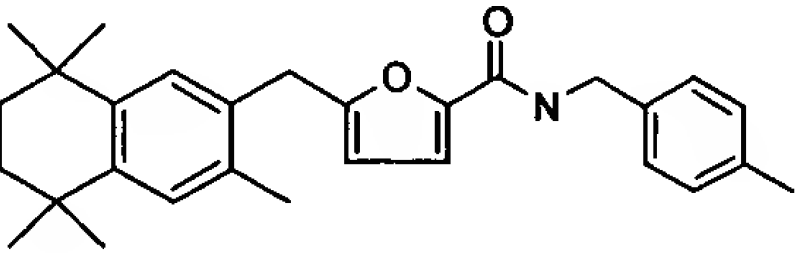
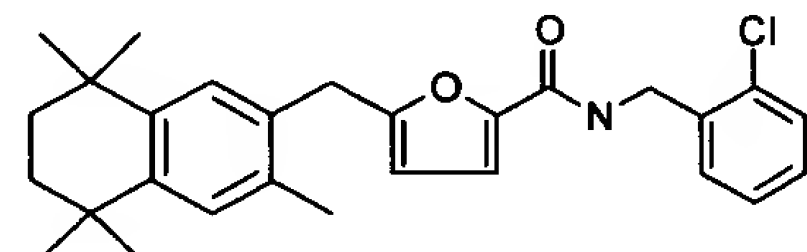
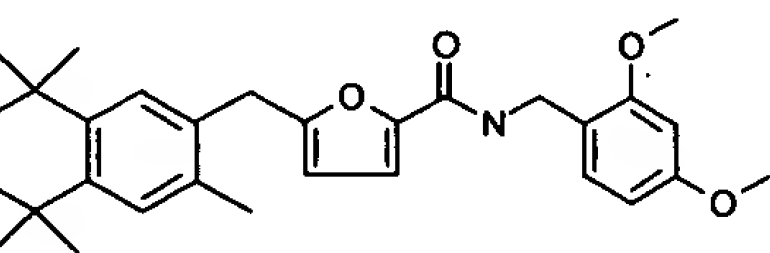
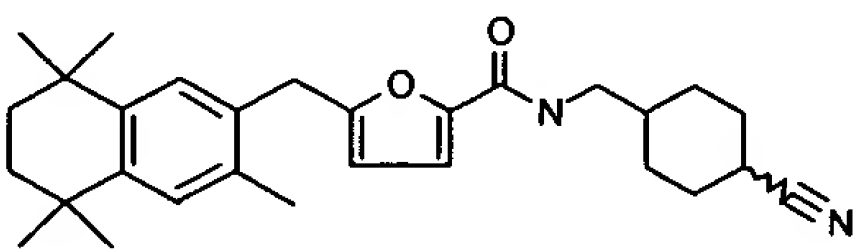
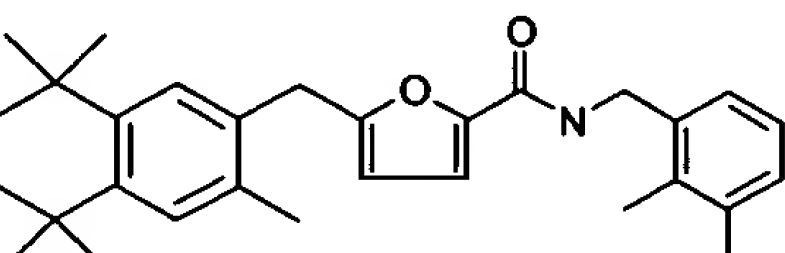
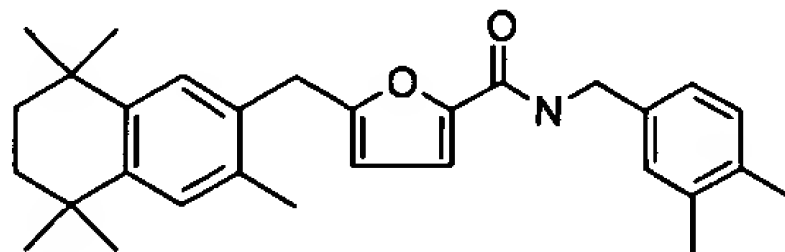
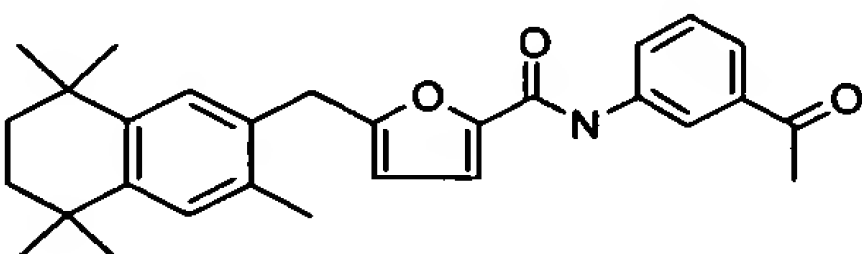
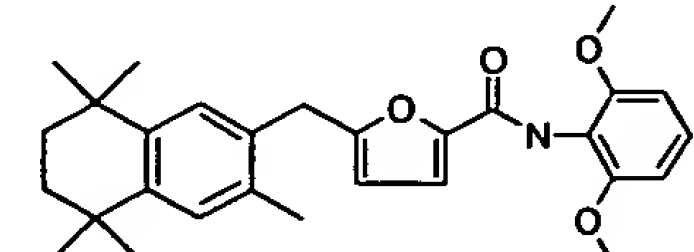
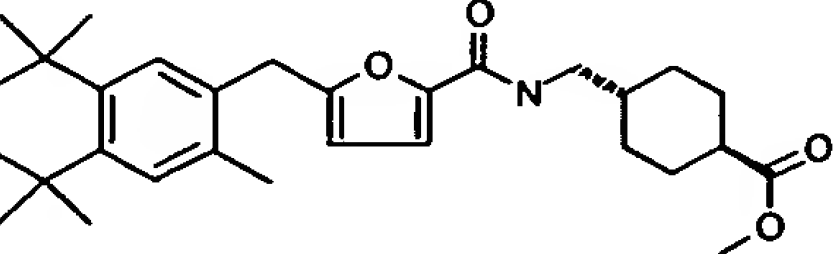
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		1 uM	1 uM		uM		
126		544					
134		630	3909	1.4 hr	0.61	1 hr	8% (20 mg/kg) ⁵
136		191	4437	2 hr	0.24	2 hr	6% (20 mg/kg) ⁴
137		594					
140		497	2521				
147		523	3524	1 hr			
148		248	1414	1.1 hr	0.03	2 hr	3% (20 mg/kg) ¹
150		655	3334				
151		225	3043				

		Human	Rat male				
		pmol/min/ mg	pmol/min/mg	T _{1/2}	C _{max}	T _{max}	F _{p.o.}
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156		867	2995				
159		793	6642				
168		407	1087	1.8 hr	0.3	1 hr	1% (20 mg/kg) ⁶
169		589	3320	1.5 hr	0.97	1 hr	27% (20 mg/kg) ¹
170				toxic			0 (20 mg/kg) ⁷
176	 Chiral	273	2341	2.8 hr			
177		876	2930	2.2 hr			0 (20 mg/kg) ⁹
181		514	1589	1.1 hr	2.43	2.5 hr2	8% (20 mg/kg) ¹
182		326					

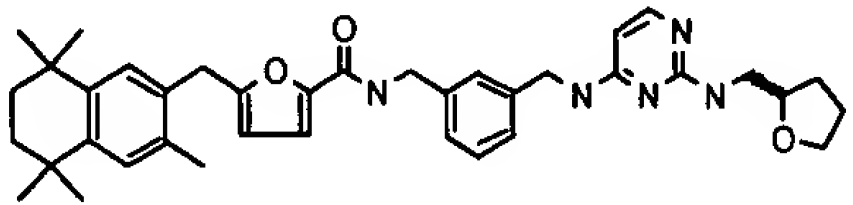
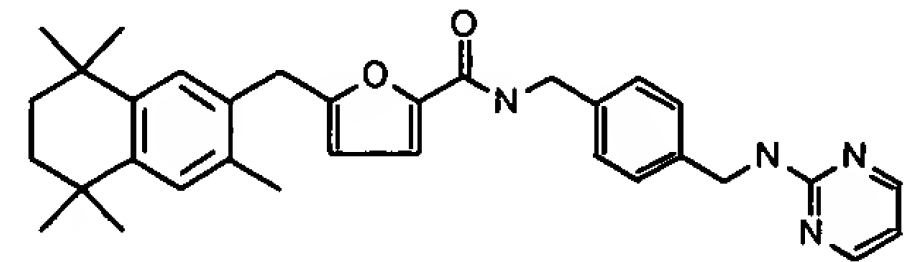
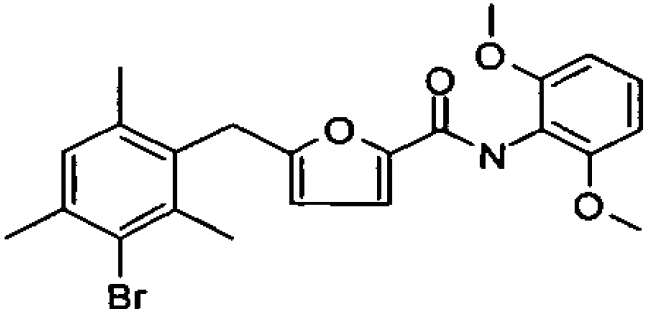
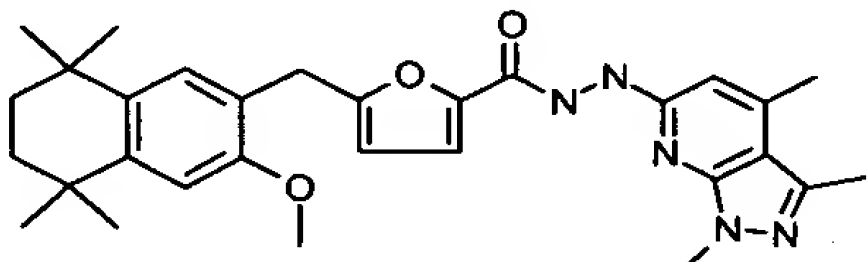
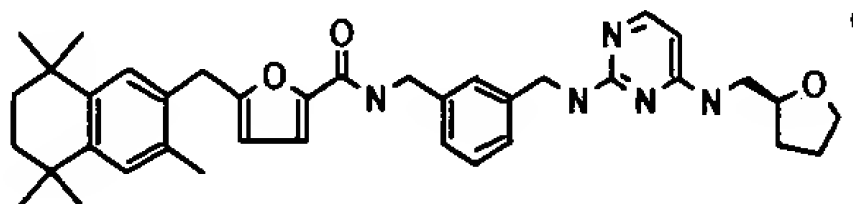
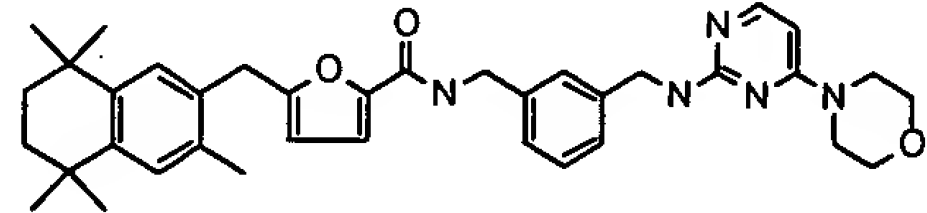
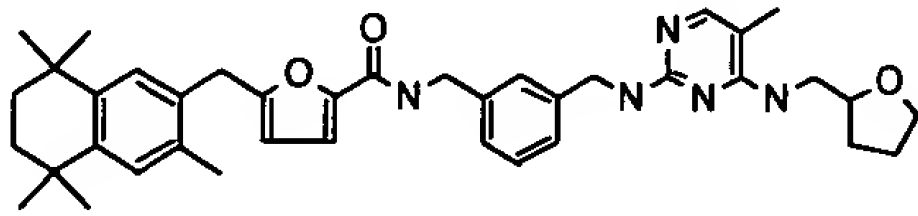
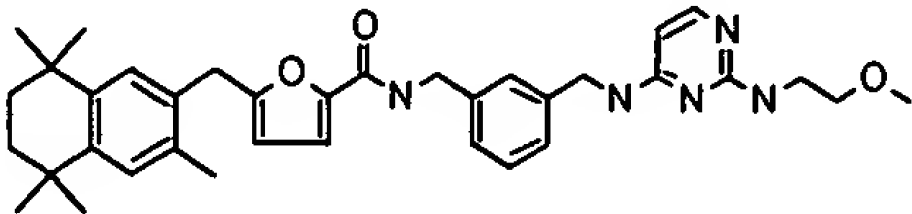
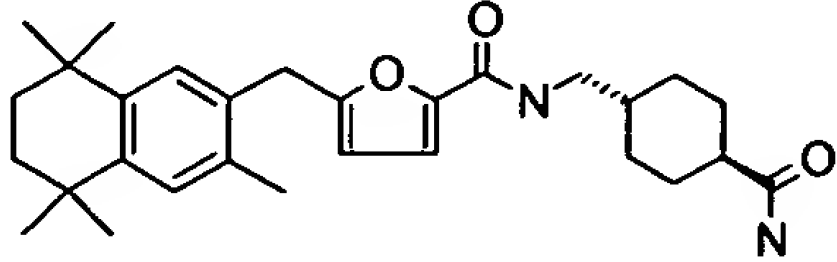
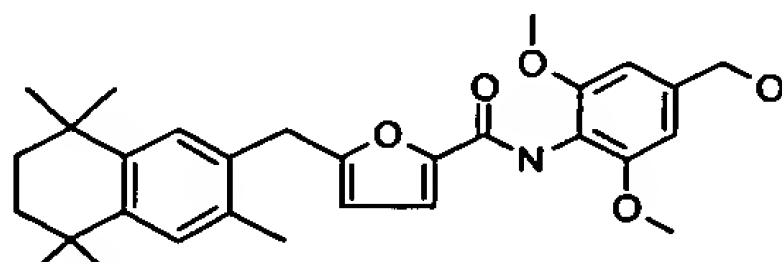
No.	Compound	Human	Rat male				F _{p.o.}
		pmol/min/ mg	pmol/min/mg	T _{1/2}	C _{max}	T _{max}	
		1 uM	1 uM		uM		
183		238	1246	4.1 hr	0.35	1 hr	29% (10 mg/kg) ¹⁰
188		524					
185		646	3713				
190.5		1124					
194		750	5978				
199		181	3387				
200		346	595				
201		479					
206		689	1794				

No.	Compound	Human	Rat male			
		pmol/min/ mg	pmol/min/mg	T _{1/2}	C _{max}	T _{max}
		1 uM	1 uM		uM	F _{p.o.}
207		157	1236			
212	 Chiral	322	3282			
217		483	953			
221		116	528			
222		115	575			
225		450	2180			
226		369	990			
228		641	320			

		Human	Rat male				
		pmol/min/ mg	pmol/min/mg	T _{1/2}	C _{max}	T _{max}	F _{p.o.}
No.	Compound	1 uM	1 uM		uM		
233		773	2988				
299							
300							
Castrated male rats:							
No.	Compound	T _{1/2}	C _{max}	T _{max}	F _{p.o.}		
134		1.7 hr	1.98	1.5 hr	23% (20 mg/kg) ¹		
¹ 10 mg/ml solution in 10% DMSO 10% cremophor 80% saline							
³ 10 mg/ml solution in 10% DMSO 10% cremophor 80% saline (when given as 40 mg/ml in PEG400 or PG, F _{p.o.} was							
⁴ 10 mg/ml solution in 10% DMSO 10% cremophor 80% saline; or 40 mg/ml in PEG400 (T _{max} =4hr); or PG (T _{max} =3.5hr							
⁵ 10 mg/ml solution in 10% DMSO 10% cremophor 80% saline; or 40 mg/ml in PEG400 (C _{max} =0.23 plateau 2-4 hr)							
⁶ 40 mg/ml solution in 50% DMSO 50% cremophor							
⁷ 10 mg/ml suspension in 10% DMSO 20% cremophor 70% saline							
⁸ 5 mg/ml suspension in 10% DMSO 10% cremophor 80% saline							
⁹ 10 mg/ml suspension in 10% DMSO 10% cremophor 80% saline							
¹⁰ 5 mg/ml solution in 10% DMSO 10% cremophor 80% saline							
Bioavailability after i.p. dosing:							
134	38% (20 mg/kg)						
169	70% (20 mg/kg)						
177	8% (20 mg/kg)						
Portal vein infusion:							
136	36% bioavailable (20 mg/kg)						
Metabolism by CYP3A4:							
20	3.4 pmol/min/pmol						

No.	Compound	Rat female					Dog	Monkey
		pmol/min/ mg	T _{1/2}	C _{max}	T _{max}	F _{p.o.}	pmol/min/ mg	pmol/min/ mg
		1 uM		uM			1 uM	1 uM
48		135						
14								
51								
56								
16								
17								
18								
19								
20							2520	7366
68								

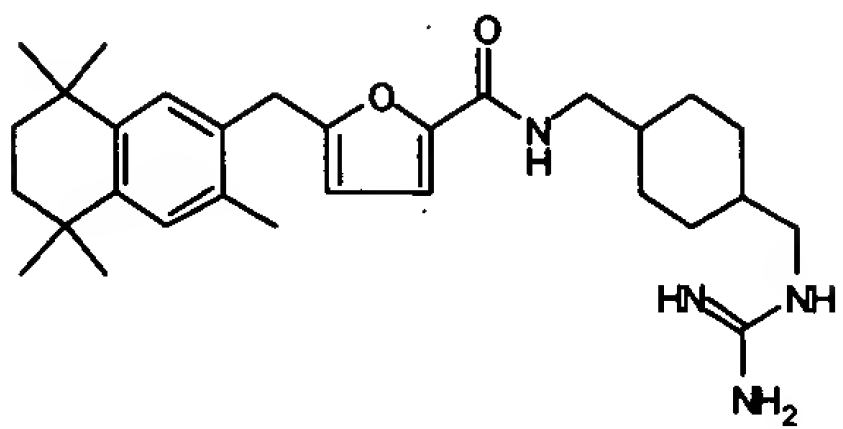
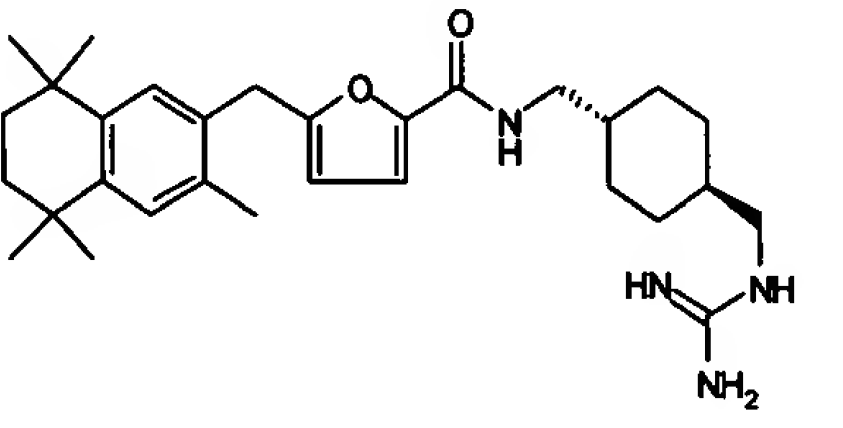
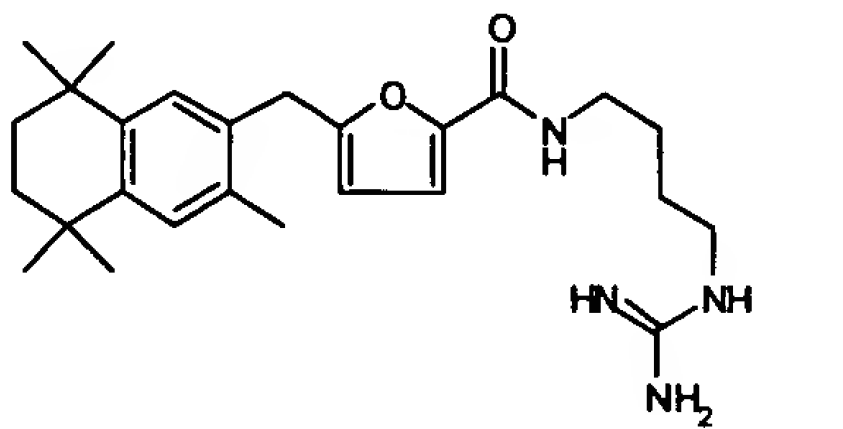
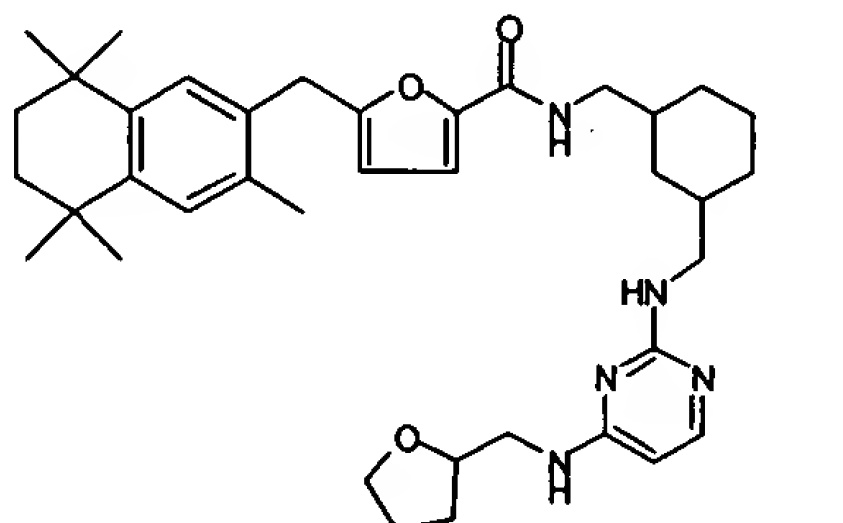
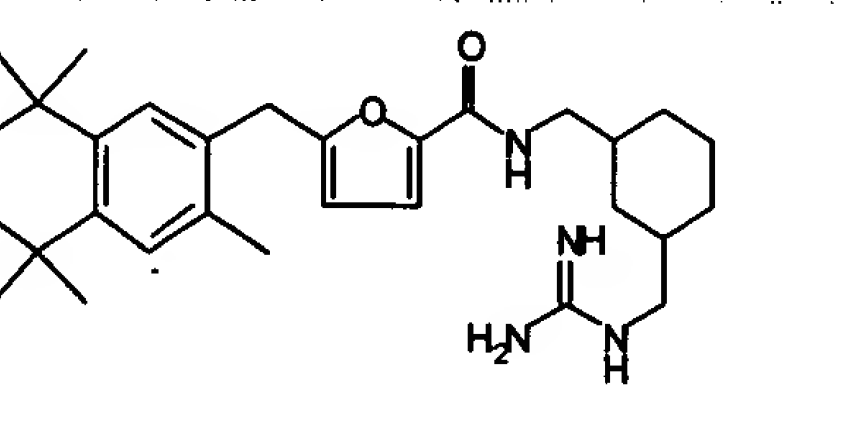
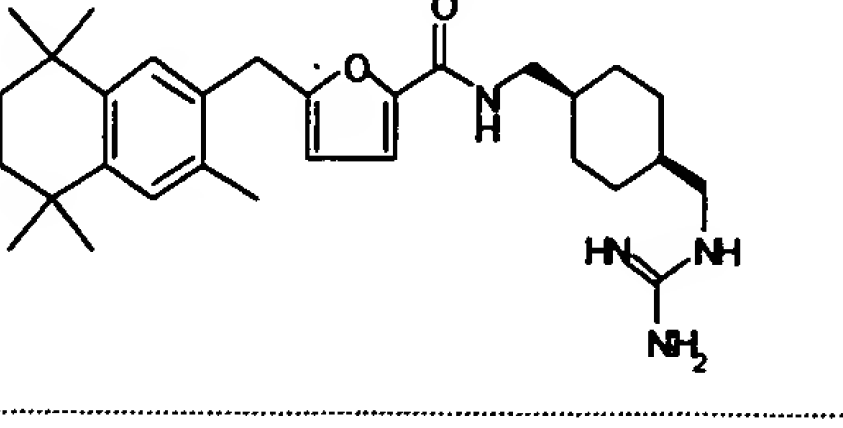
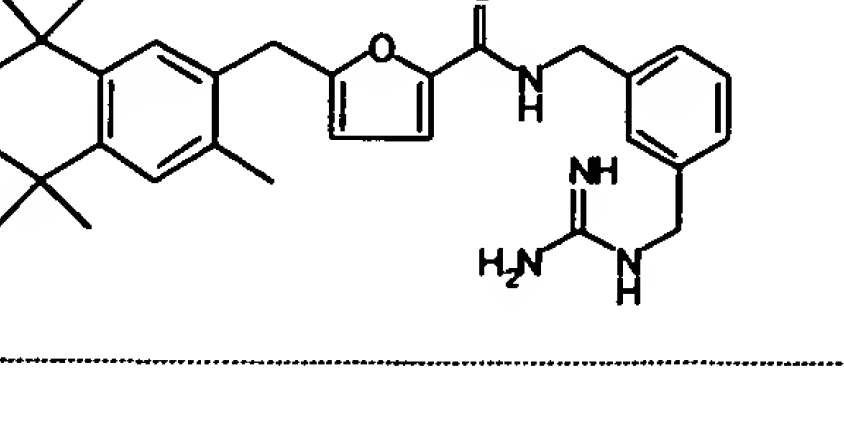
No.	Compound	Rat female				Dog	Monkey
		pmol/min/ mg	T _{1/2}	C _{max}	T _{max}	F _{p.o.}	pmol/min/ mg
		1 uM		uM			1 uM
70							
72							
92							
95							
96							
99							
109							
126			1.9 hr	0.02	30 min	2% (10 mg/kg) ¹⁰	
134		94	1.7 hr	2.31	1 hr	24% (20 mg/kg) ¹	2159
							9277

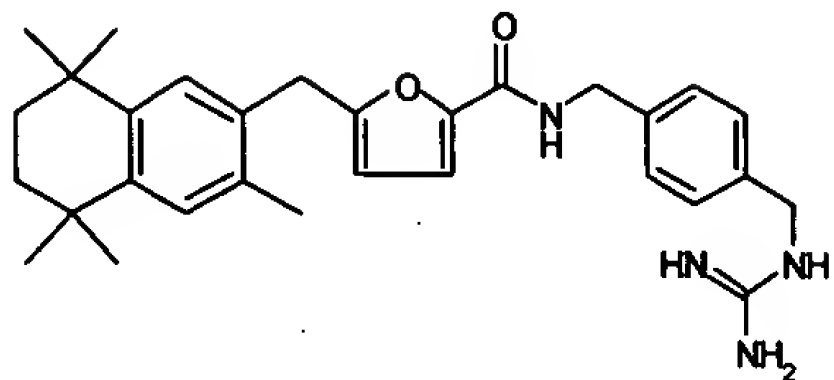
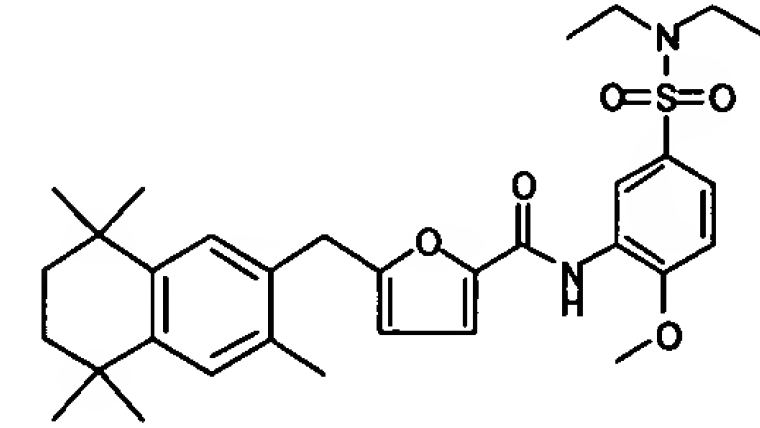
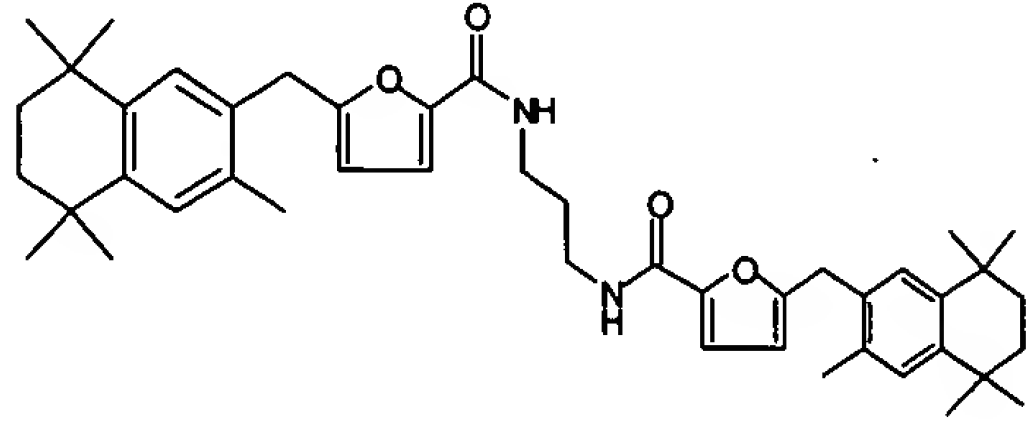
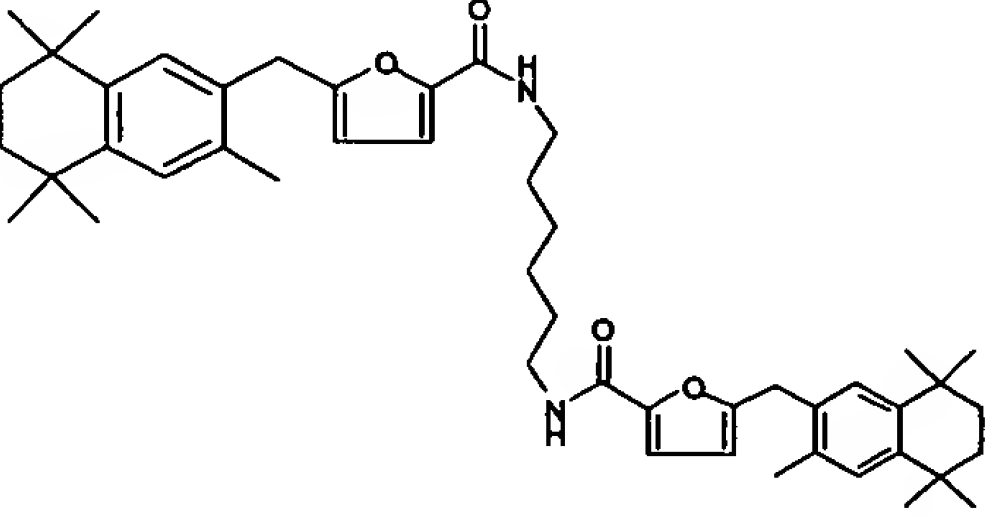
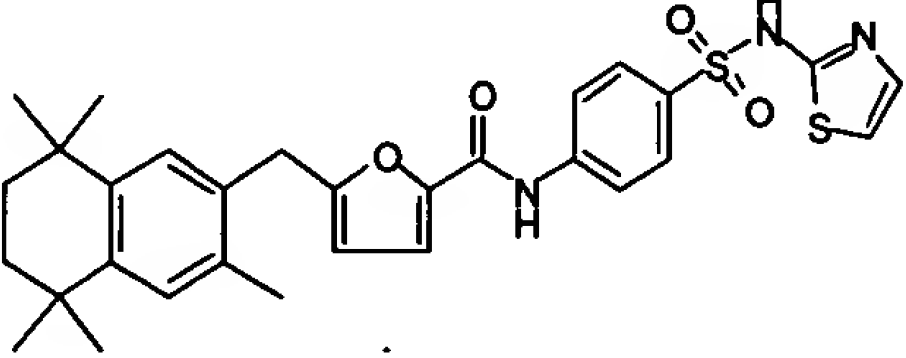
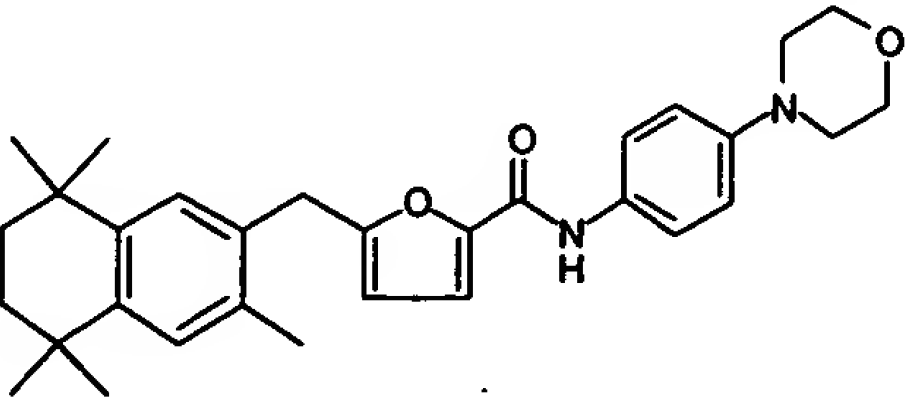
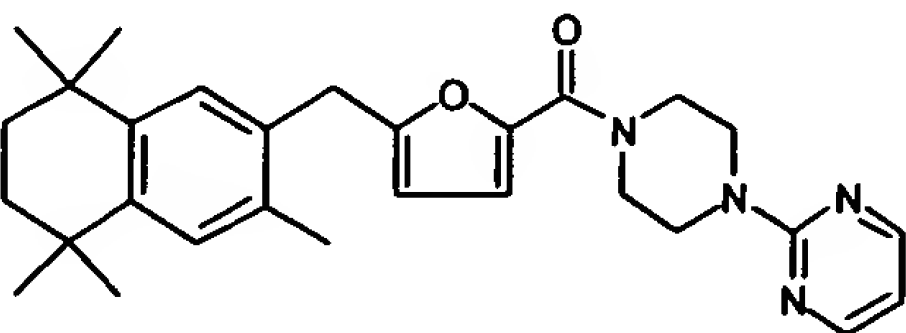
No.	Compound	Rat female				Dog	Monkey
		pmol/min/ mg	T _{1/2}	C _{max}	T _{max}	F _{p.o.}	pmol/min/ mg
		1 uM		uM			1 uM
136	 Chiral	271	3.2 hr	0.24	2.6 hr	8% (20 mg/kg) ¹	3071
137		182	7.9 hr	0.23	1 hr	1% (10 mg/kg) ¹⁰	
140			4.6 hr	0.44	1 hr	59% (10 mg/kg) ¹⁰	
147							
148	 Chiral	133					1960
150							
151							
154							
156							
159							

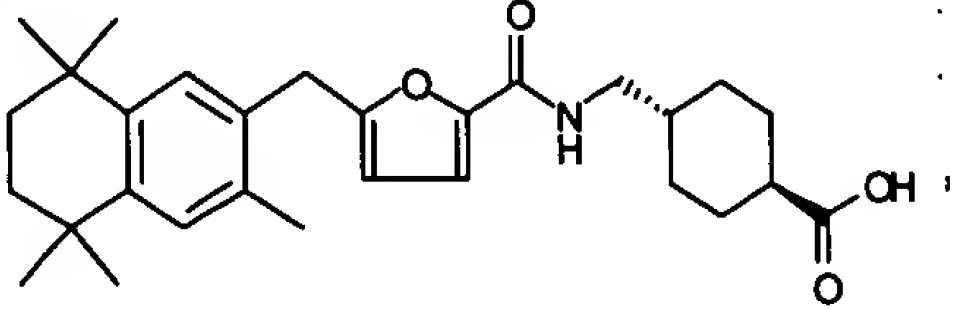
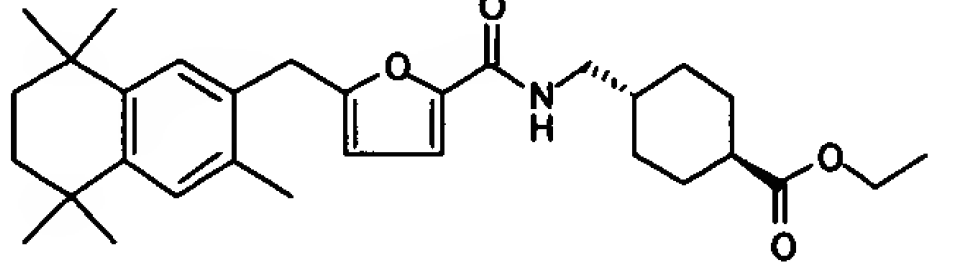
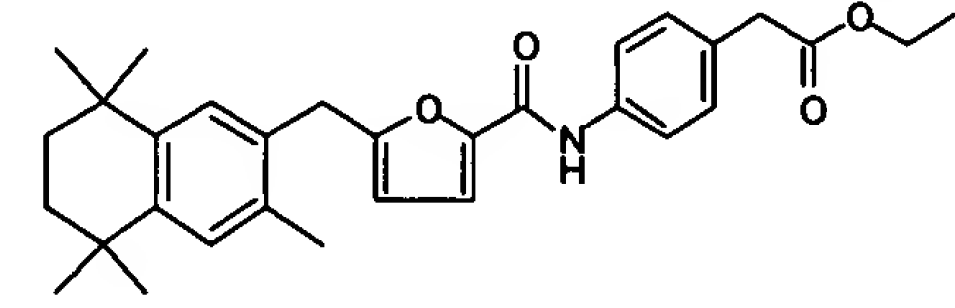
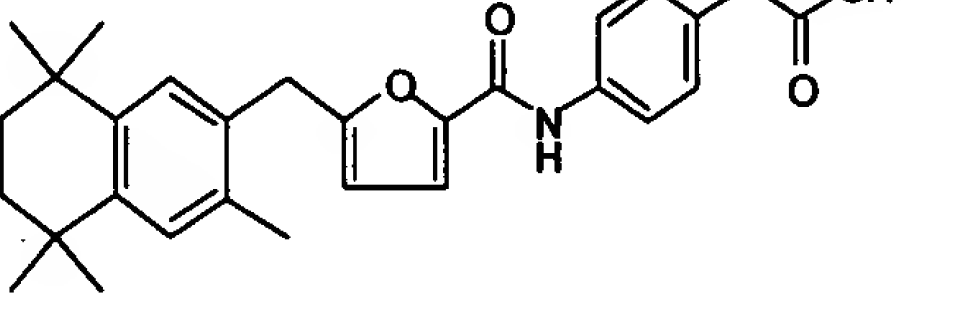
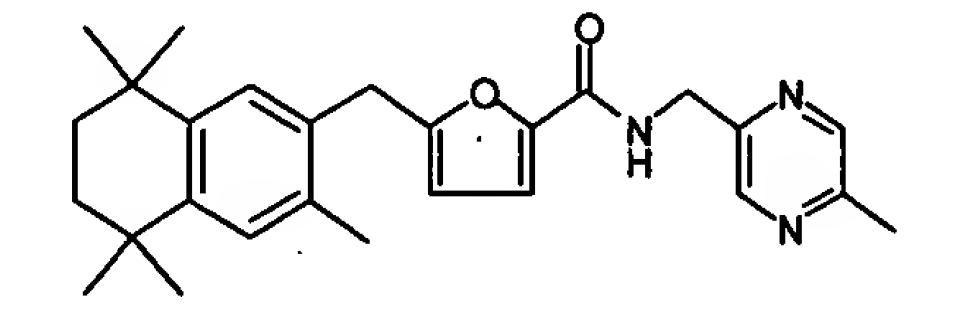
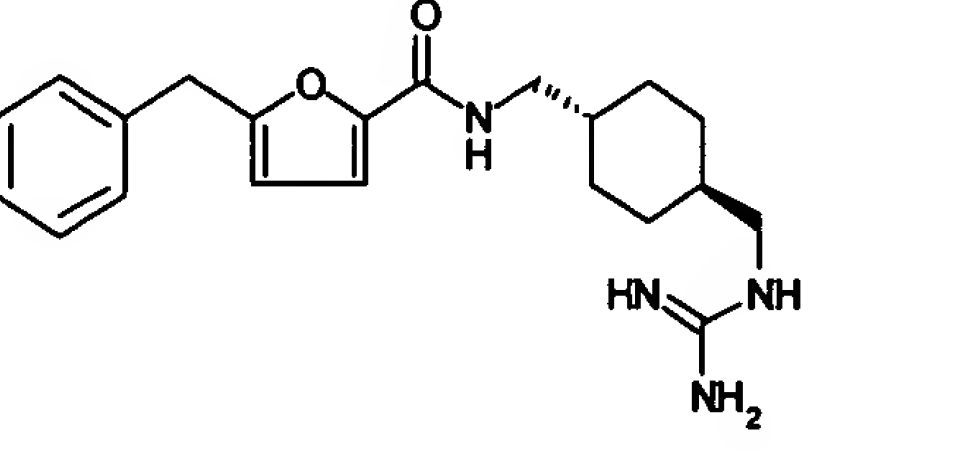
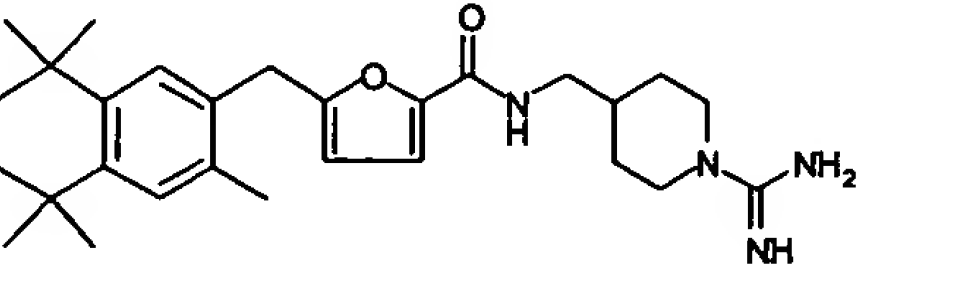
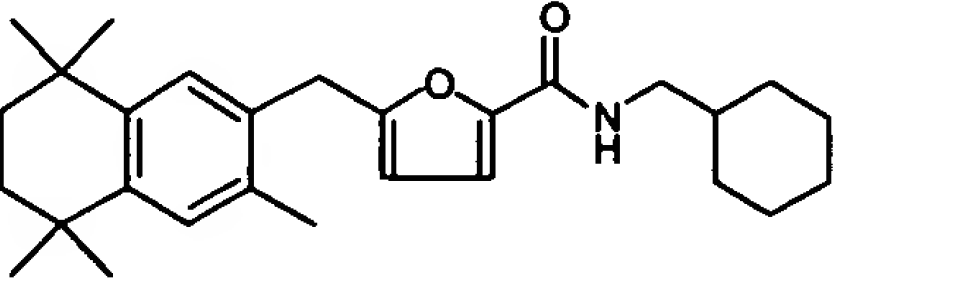
No.	Compound	Rat female				Dog	Monkey
		pmol/min/ mg	T _{1/2}	C _{max}	T _{max}	F _{p.o.}	pmol/min/ mg
		1 uM		uM			1 uM
168							
169						2301	5940
170							
176	 Chiral						
177						2456	3120
181		139	1.9 hr	3.64	4 hr	59% (20 mg/kg) ¹	
182			3.2 hr	1.1	1.5 hr	33% (10 mg/kg) ¹⁰	
183		64	3.6 hr	0.51	1 hr	37% (10 mg/kg) ¹⁰	
188							
185		224					

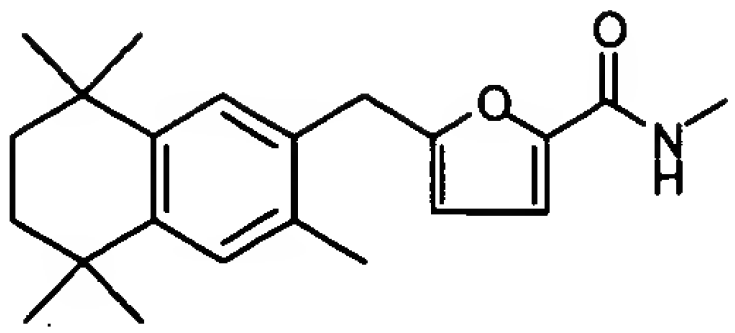
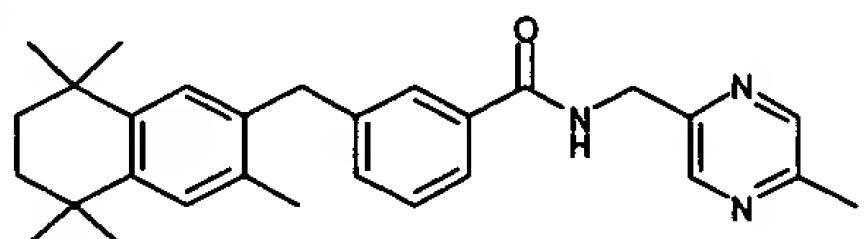
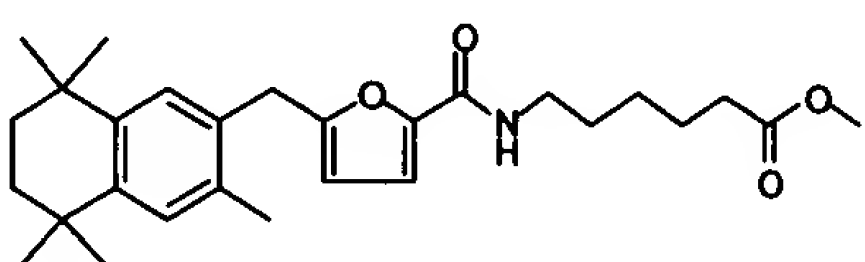
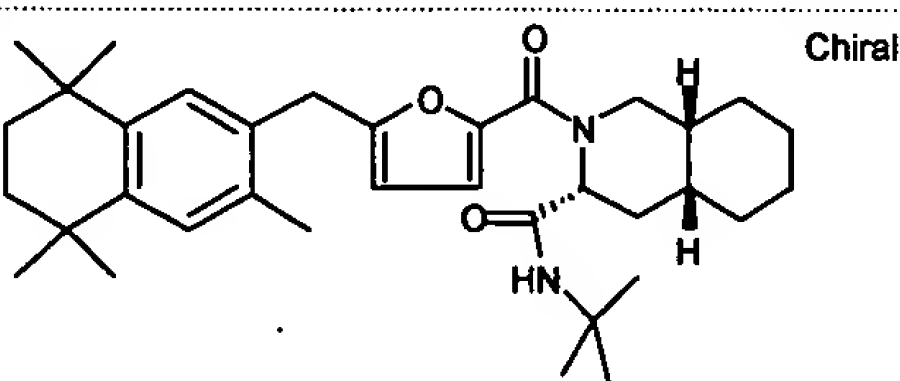
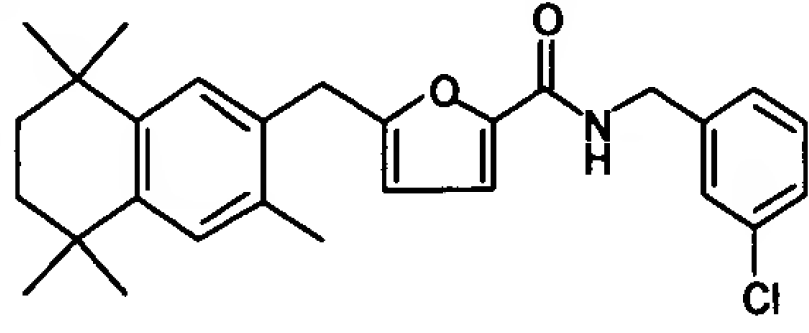
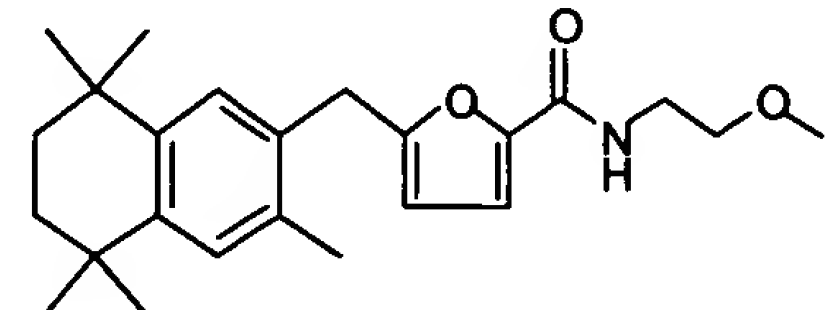
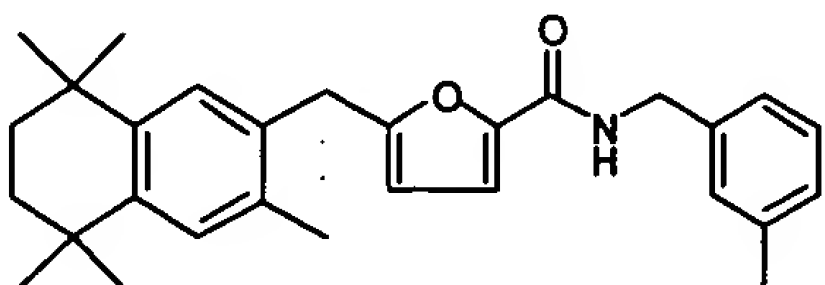
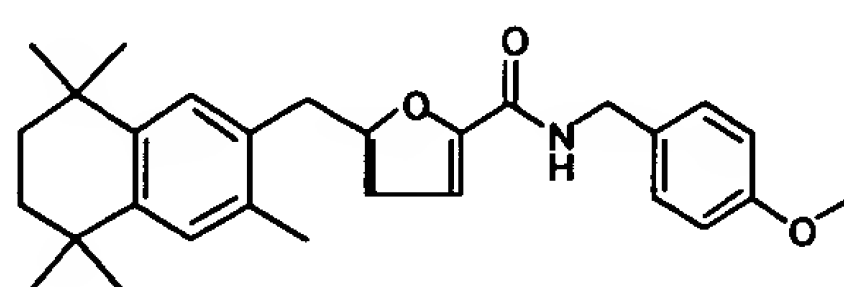
No.	Compound	Rat female				Dog		Monkey
		pmol/min/ mg	T _{1/2}	C _{max}	T _{max}	F _{p.o.}	pmol/min/ mg	pmol/min/ mg
		1 uM		uM			1 uM	1 uM
191								
194		205						
199		181	1.5 hr	0.08	4 hr	10% (10 mg/kg) ¹⁰		
200		86	2.0 hr	0.96	1.5 hr	33% (10 mg/kg) ¹⁰		
201			2.5 hr	1.9	30 min	21% (10 mg/kg) ¹⁰		
206		330						
207		169	1.6 hr	2.81	1 hr	41% (10 mg/kg) ¹⁰		
212		430	1.6 hr	0.05	1 hr	1% (10 mg/kg) ¹⁰		
217		203	3.8 hr	0.1	1 hr	16% (10 mg/kg) ¹⁰		

No.	Compound	Rat female				Dog	Monkey
		pmol/min/ mg	T _{1/2}	C _{max}	T _{max}	F _{p.o.}	pmol/min/ mg
		1 uM		uM			1 uM
221		223	3 hr	0.34	1 hr	41% (10mg/kg) ¹⁰	
222		167	3.2	0.81	1.5	54% (10mg/kg) ¹⁰	
225		330	1.1 hr	0.34	1 hr	24% (10mg/kg) ¹⁰	
226		173	4.6 hr	0.42	1 - 5 hr	34% (10mg/kg) ¹⁰	
228		227	2.4 hr	0.07	0.5 hr	2% (10mg/kg) ¹⁰	

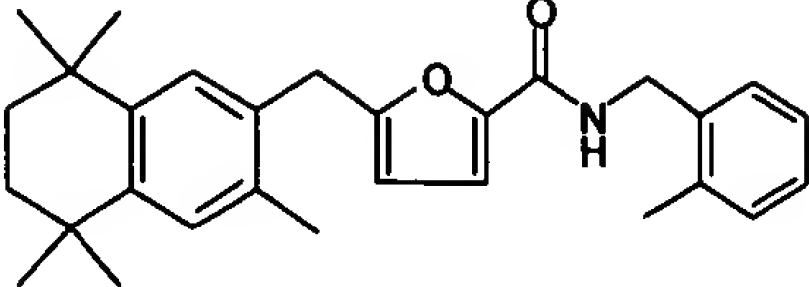
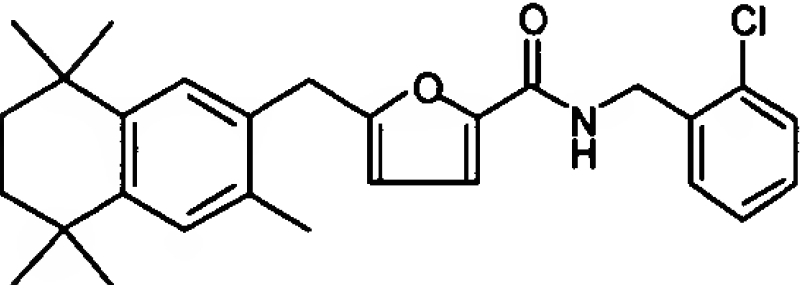
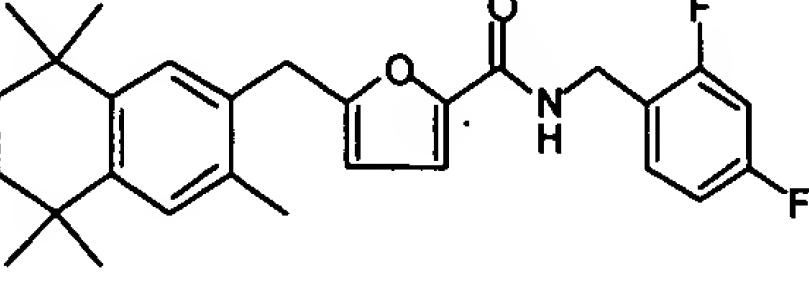
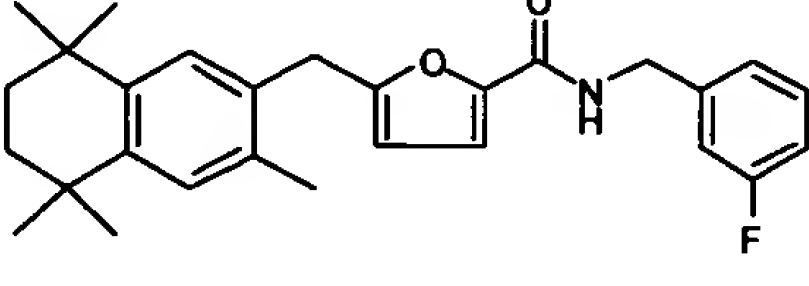
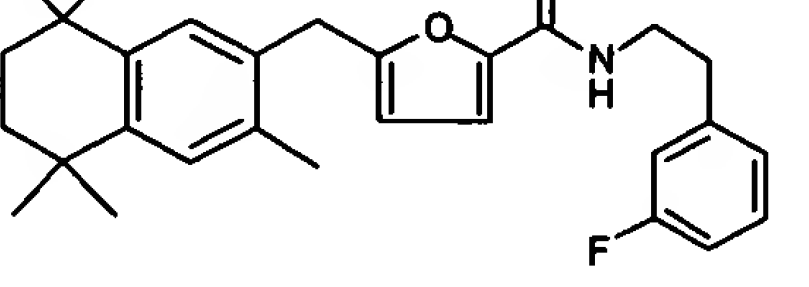
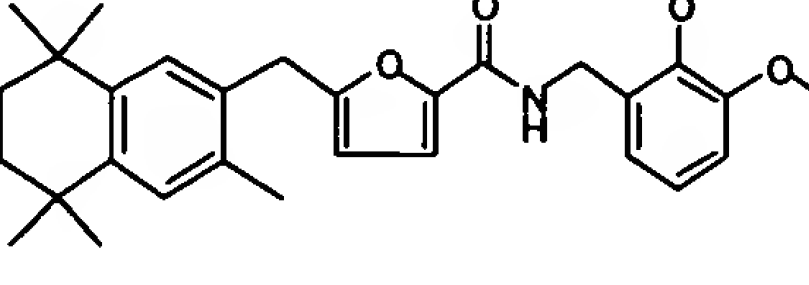
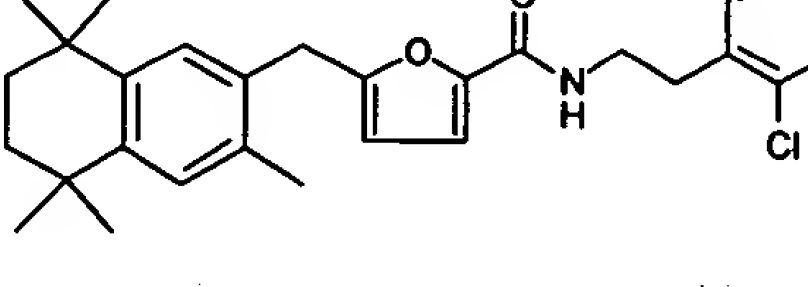
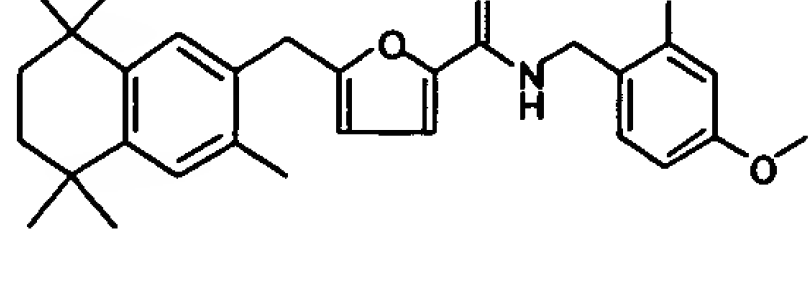
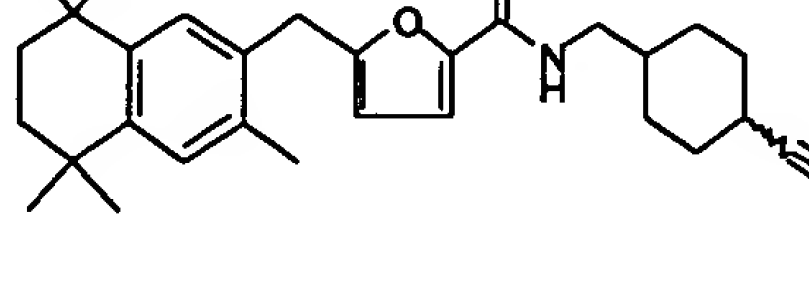
Cmpd. #	Structure	hGnRH IC50 (nM)	bGnRH IC50 (nM)	rGnRH IC50 (nM)	mGnRH IC50 (nM)	GnRH functional	GnRH Kb (nM)
9		220 (40*)	3800	680	2300	Antagonist	30
10		130 (20*)	1500	390	1400	Antagonist	95
21		920	3340	540	1200	Antagonist	
11		190 (80*)	320	9	50	Antagonist	
22		540	1710	310	1100		
23		220 (40*)	2830	730	2100	Antagonist	
24		550	760	150	560		

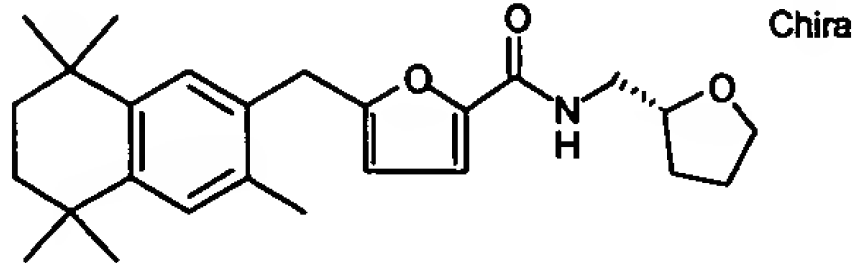
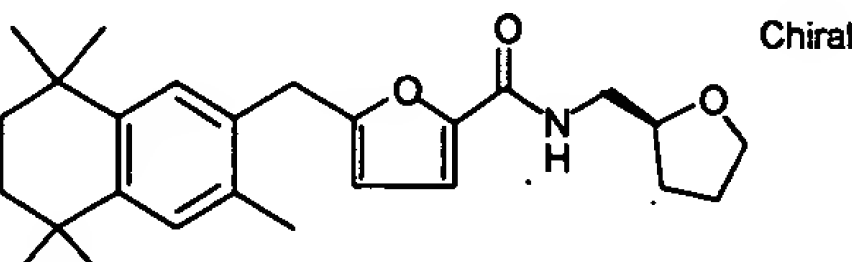
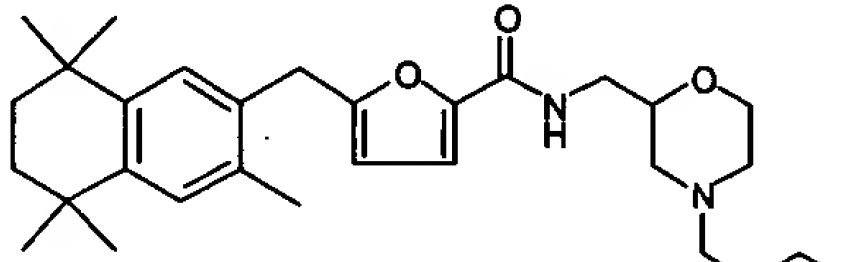
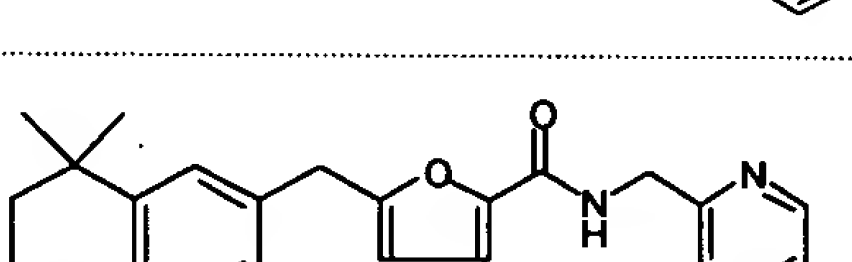
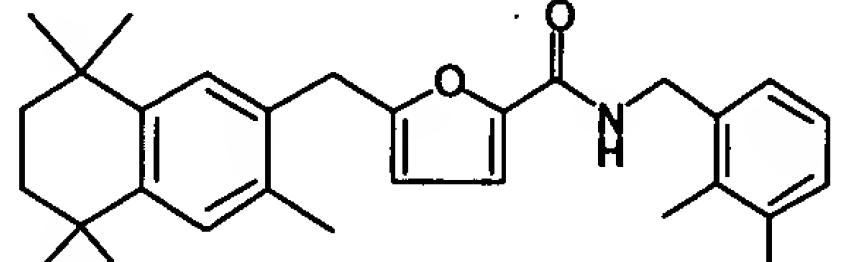
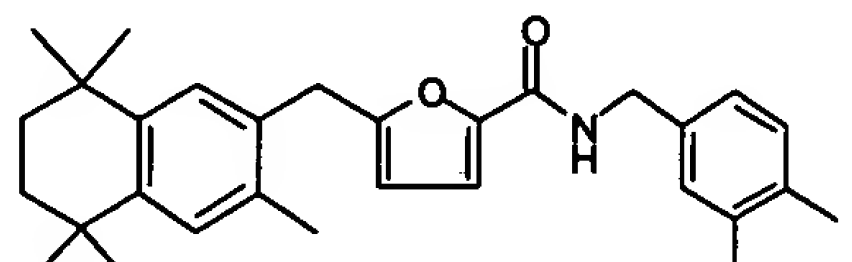
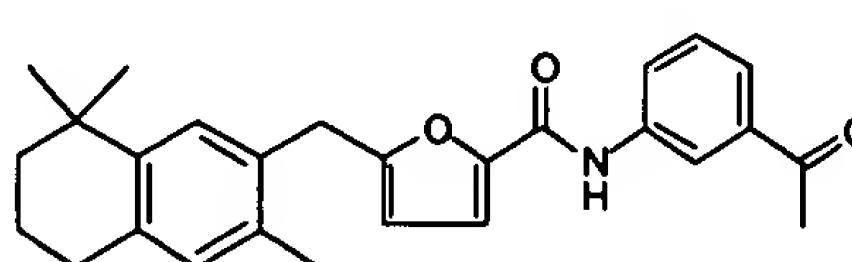
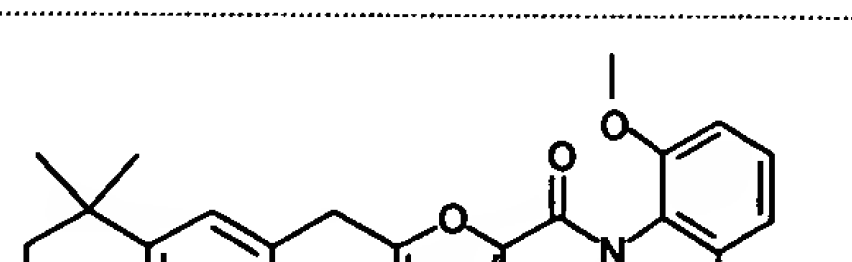
Cmpd. #	Structure	hGnRH IC50 (nM)	bGnRH IC50 (nM)	rGnRH IC50 (nM)	mGnRH IC50 (nM)	GnRH functional	GnRH Kb (nM)
25		410	940	260	950		
26		1010					
27		>10000					
28		>10000					
29		1420					
30		1350					
30.5		5030					

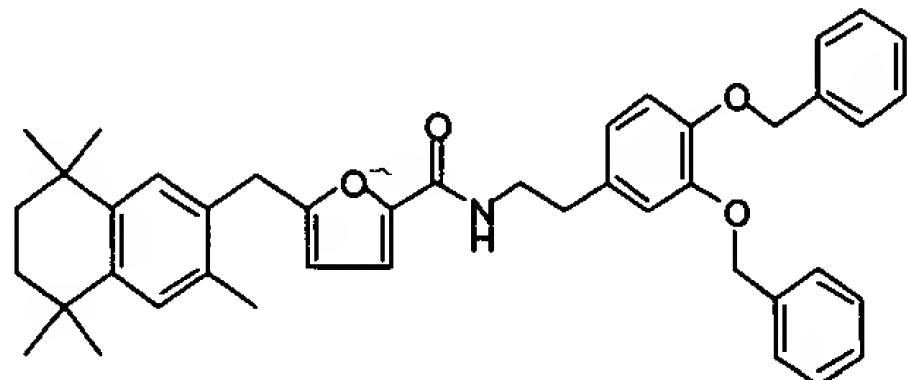
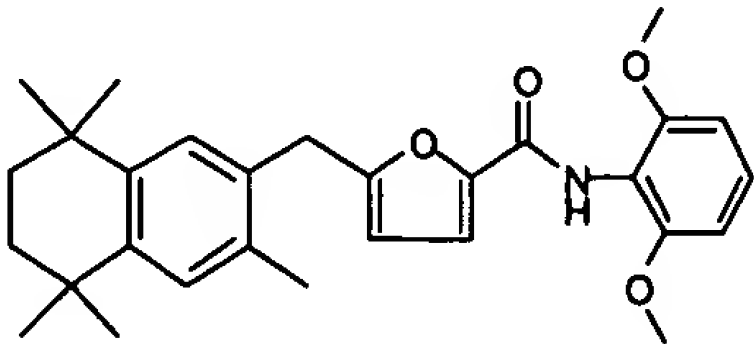
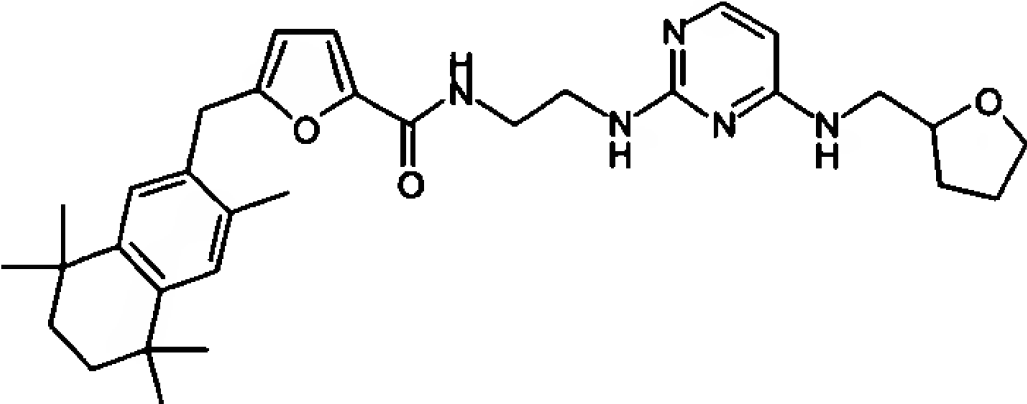
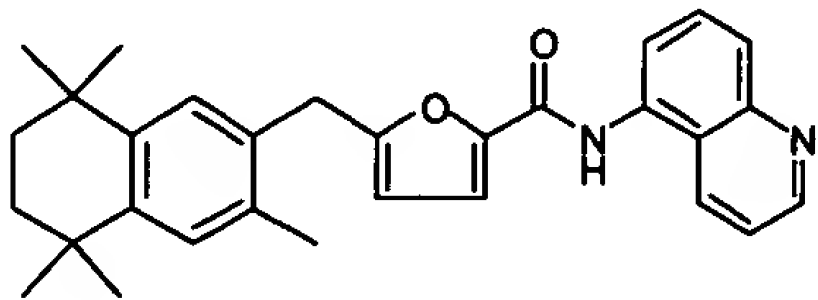
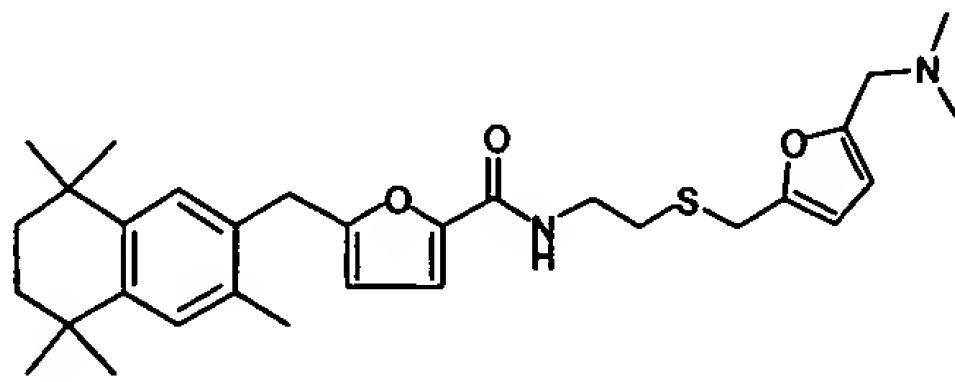
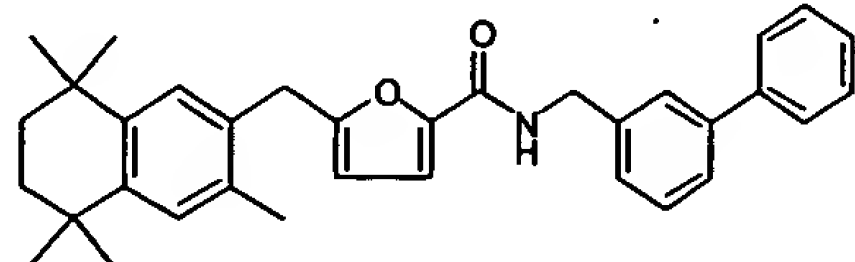
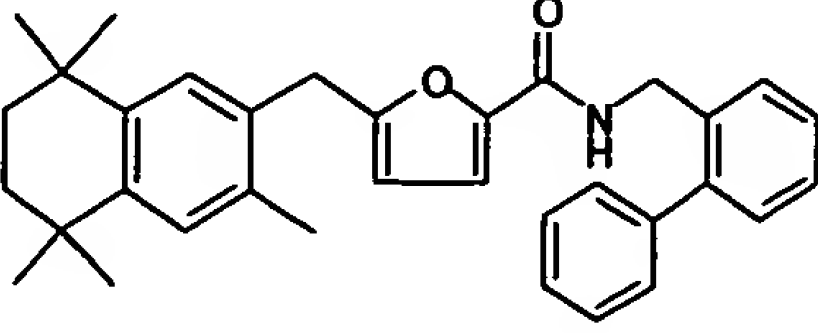
Cmpd. #	Structure	hGnRH IC50 (nM)	bGnRH IC50 (nM)	rGnRH IC50 (nM)	mGnRH IC50 (nM)	GnRH functional	GnRH Kb (nM)
12		230 (210*)	10400	3080	7130	Antagonist	170
31		140	8900	2800	1880	Antagonist	
32		3800					
33		5300					
13		110 (40*)	530	60	120	Antagonist	85
34		>10000					
35		1200					
36		215	12500	2900	3800	Antagonist	

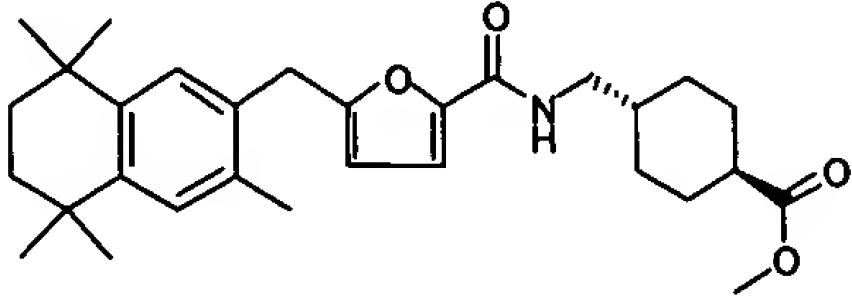
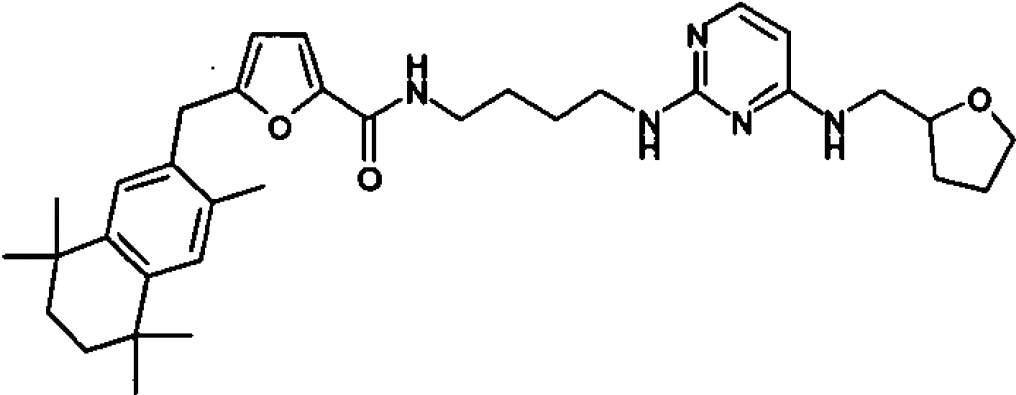
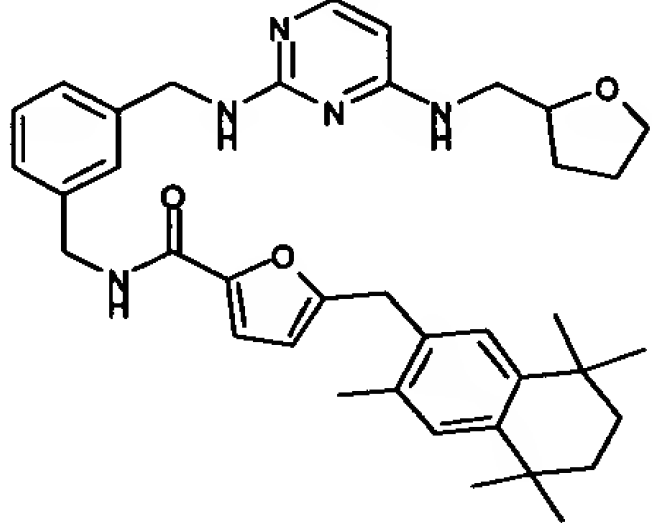
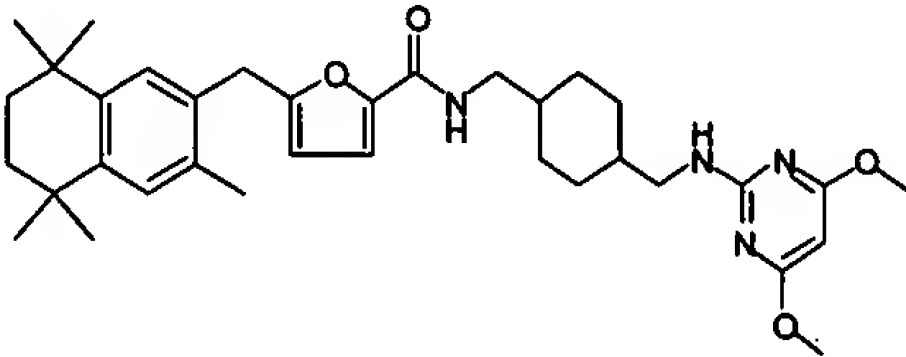
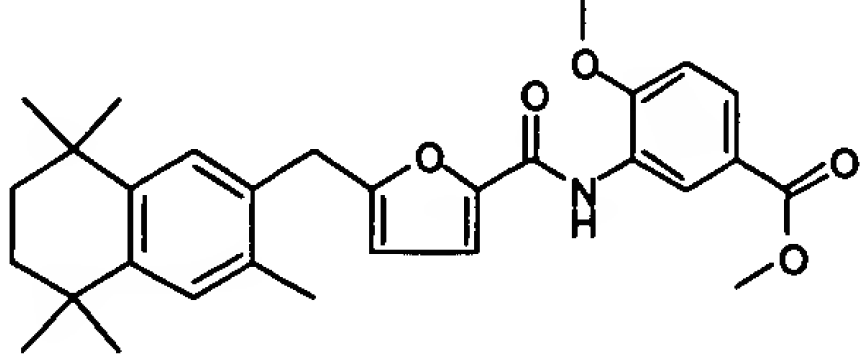
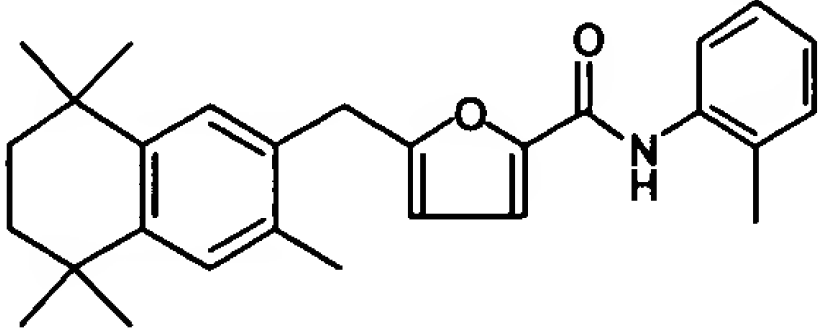
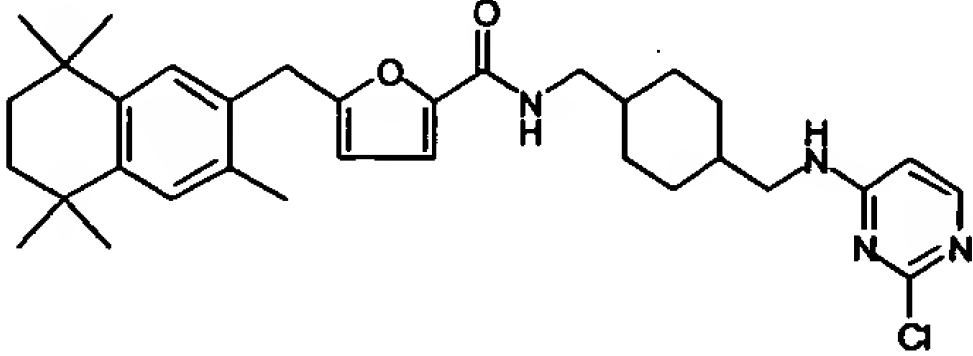
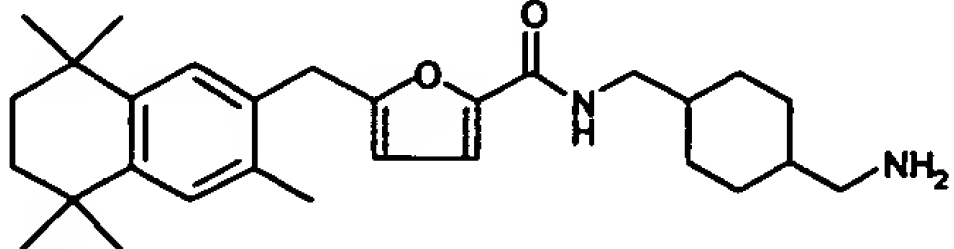
Cmpd. #	Structure	hGnRH IC50 (nM)	bGnRH IC50 (nM)	rGnRH IC50 (nM)	mGnRH IC50 (nM)	GnRH functional	GnRH Kb (nM)
37		2100					
37.5		7800					
38		370					
38.5		>10000					
39		190	2340	220	830		
40		530					
41		140	1400	210	490		
42		75(65*)	1780	100	280		

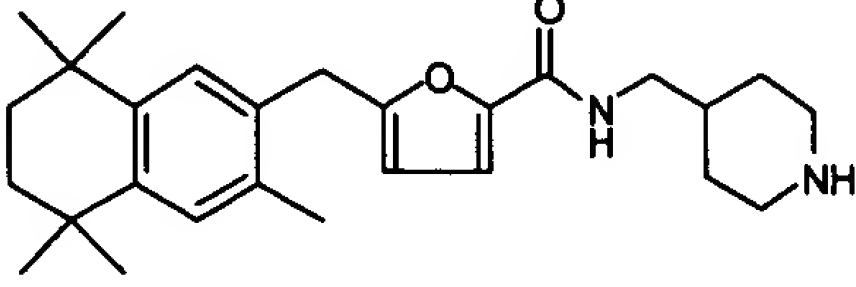
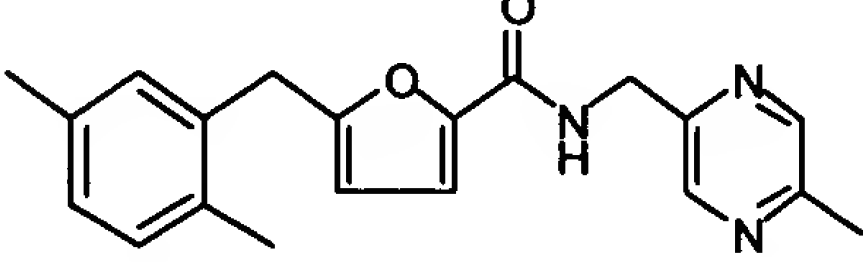
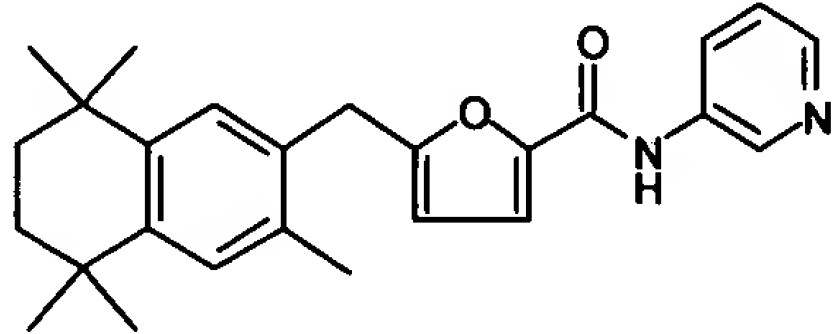
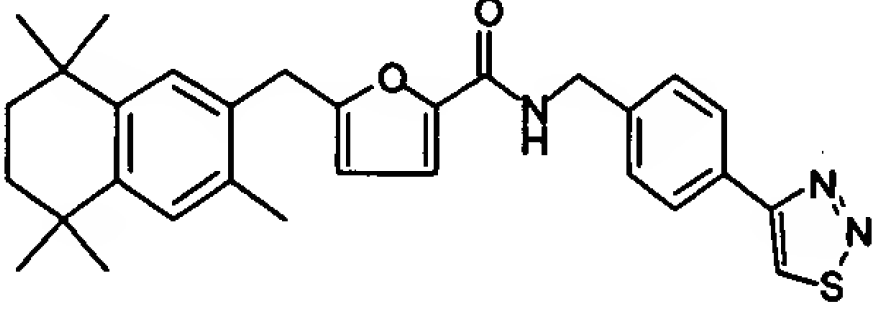
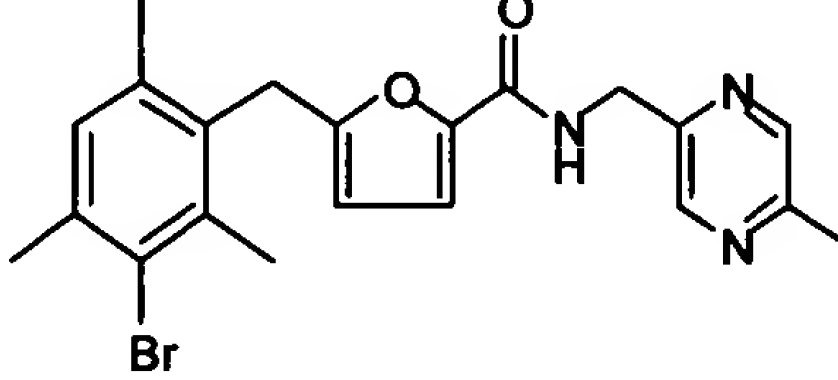
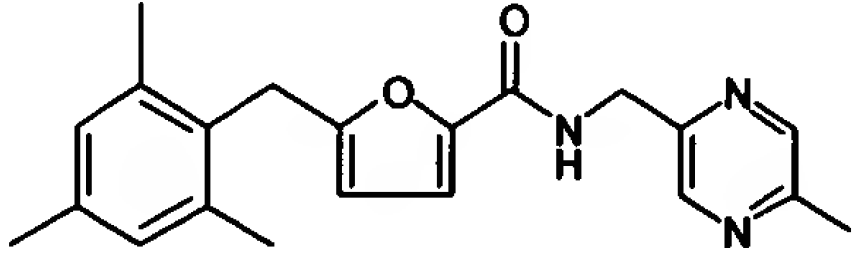
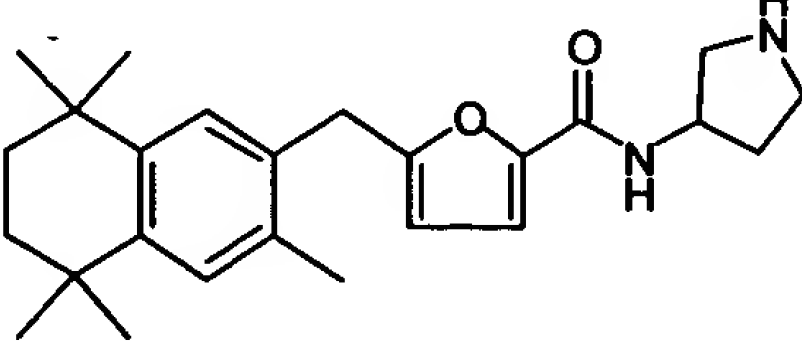
Cmpd. #	Structure	hGnRH IC50 (nM)	bGnRH IC50 (nM)	rGnRH IC50 (nM)	mGnRH IC50 (nM)	GnRH functional	GnRH Kb (nM)
43		320(130*)	4000	330	630		
44		390(120*)	4440	510	610		
45		>10000					
46		330*	8700	900	1880		
47		230(80*)	1920	130	240		
48		50(40*)	490	6	10	Antagonist	
14		80	1050	60	290		
49		3700					

Cmpd. #	Structure	hGnRH IC50 (nM)	bGnRH IC50 (nM)	rGnRH IC50 (nM)	mGnRH IC50 (nM)	GnRH functional	GnRH Kb (nM)
50		230	1520	150	370		
51		150	1930	120	400		
52		280	2900	270	660		
53		250	2930	270	630		
54		680					
15		290					
55		570					
56		100	1000	70	220		
16		30	4380	560	1290		

Cmpd. #	Structure	hGnRH IC50 (nM)	bGnRH IC50 (nM)	rGnRH IC50 (nM)	mGnRH IC50 (nM)	GnRH functional	GnRH Kb (nM)
57	 Chiral	830					
58	 Chiral	205					
59		980					
60		4200					
17		80	670	30	80		
18		55	460	40	115		
19		50	605	60	160		
61		760					

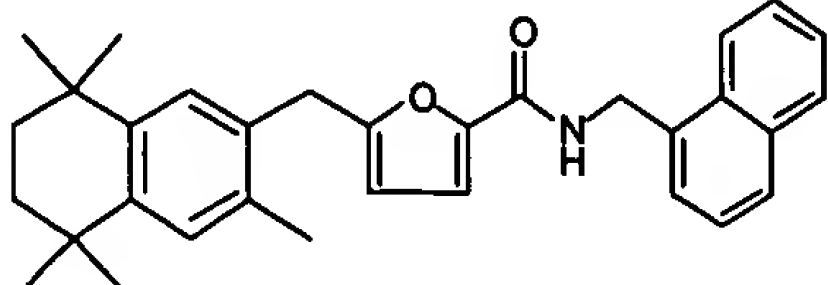
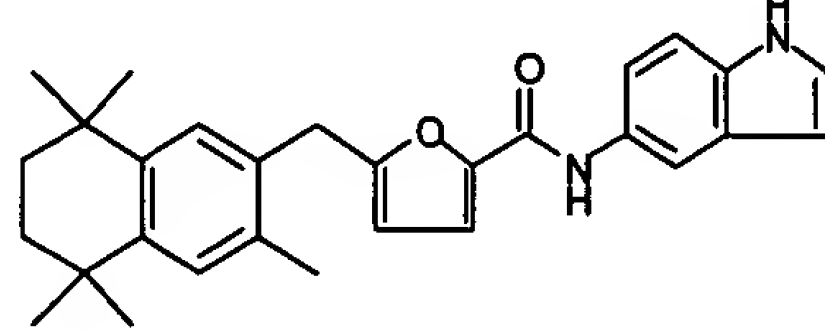
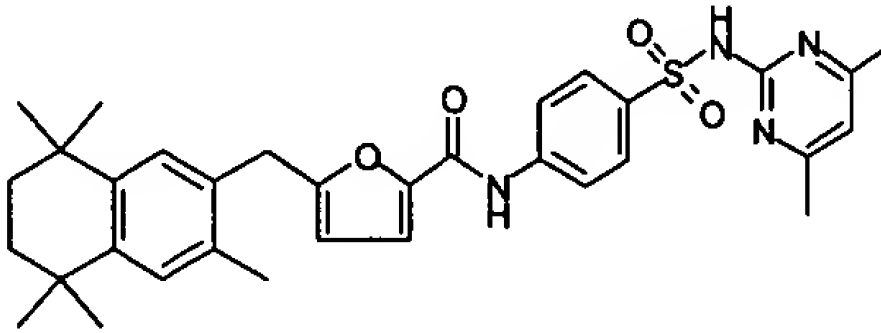
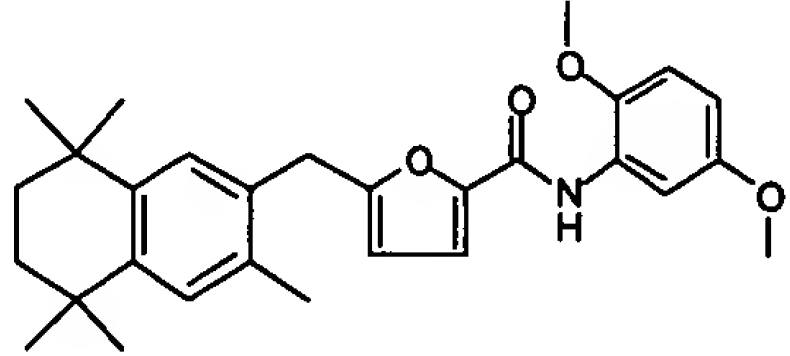
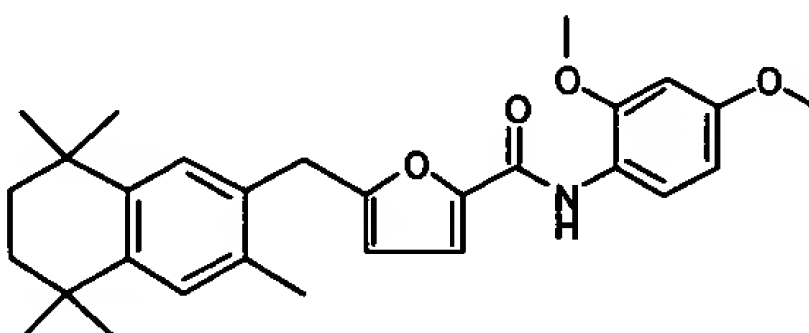
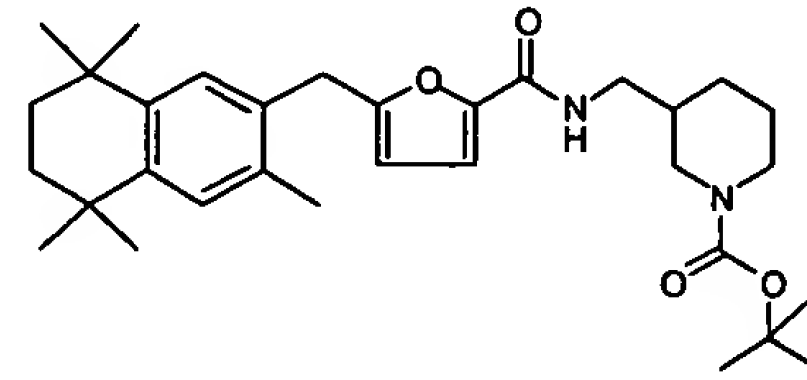
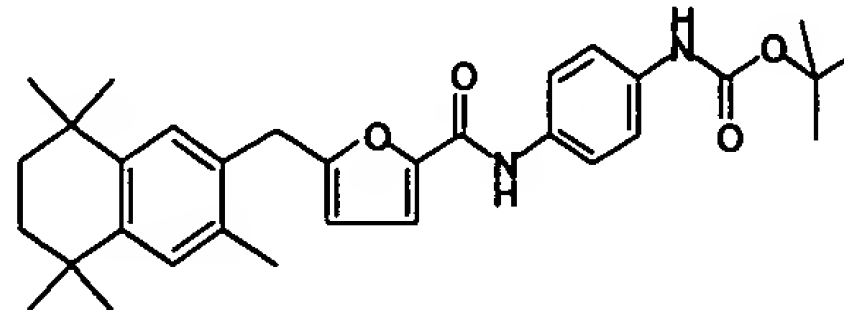
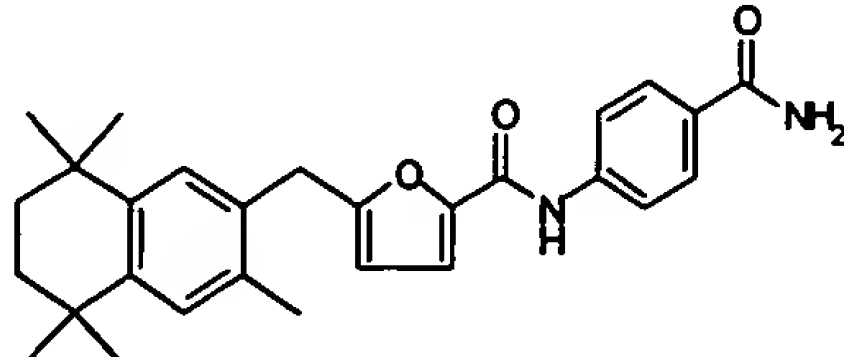
Cmpd. #	Structure	hGnRH IC50 (nM)	bGnRH IC50 (nM)	rGnRH IC50 (nM)	mGnRH IC50 (nM)	GnRH functional	GnRH Kb (nM)
62		>10000					
20		8	130	5	20	Antagonist (rat)	9 (rat)
63		2500					
64		125	380	60	34		
65		710					
66		3500					
67				1160			

Cmpd. #	Structure	hGnRH IC50 (nM)	bGnRH IC50 (nM)	rGnRH IC50 (nM)	mGnRH IC50 (nM)	GnRH functional	GnRH Kb (nM)
68		90		890	1010		
69		1410					
70		50	43	6	6	Antagonist (rat)	8 (rat)
71		2000					
72		130	1440	84	85		
73		580					
73		180		2140	2940		
75		115		740	710		

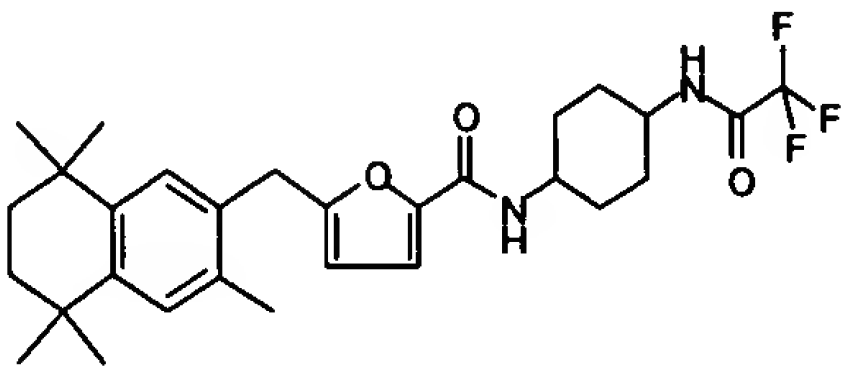
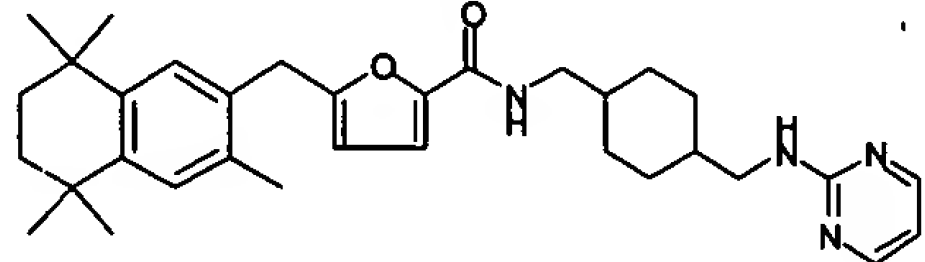
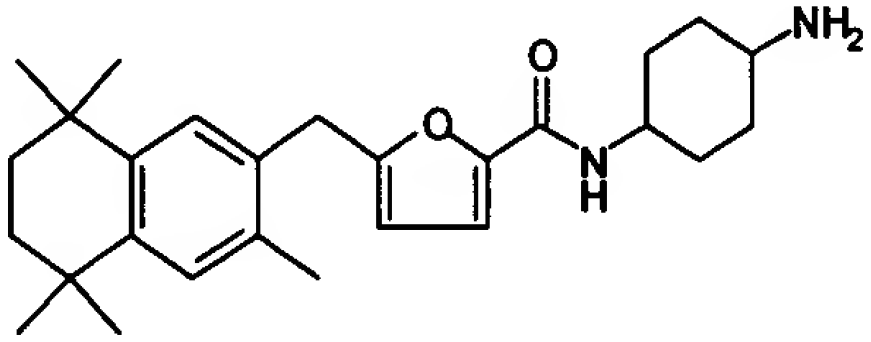
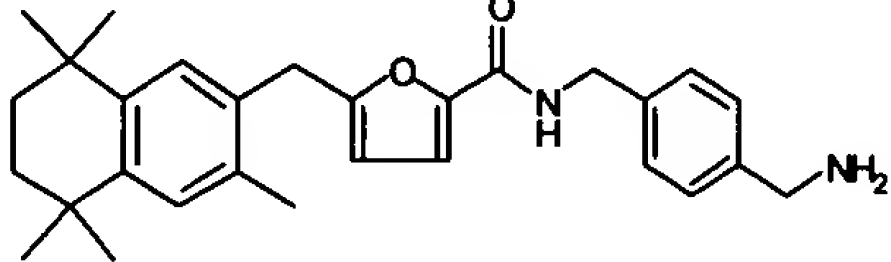
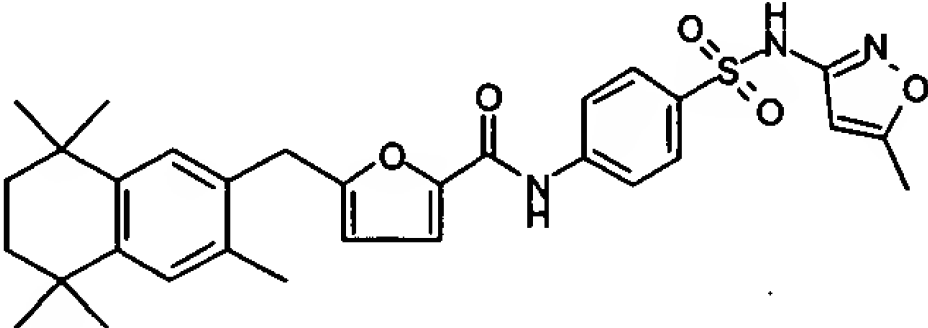
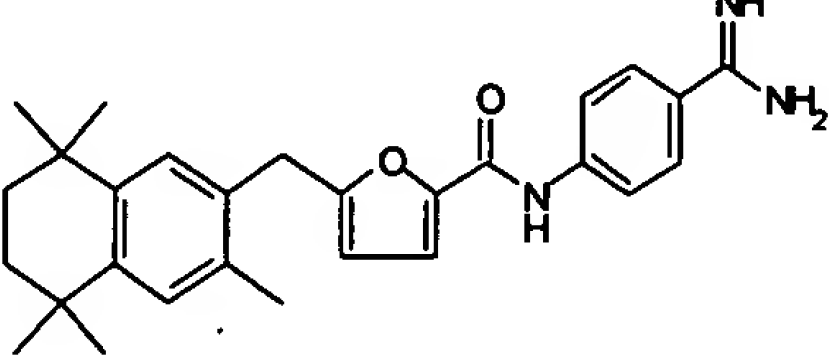
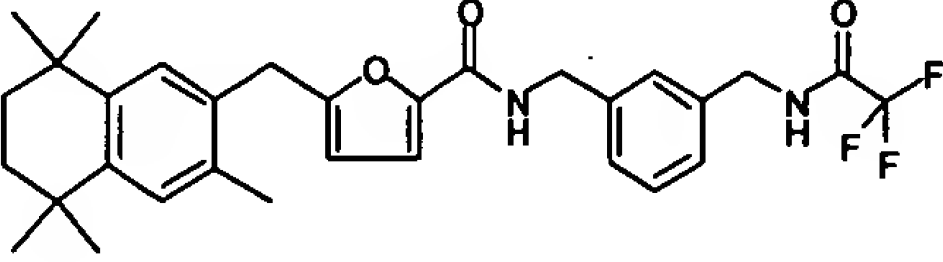
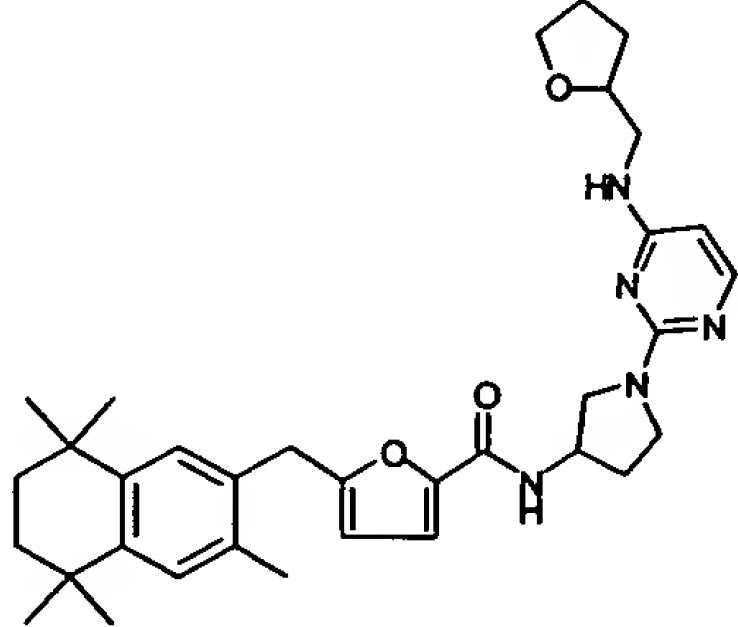
Cmpd. #	Structure	hGnRH IC50 (nM)	bGnRH IC50 (nM)	rGnRH IC50 (nM)	mGnRH IC50 (nM)	GnRH functional	GnRH Kb (nM)
76		880		5300			
77		>10000		8450			
78		>1000 (<10000)		1300			
79		250		370			
80		10000		5610			
81		12000		8080			
82		10000		9200			

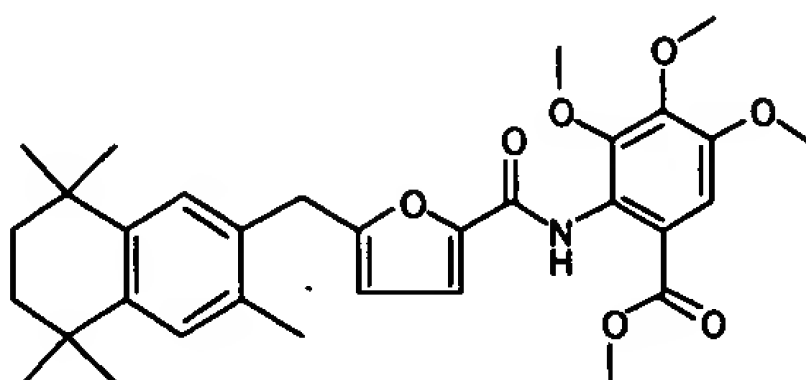
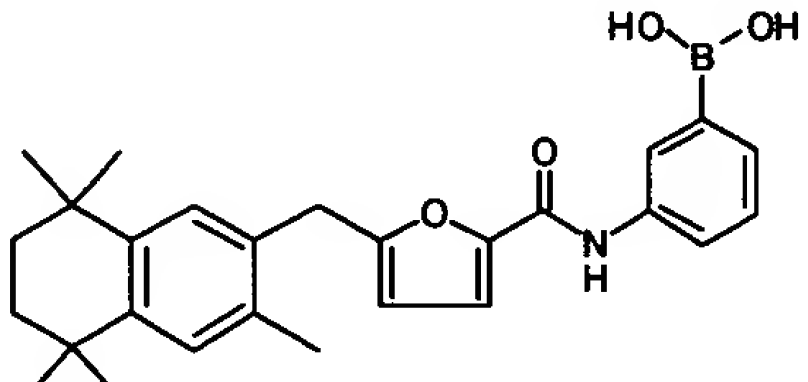
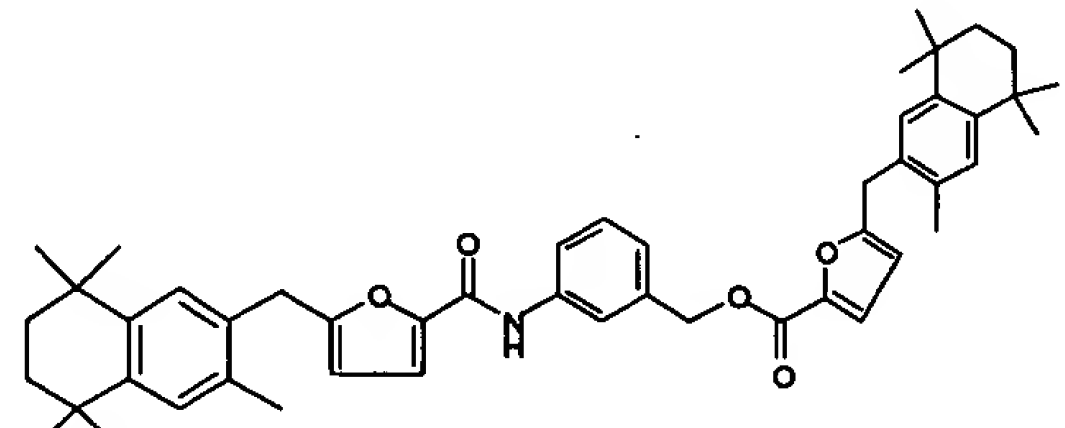
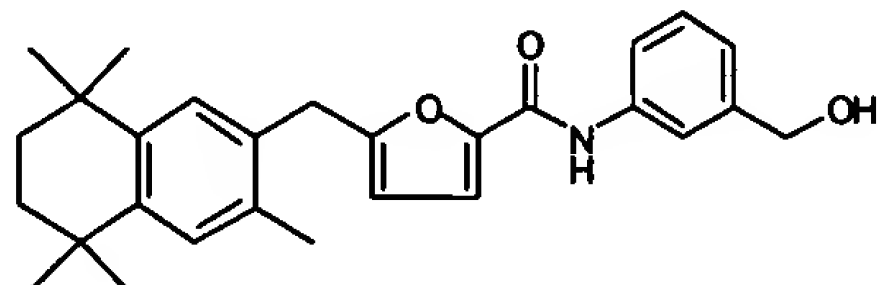
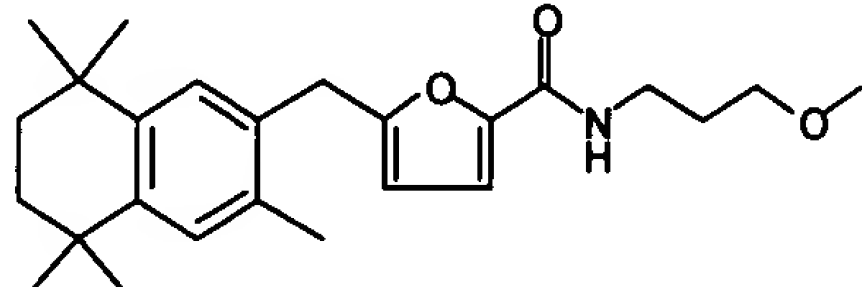
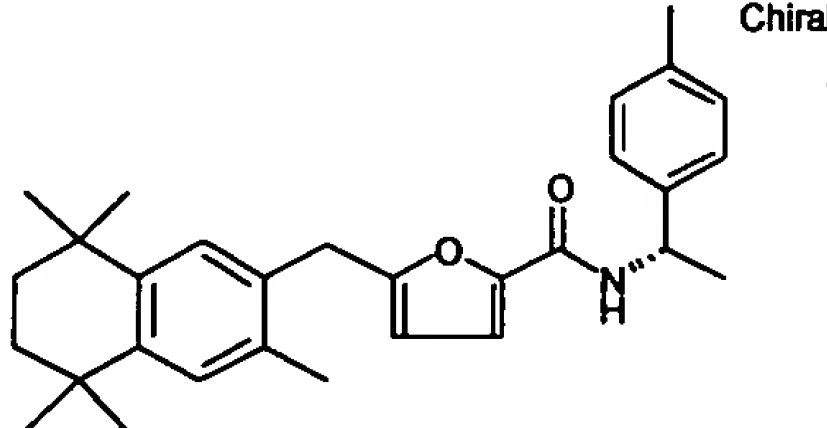
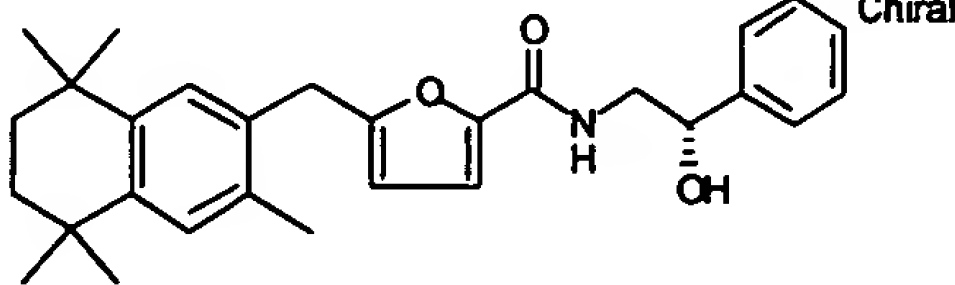
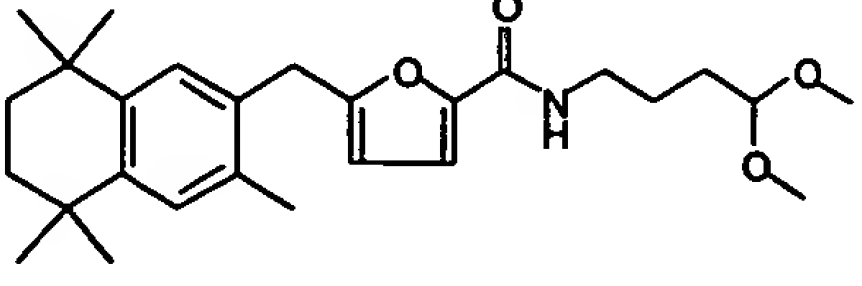
Cmpd. #	Structure	hGnRH IC50 (nM)	bGnRH IC50 (nM)	rGnRH IC50 (nM)	mGnRH IC50 (nM)	GnRH functional	GnRH Kb (nM)
83		620		1740			
84		8000		770			
85		12000		>10000			
86		8000		610			
86.5		2570		2050			
87		270		3310			
88		1000		2340			

Cmpd. #	Structure	hGnRH IC50 (nM)	bGnRH IC50 (nM)	rGnRH IC50 (nM)	mGnRH IC50 (nM)	GnRH functional	GnRH Kb (nM)
89		250		2730			
90		560		1260			
91		120		950	790		
92		80		27	42		
93		8000		5470			
94		315		1490	1300		
94.2		3000		1580			
94.5		>1000 (<10000)		4830			

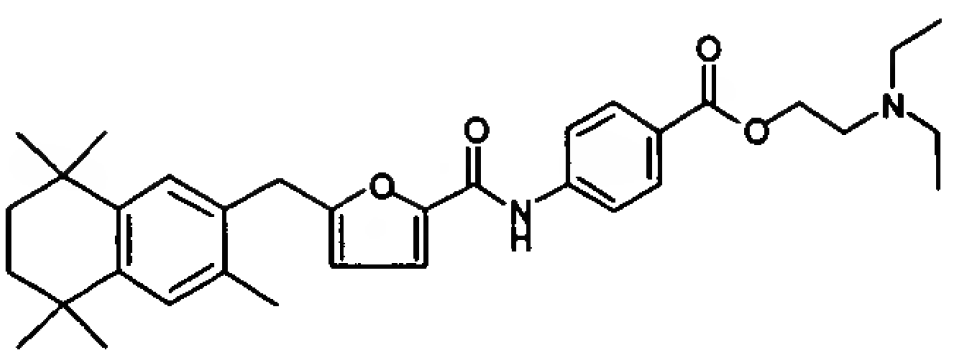
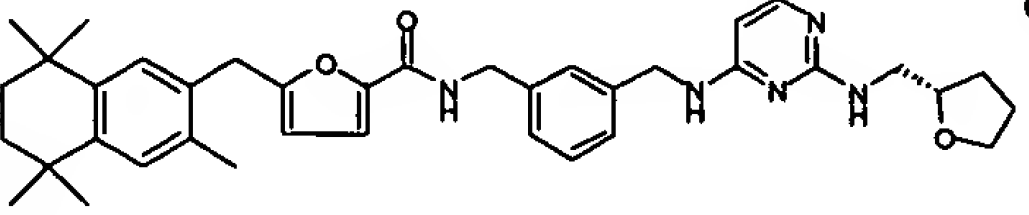
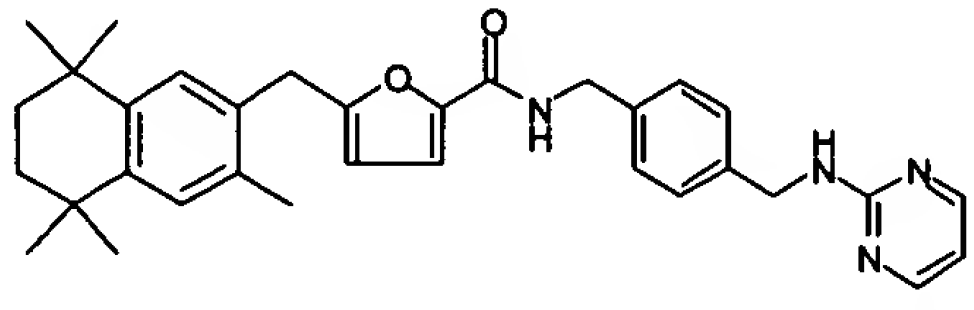
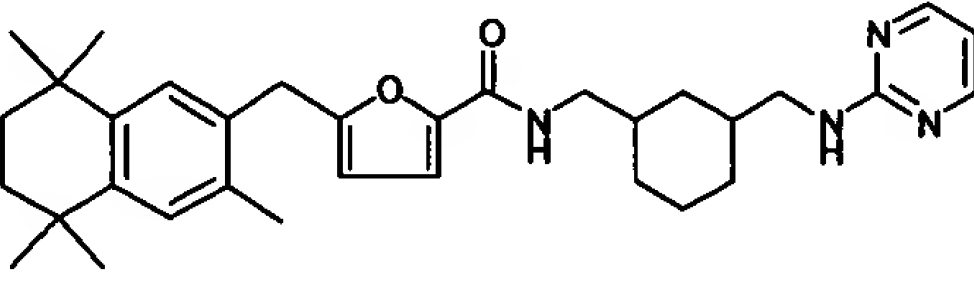
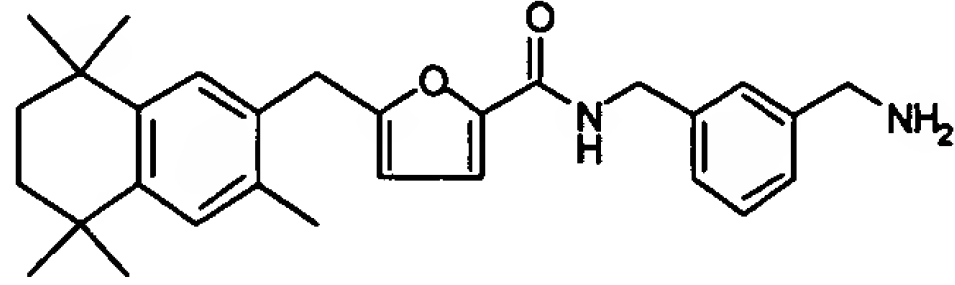
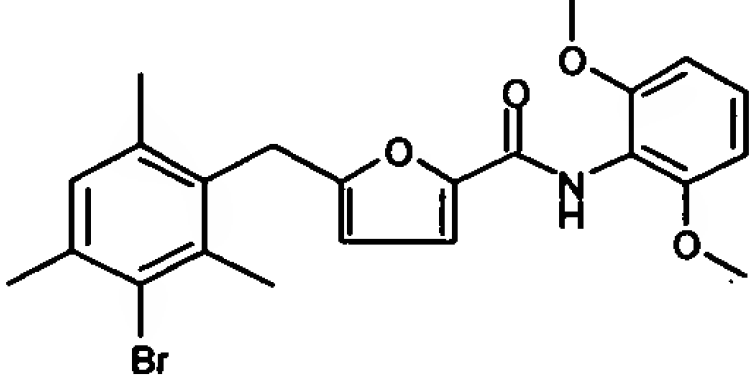
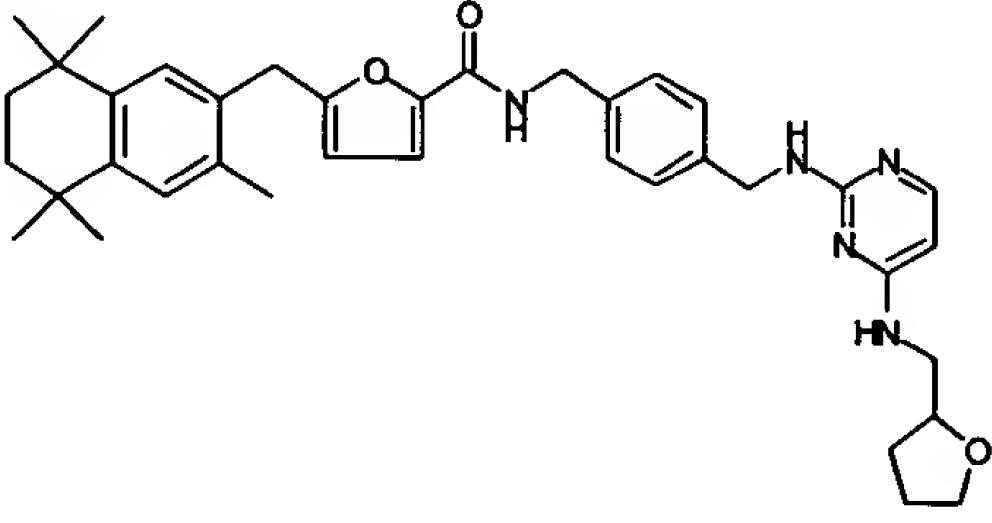
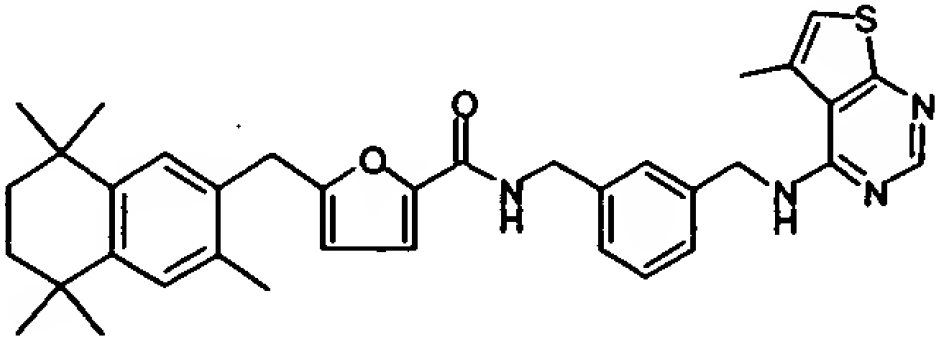
Cmpd. #	Structure	hGnRH IC50 (nM)	bGnRH IC50 (nM)	rGnRH IC50 (nM)	mGnRH IC50 (nM)	GnRH functional	GnRH Kb (nM)
95		31		60	21		
96		40		90	130		
97		1700		1690			
98		80		220	84		
99		56		108	105		
100		585		6000			
101		>10000		>10000			
102		1180		970			

Cmpd. #	Structure	hGnRH IC ₅₀ (nM)	bGnRH IC ₅₀ (nM)	rGnRH IC ₅₀ (nM)	mGnRH IC ₅₀ (nM)	GnRH functional	GnRH K _b (nM)
103		360		3460			
104		>1000 (<10000)		1260			
105		22		250	200		
106		2000		510			
107		>10000		>10000			
108		1000		2740			
109		75		140	140		

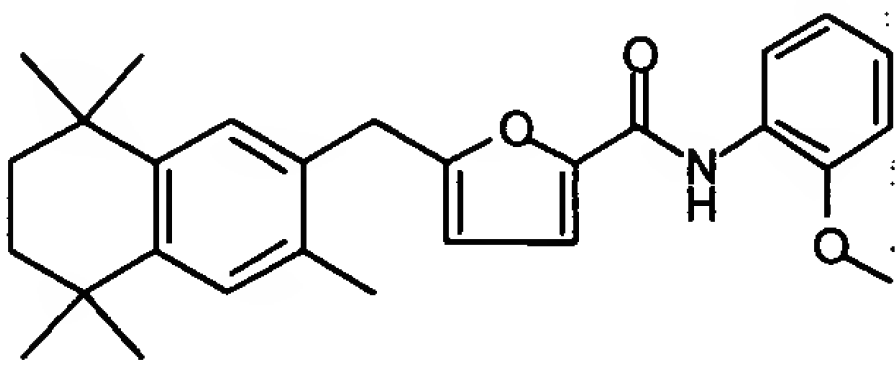
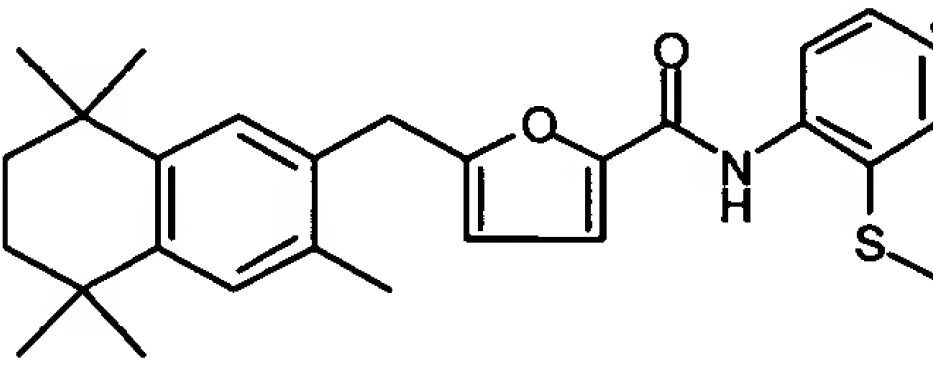
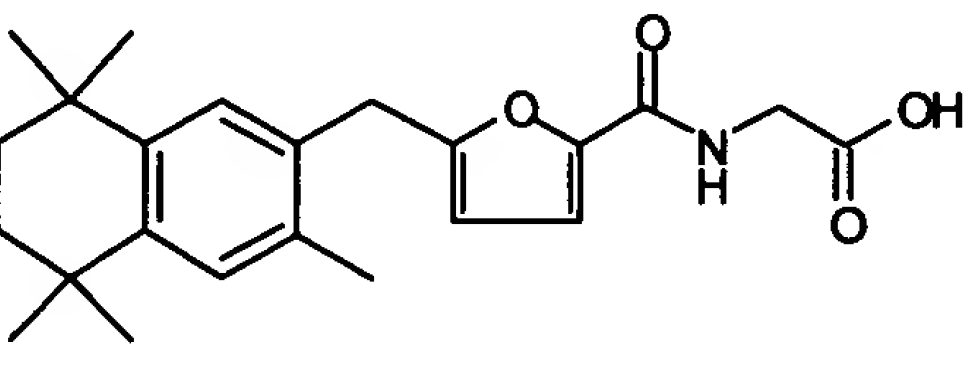
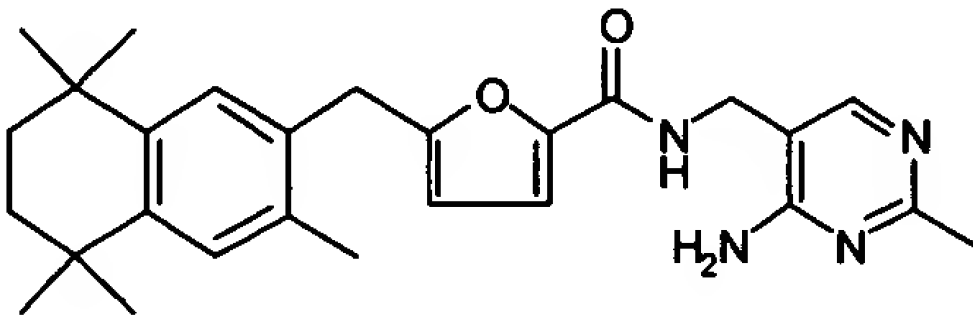
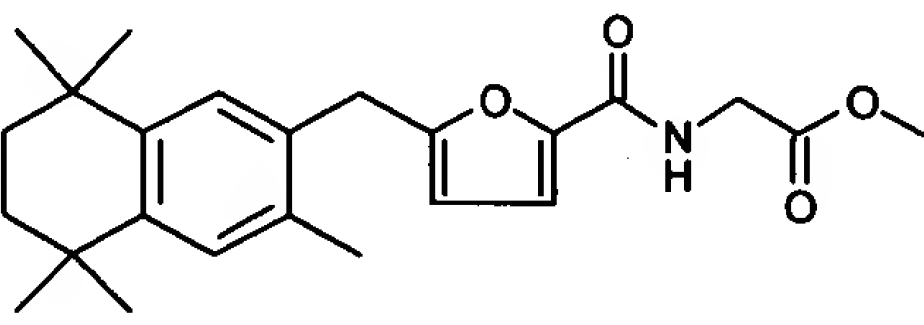
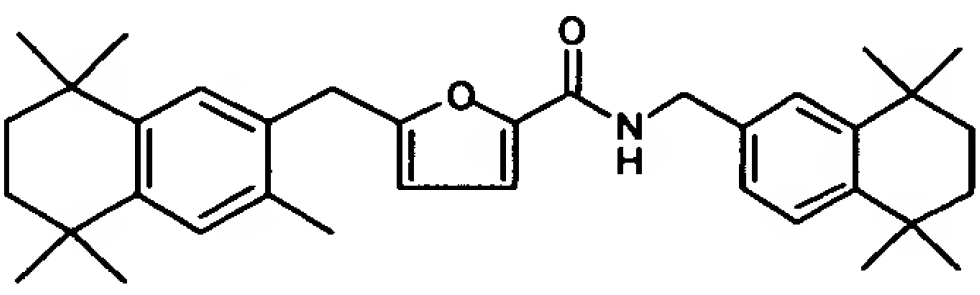
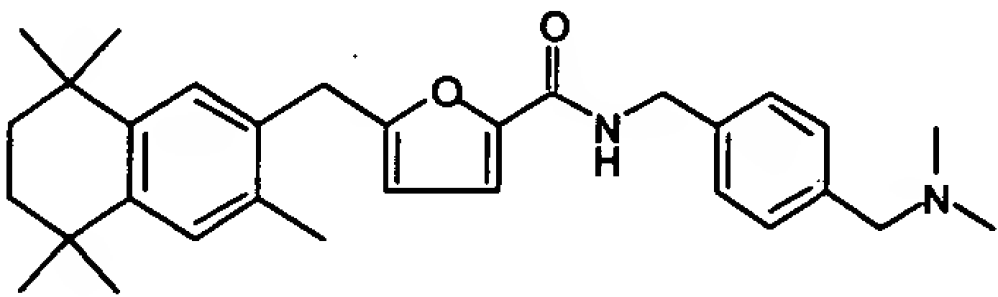
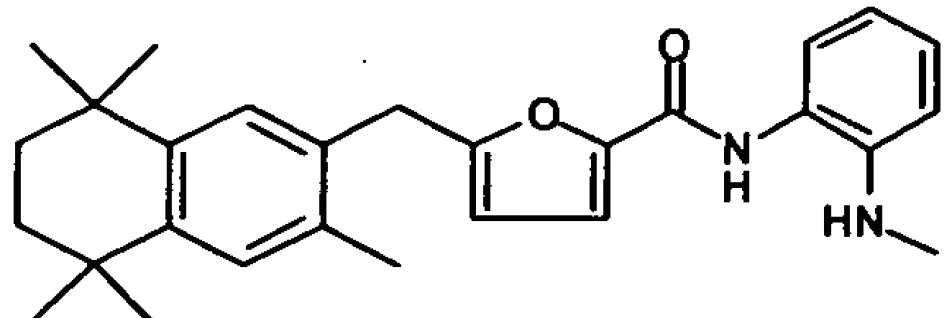
Cmpd. #	Structure	hGnRH IC50 (nM)	bGnRH IC50 (nM)	rGnRH IC50 (nM)	mGnRH IC50 (nM)	GnRH functional	GnRH Kb (nM)
110		12000		>10000			
111		630		1740			
112		12000		7780			
113		340		300			
114		10000		7000			
115		6000		6130			
116		1170		390			
117		1810		3000			

Cmpd. #	Structure	hGnRH IC50 (nM)	bGnRH IC50 (nM)	rGnRH IC50 (nM)	mGnRH IC50 (nM)	GnRH functional	GnRH Kb (nM)
118		380		430			
119		12000		1300			
120		>10000		>10000			
121		1100					
122		730		1260			
123		510					
124		1000		750			
125		135		560	1050		

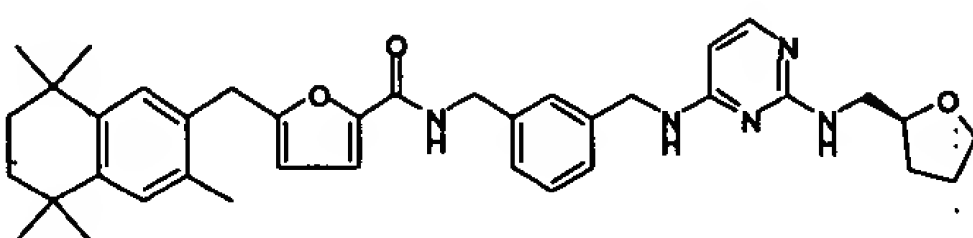
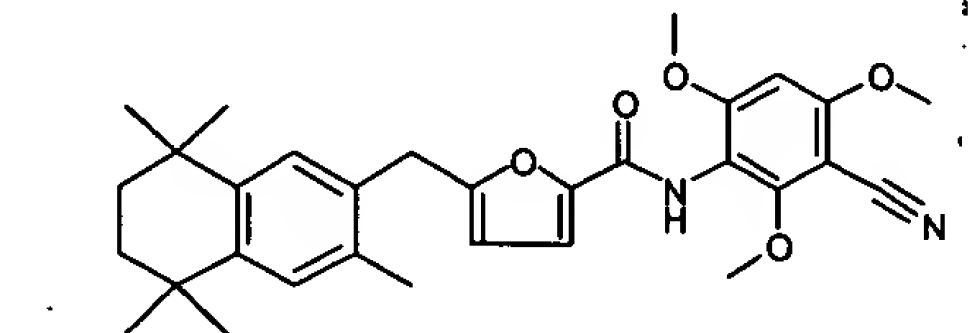
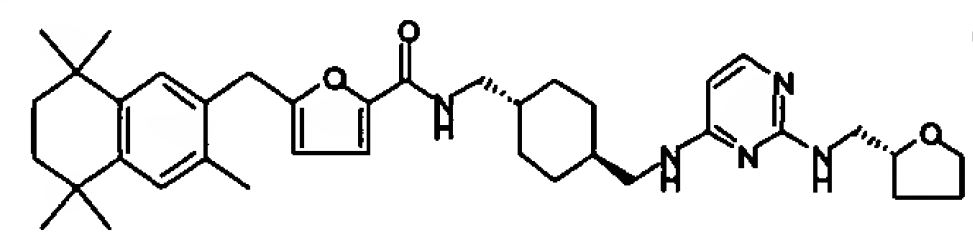
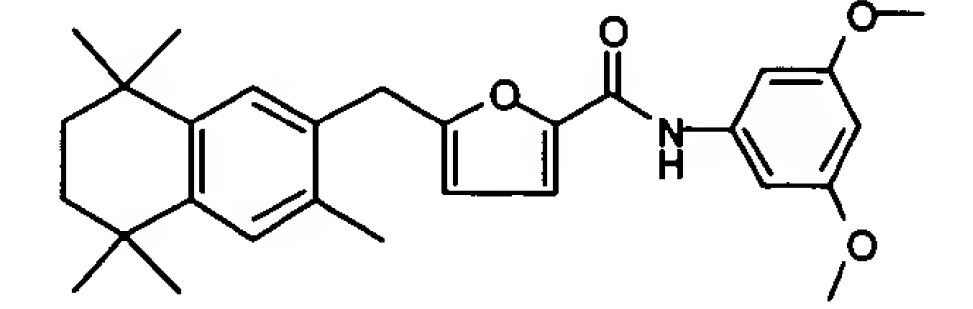
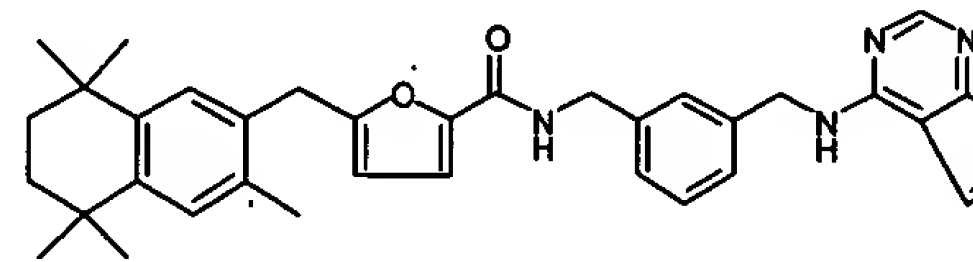
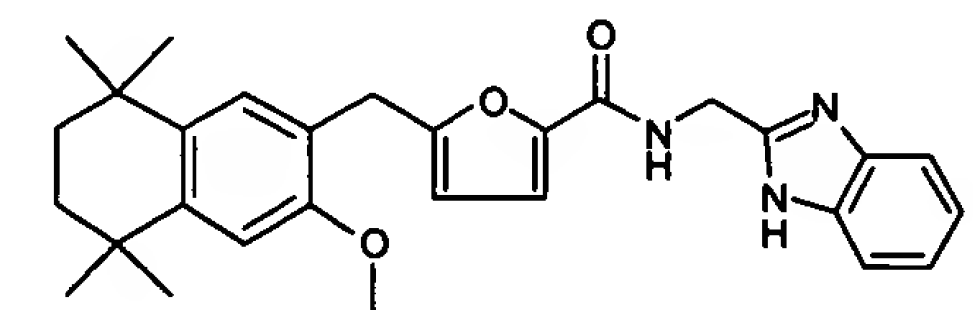
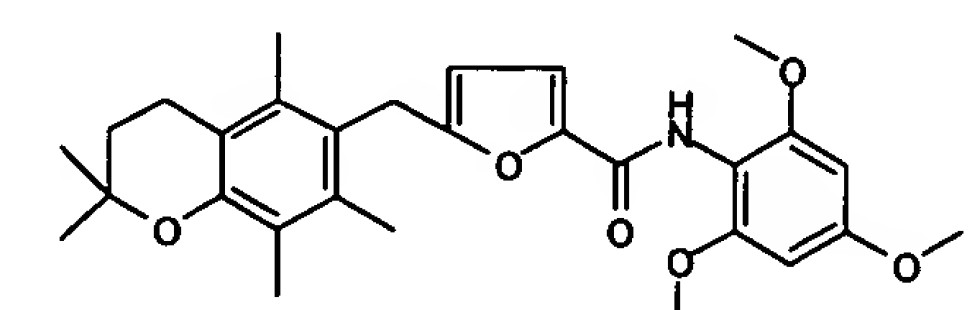
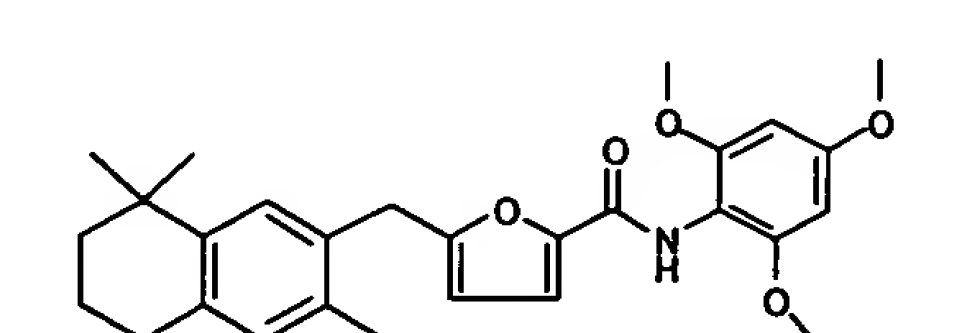
Cmpd. #	Structure	hGnRH IC50 (nM)	bGnRH IC50 (nM)	rGnRH IC50 (nM)	mGnRH IC50 (nM)	GnRH functional	GnRH Kb (nM)
126		128**		325**	684**		
127		510		910			
128		220		170	220		
129		12000		1390			
130		3000		590			
131		600					
132		640					
133		490					
134		6		3	3	Antagonist	13 (rpH)13 (hPI)

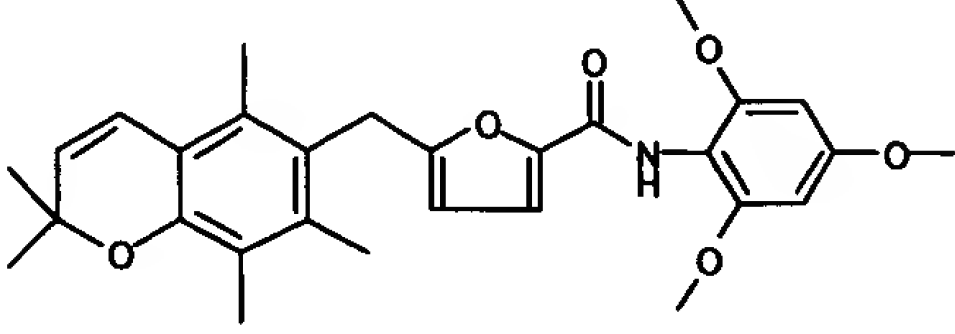
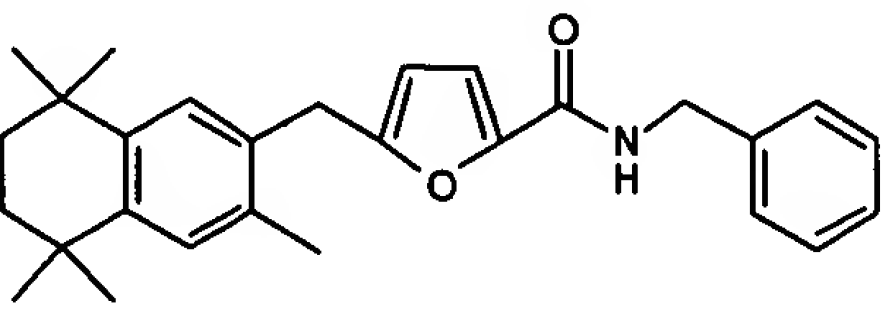
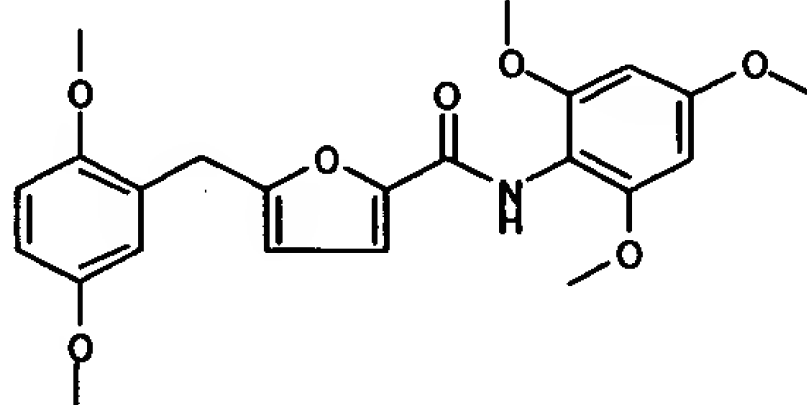
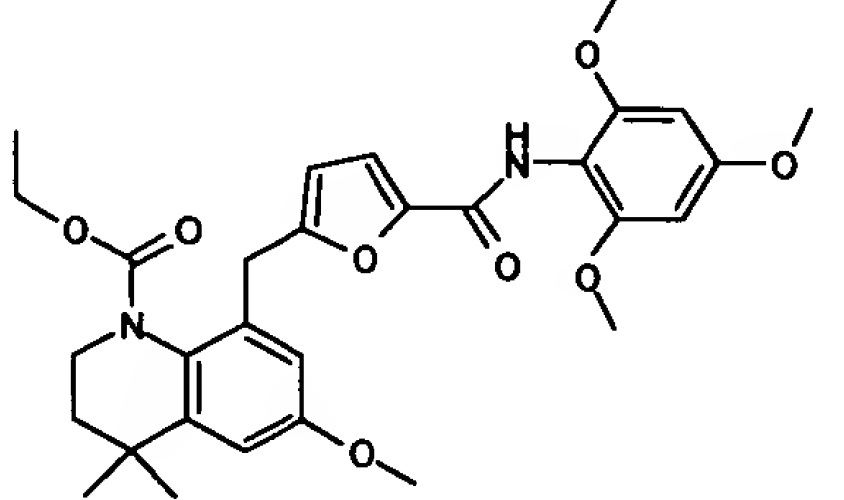
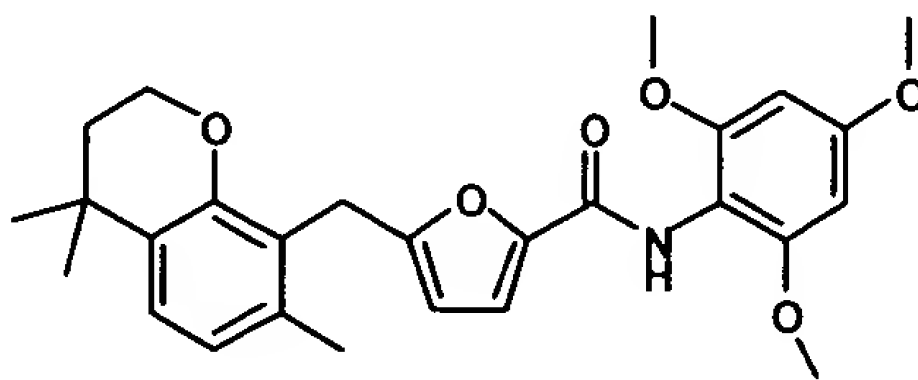
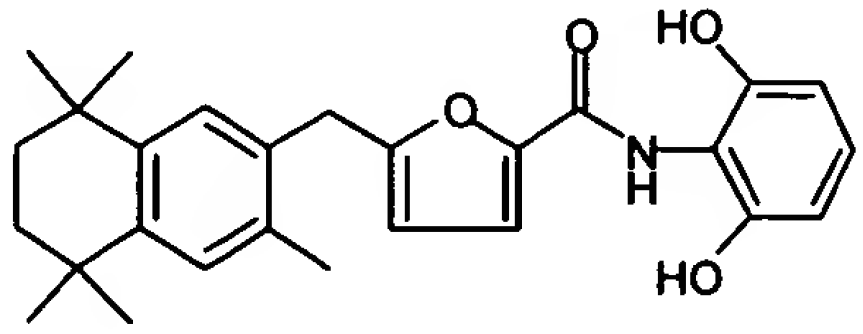
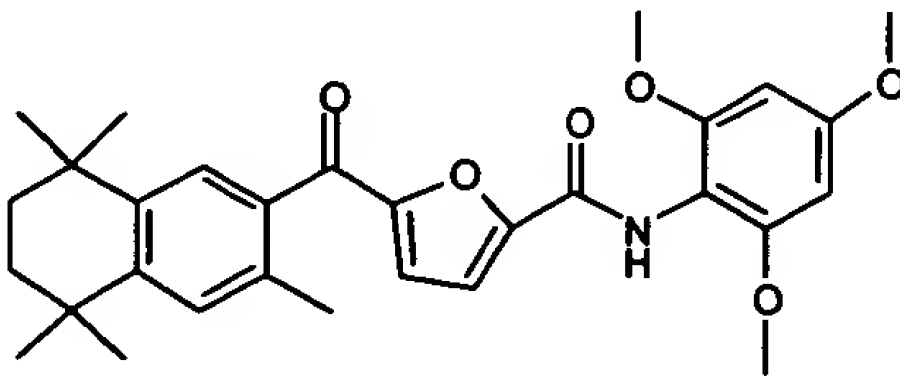
Cmpd. #	Structure	hGnRH IC50 (nM)	bGnRH IC50 (nM)	rGnRH IC50 (nM)	mGnRH IC50 (nM)	GnRH functional	GnRH Kb (nM)
135		1440					
136	 Chiral	9.4		4.7	8.6	Antagonist	6.3 & 8 rat PI, pH
137		200		400	360		
138		200		1630	1570		
139		110		113	150		
140		110		530	320		
141		150		35	64		
142		3000 (solubility?)					

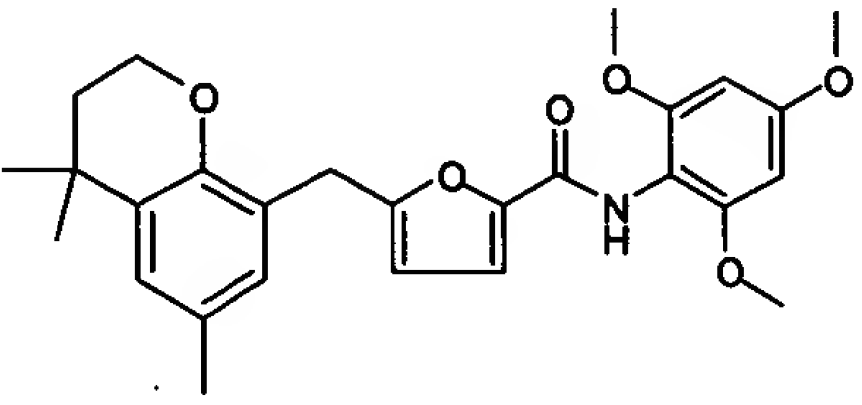
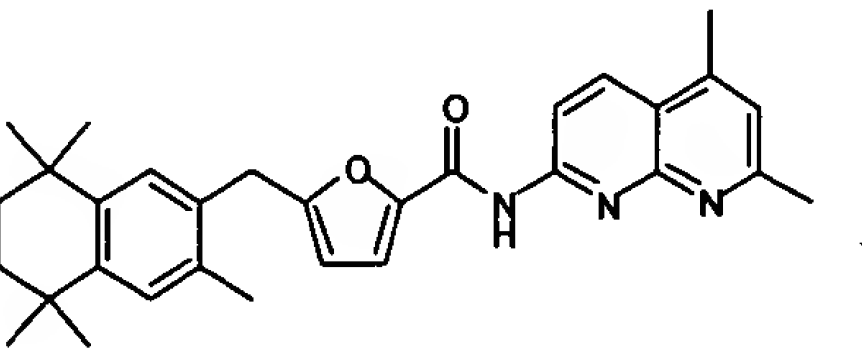
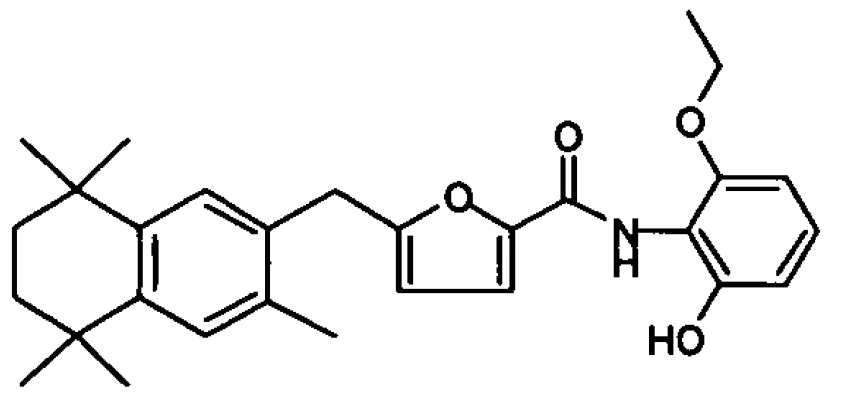
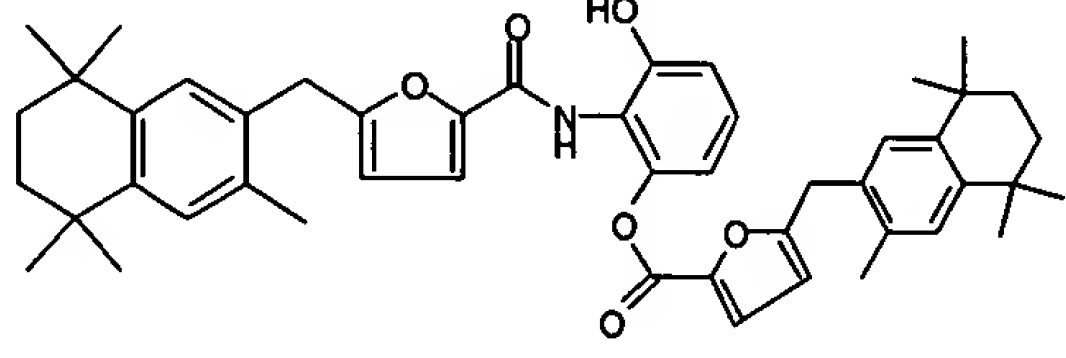
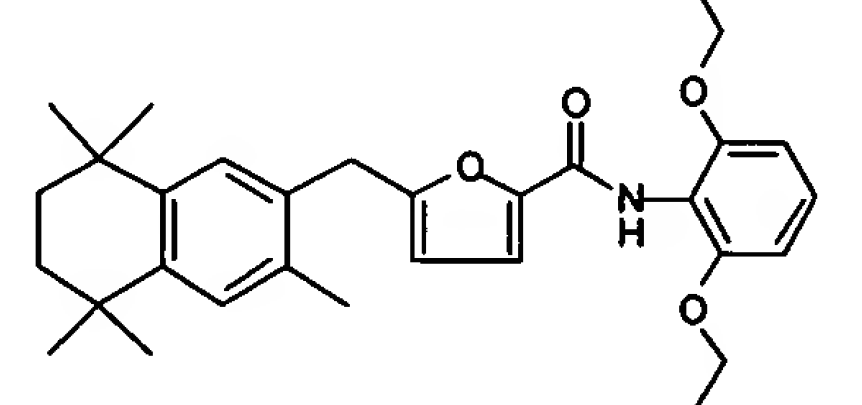
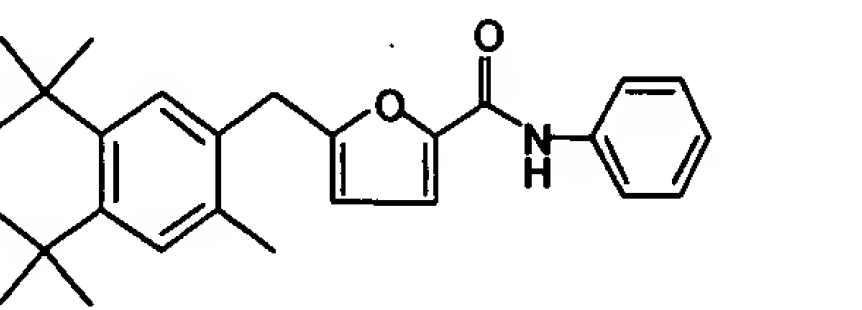
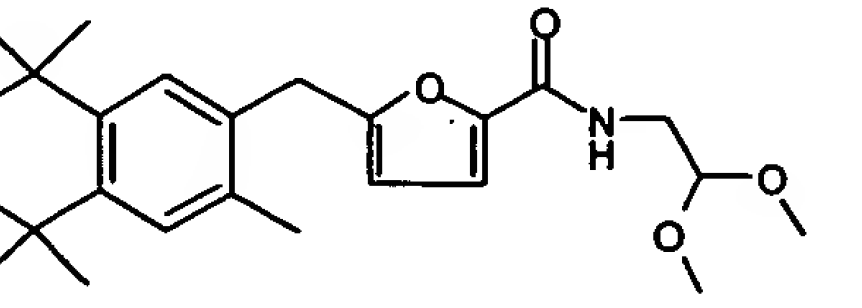
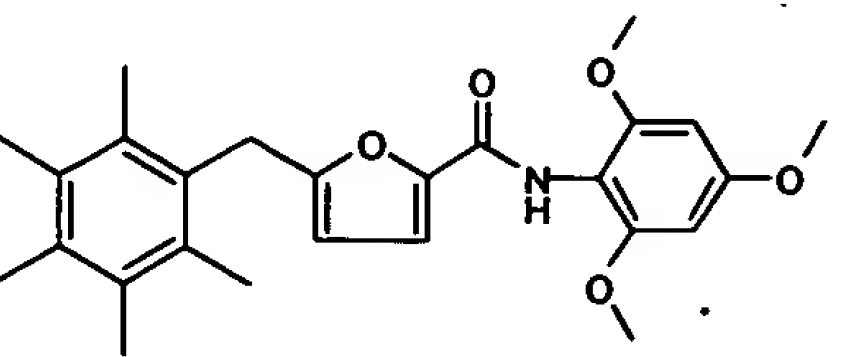
Cmpd. #	Structure	hGnRH IC50 (nM)	bGnRH IC50 (nM)	rGnRH IC50 (nM)	mGnRH IC50 (nM)	GnRH functional	GnRH Kb (nM)
143		1870					
144		230					
145		200					
146		>10000					
147		60		3	5		
148		12		1.6	1.3		
149		360					
150		80		8	13		

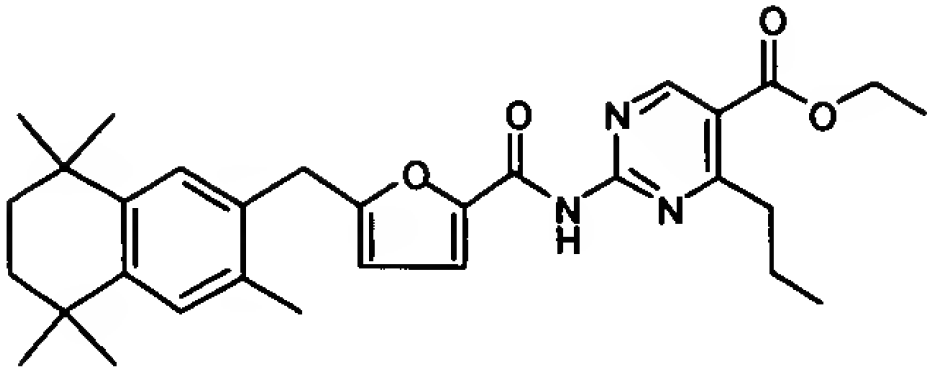
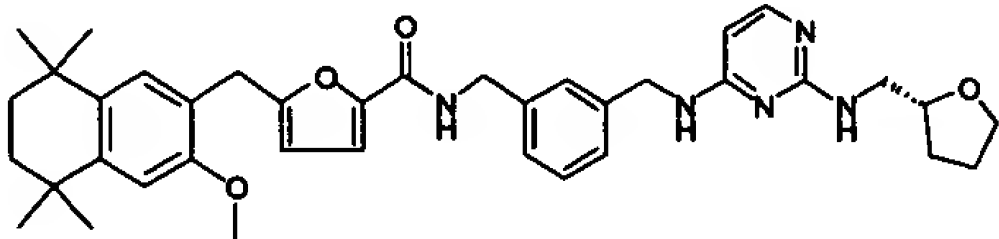
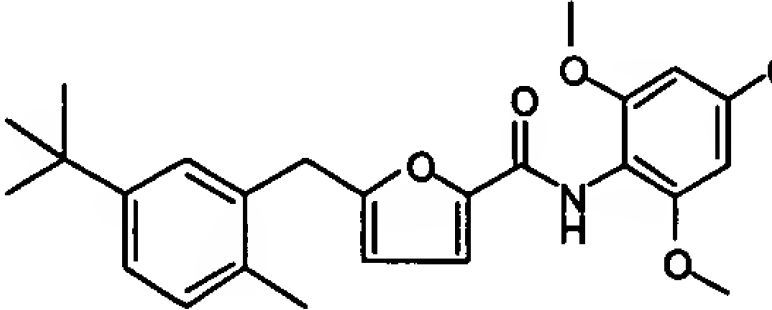
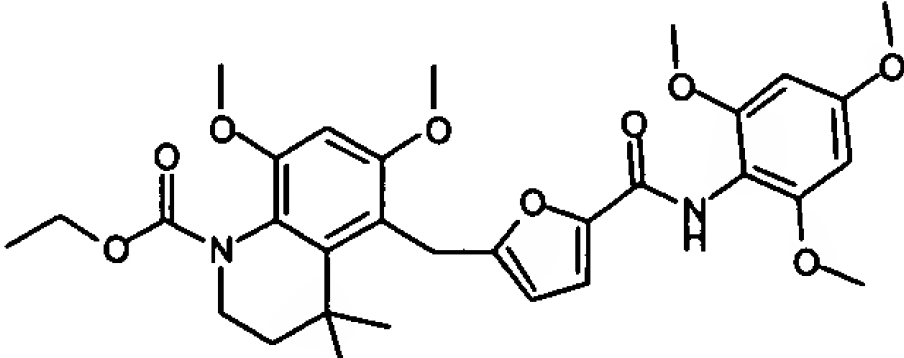
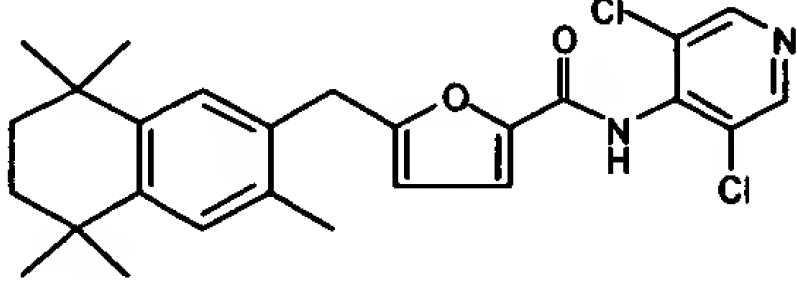
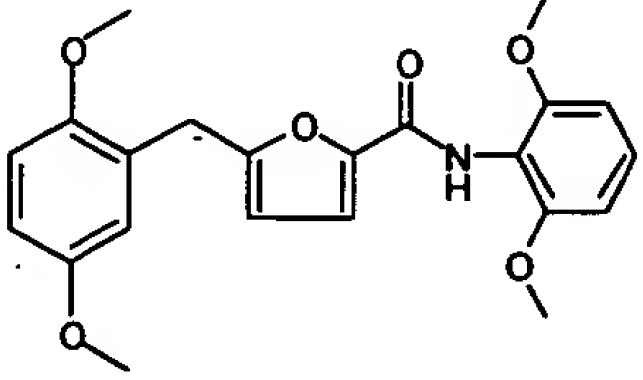
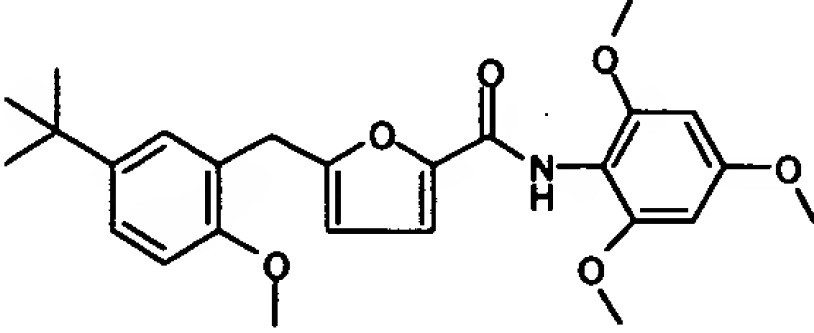
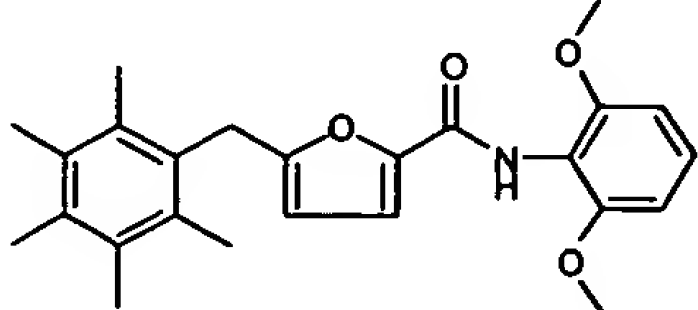
Cmpd. #	Structure	hGnRH IC50 (nM)	bGnRH IC50 (nM)	rGnRH IC50 (nM)	mGnRH IC50 (nM)	GnRH functional	GnRH Kb (nM)
160		390					
161		300					
162		>10000					
163		1960					
164		1970					
165		>10000 (solubility?)					
166		260					
167		7640					

Cmpd. #	Structure	hGnRH IC50 (nM)	bGnRH IC50 (nM)	rGnRH IC50 (nM)	mGnRH IC50 (nM)	GnRH functional	GnRH Kb (nM)
168		230		130	130		
169		12		7	5	Antagonist	17 (rat)
170		50		30	22		
171		1140					
172		3200					
173		2800					
174		220		385	570		
175		>10000					

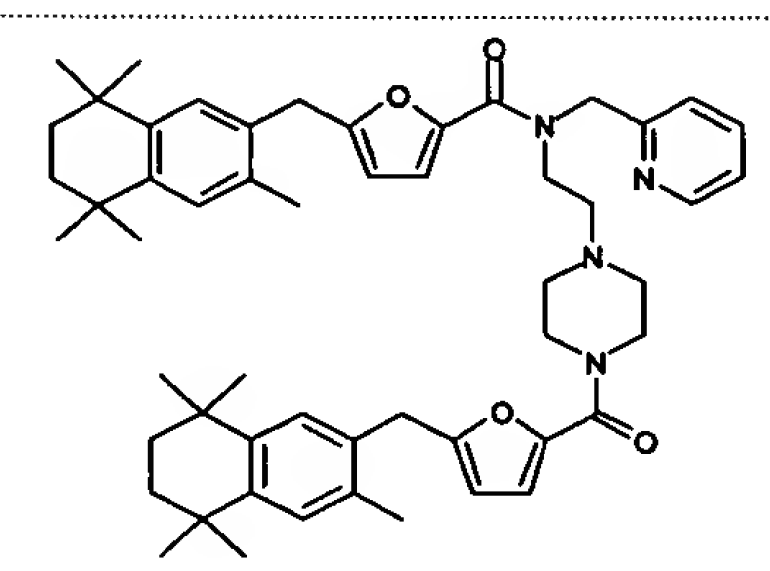
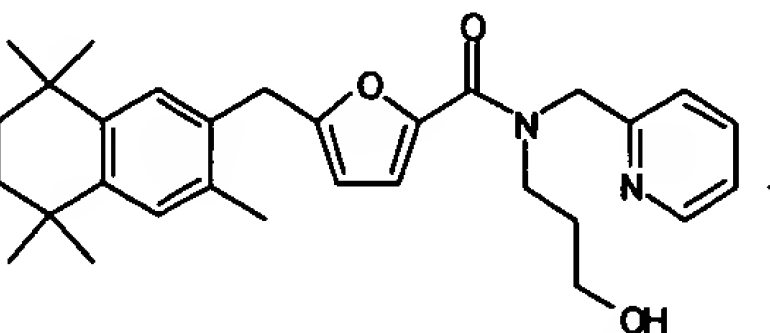
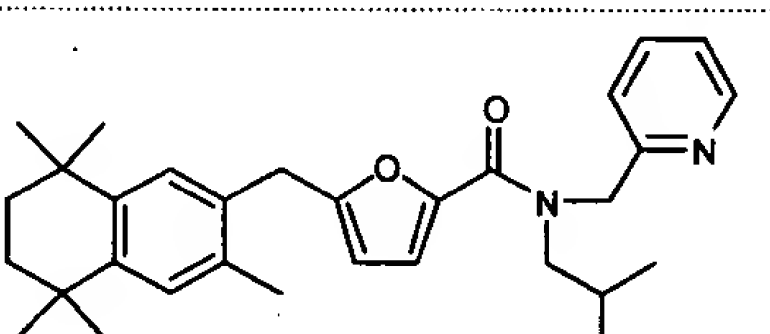
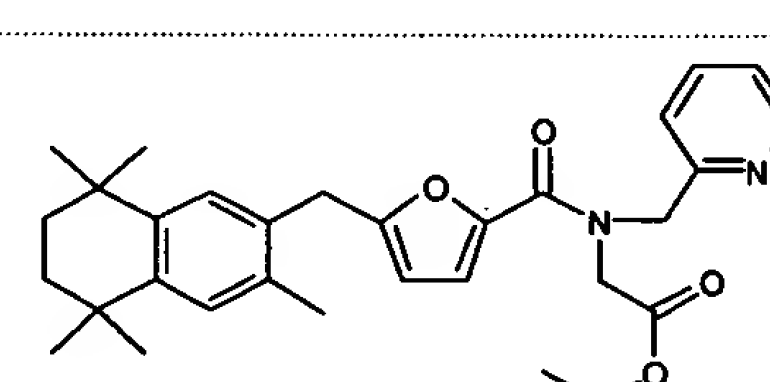
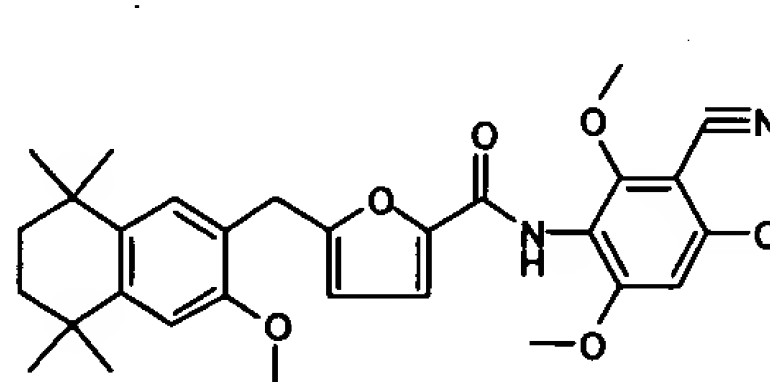
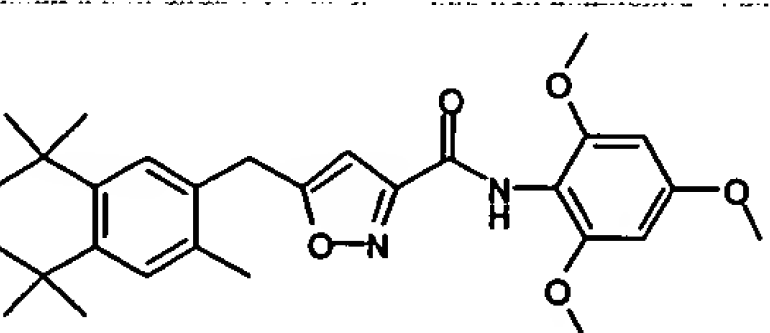
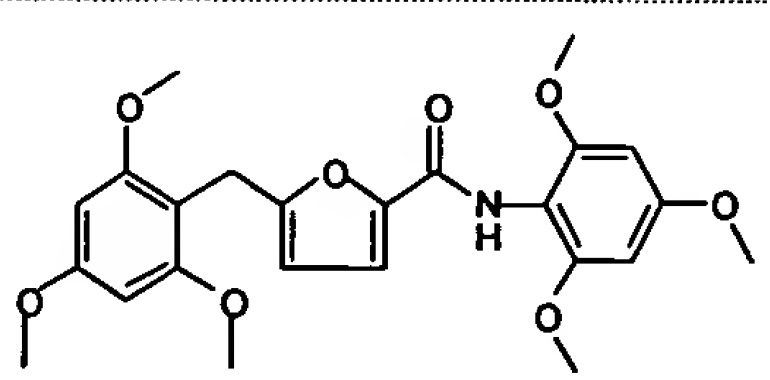
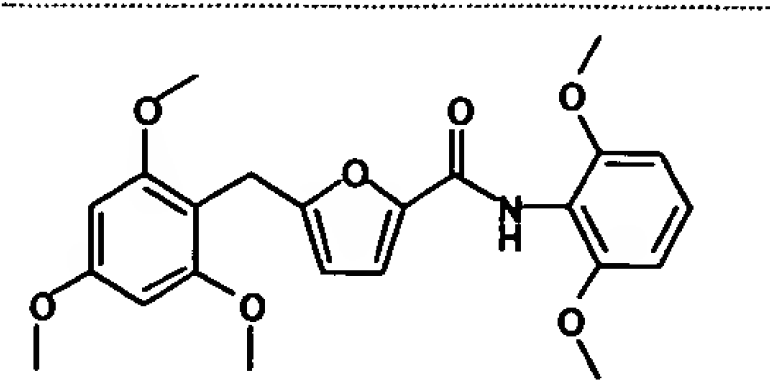
Cmpd. #	Structure	hGnRH IC50 (nM)	bGnRH IC50 (nM)	rGnRH IC50 (nM)	mGnRH IC50 (nM)	GnRH functional	GnRH Kb (nM)
176	 Chiral	54		3	4		
177		7.4		5.7	7.3	Antagonist	9 (rat)
178	 Chiral	185					
179		190					
180		350					
181		190					
182		430					
183		6.8		2.5	2.7	Antagonist	11.0 (rat)

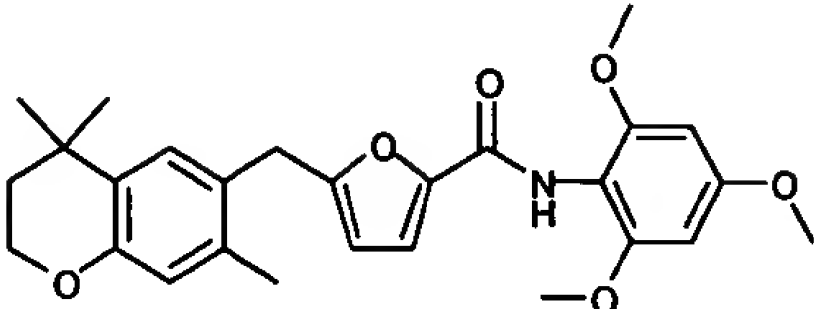
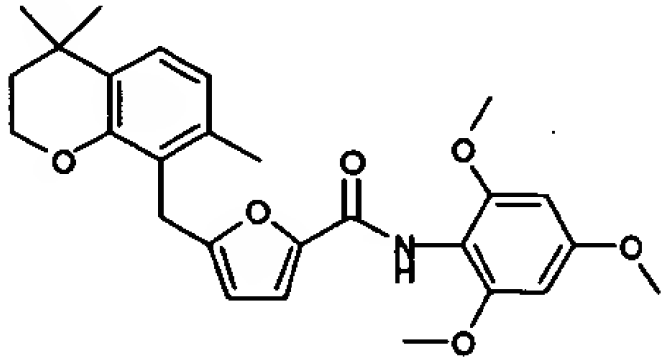
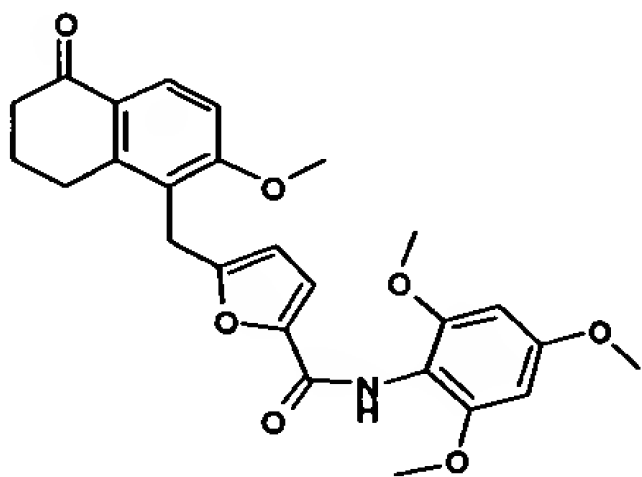
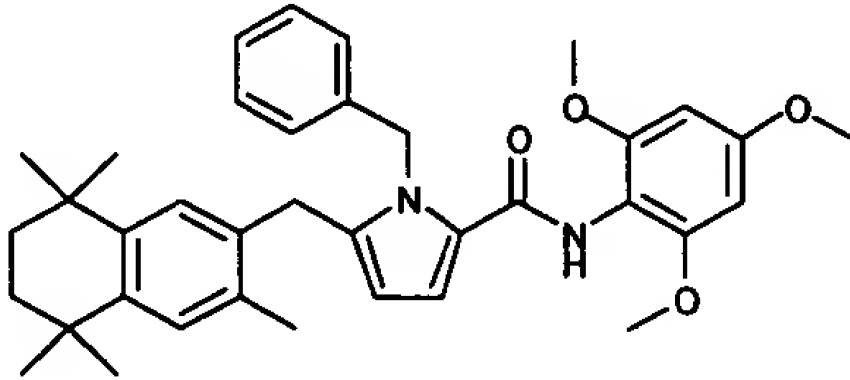
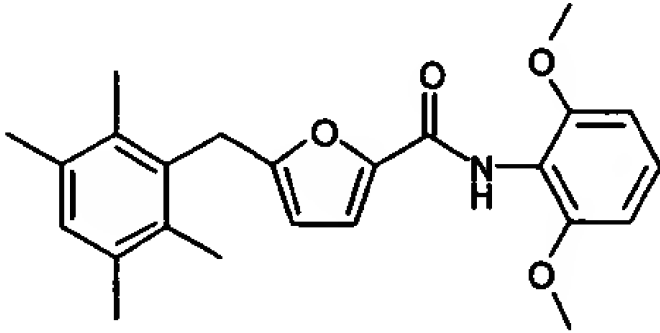
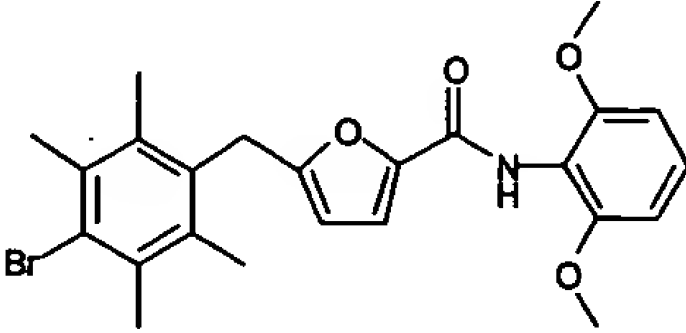
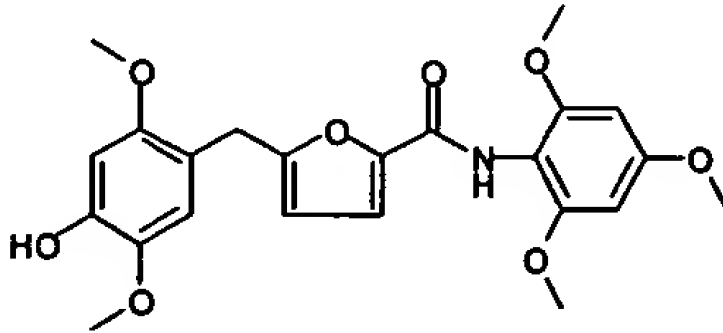
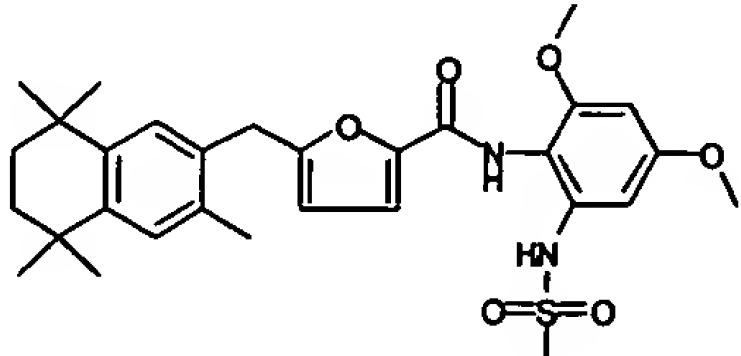
Cmpd. #	Structure	hGnRH IC50 (nM)	bGnRH IC50 (nM)	rGnRH IC50 (nM)	mGnRH IC50 (nM)	GnRH functional	GnRH Kb (nM)
184		560					
185		120		250	270		
186		7310					
187		9000					
188		290					
189		940					
190		420					

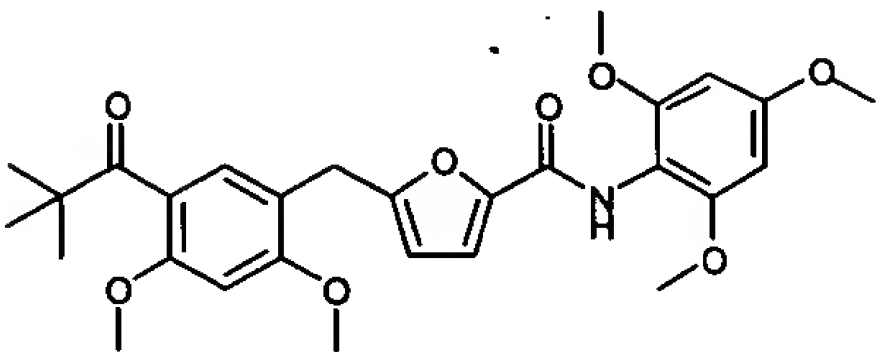
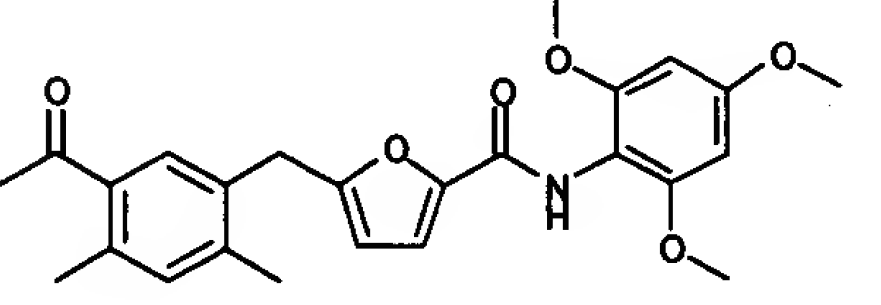
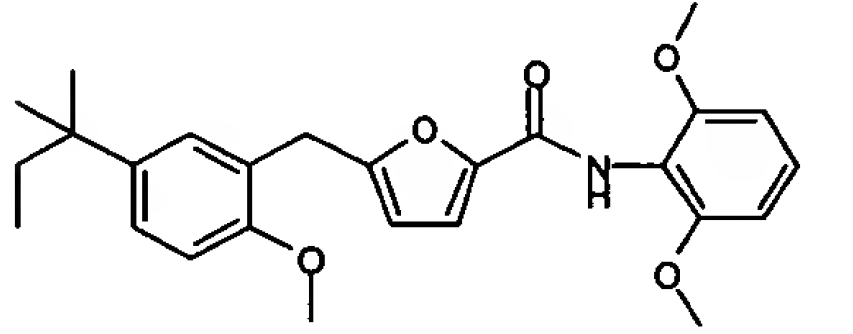
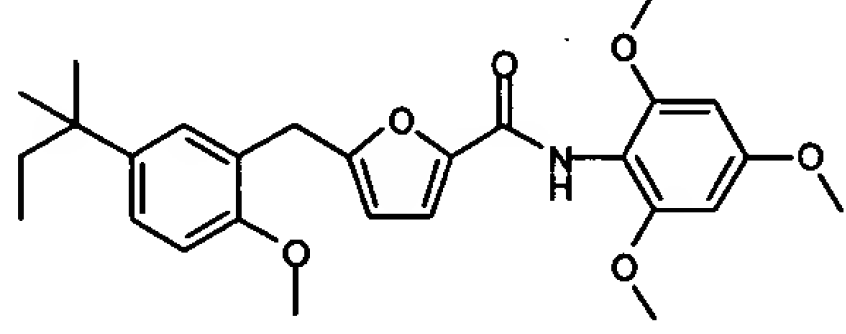
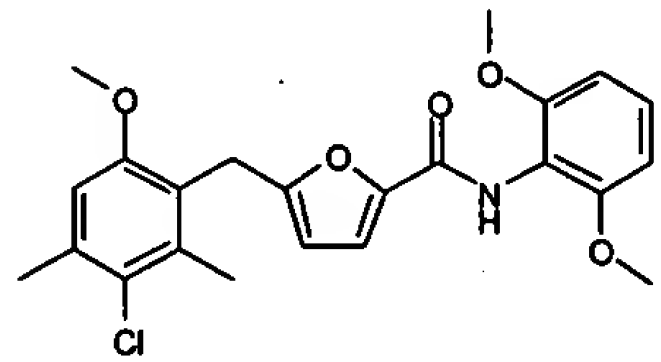
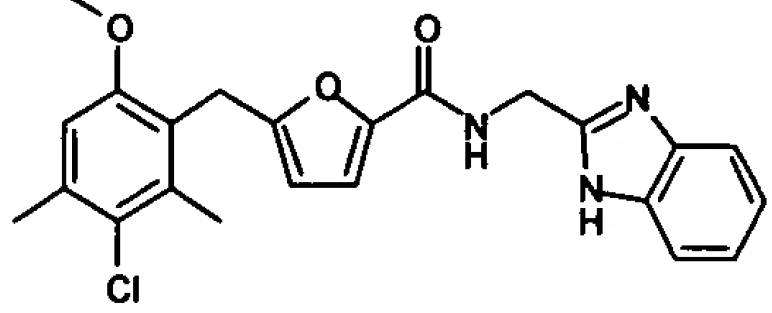
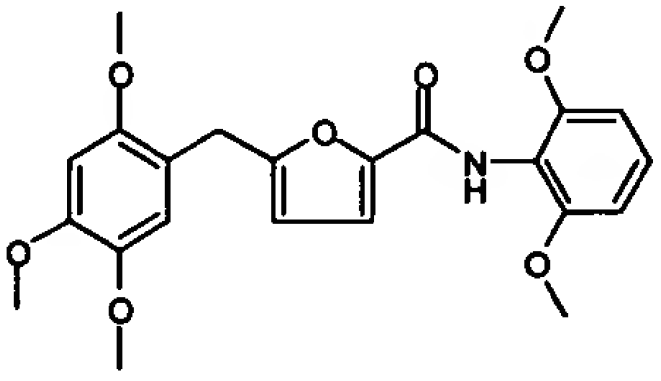
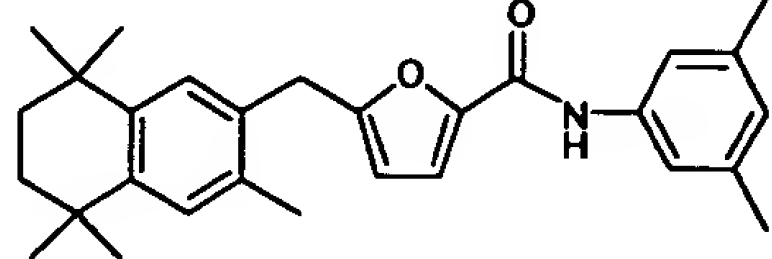
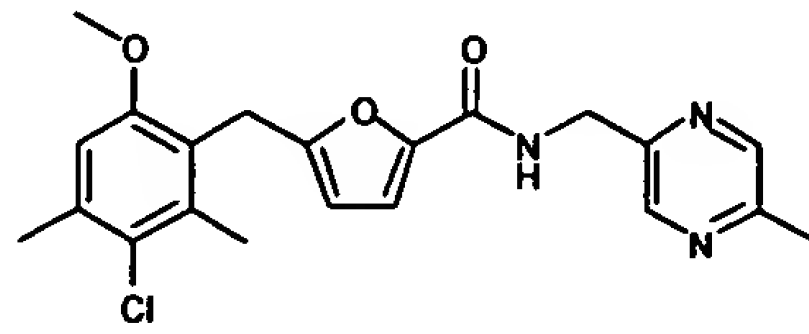
Cmpd. #	Structure	hGnRH IC50 (nM)	bGnRH IC50 (nM)	rGnRH IC50 (nM)	mGnRH IC50 (nM)	GnRH functional	GnRH Kb (nM)
190.5		350					
191		4240					
192		235					
193		>10000					
194		18		35	14		
195		320(Insol.>3 uM)					
196		760					
197		1040					

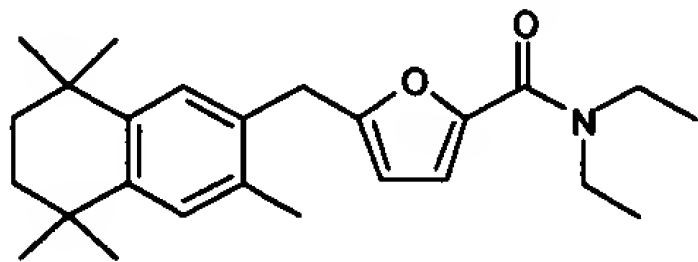
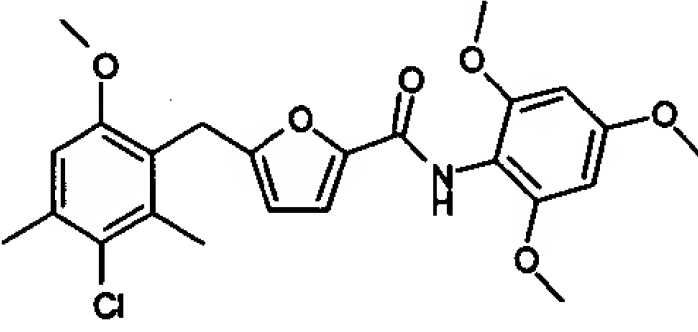
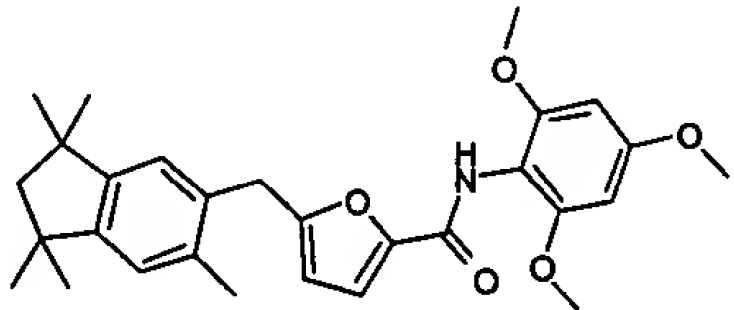
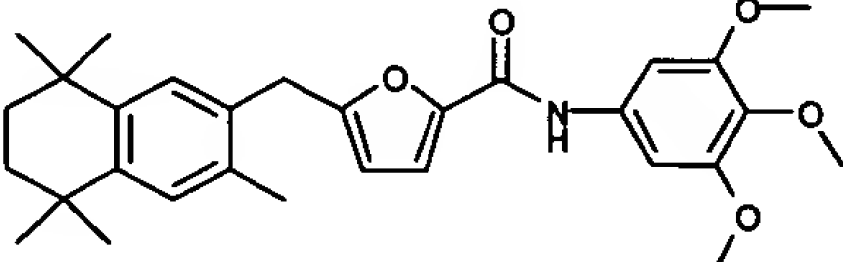
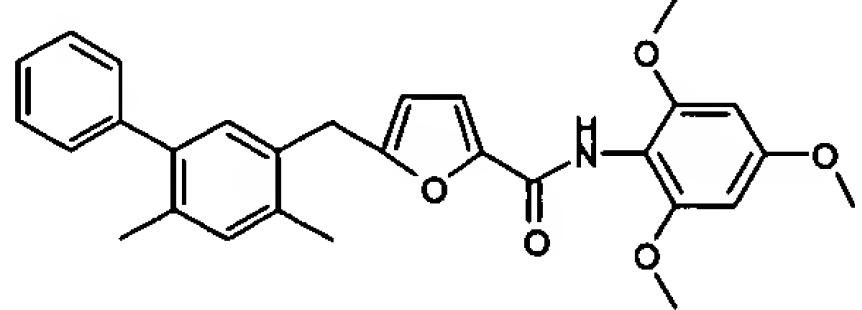
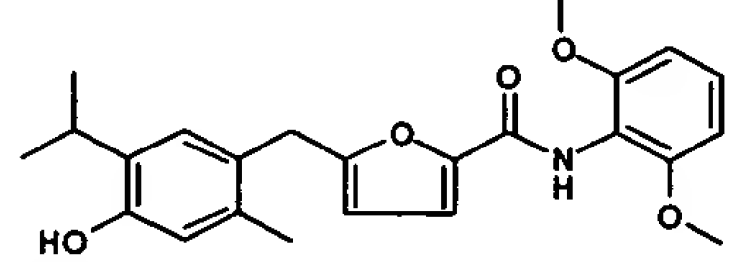
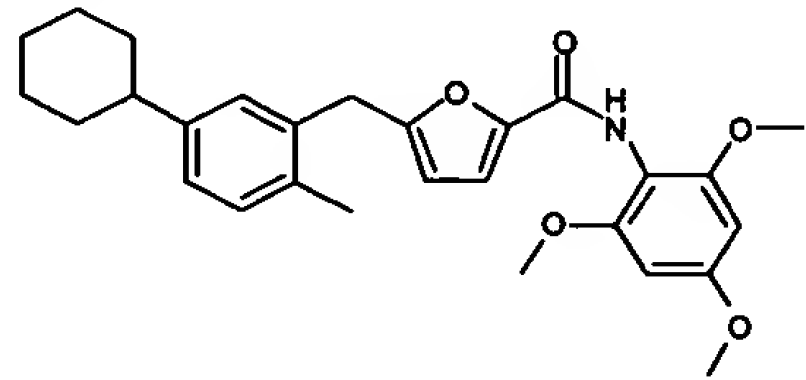
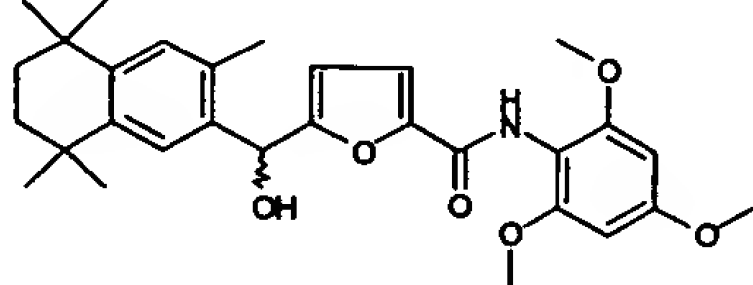
Cmpd. #	Structure	hGnRH IC50 (nM)	bGnRH IC50 (nM)	rGnRH IC50 (nM)	mGnRH IC50 (nM)	GnRH functional	GnRH Kb (nM)
198		780(Insol.>3 uM)					
199	 Chiral	92		9.6	15		
200		42		2.8	13	Antagonist	0.9 nM (rat)
200.5		1230					
201		380					
202		1425					
203		130		58	62		
204		1270					

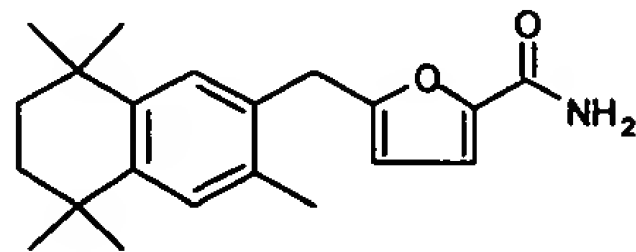
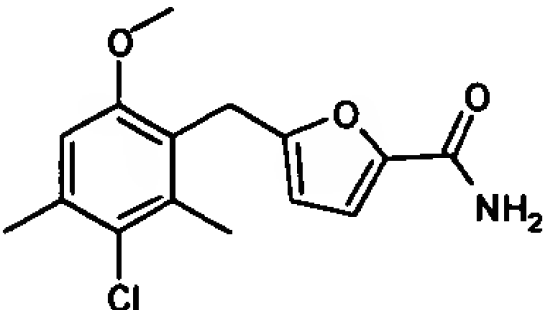
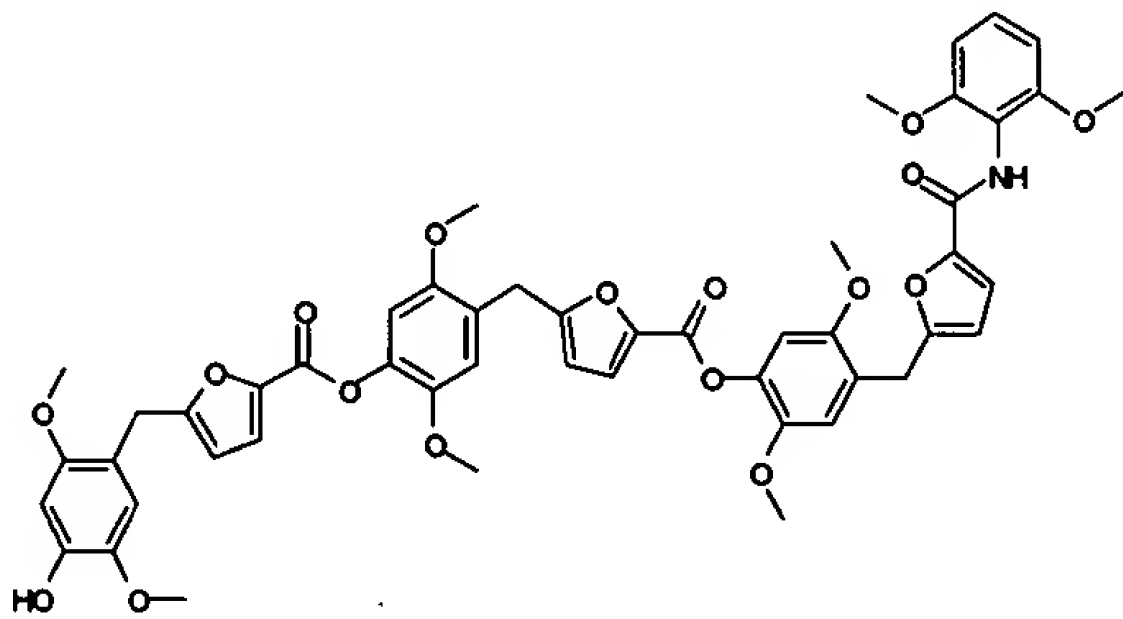
Cmpd. #	Structure	hGnRH IC50 (nM)	bGnRH IC50 (nM)	rGnRH IC50 (nM)	mGnRH IC50 (nM)	GnRH functional	GnRH Kb (nM)
205		>10000					
206		52		23	83		
207		200					
208		8620					
209		4380					
210		12000					
211		1320					
212		80		12	62		

Cmpd. #	Structure	hGnRH IC50 (nM)	bGnRH IC50 (nM)	rGnRH IC50 (nM)	mGnRH IC50 (nM)	GnRH functional	GnRH Kb (nM)
213		>10000					
214		>10000					
215		7800					
216		4400					
217		29		21	11		
218		6460					
219		875					
220		630					

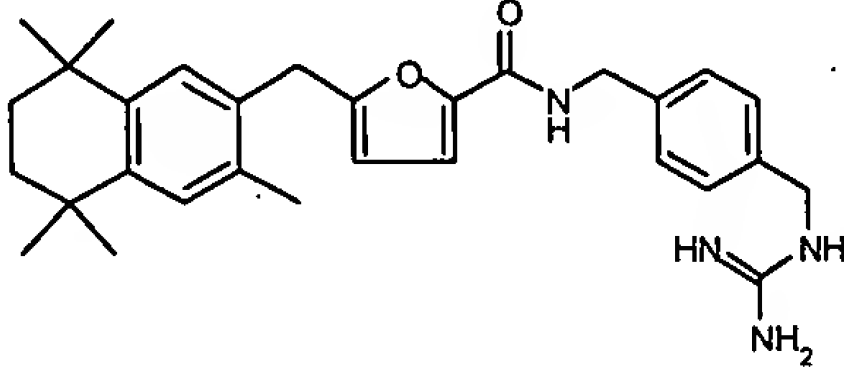
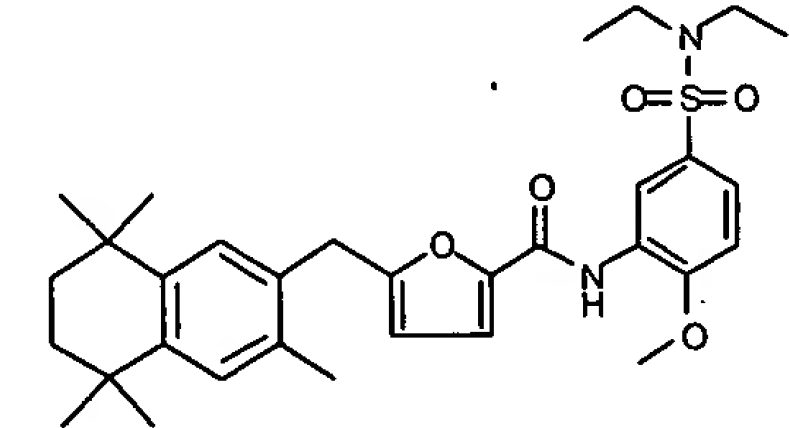
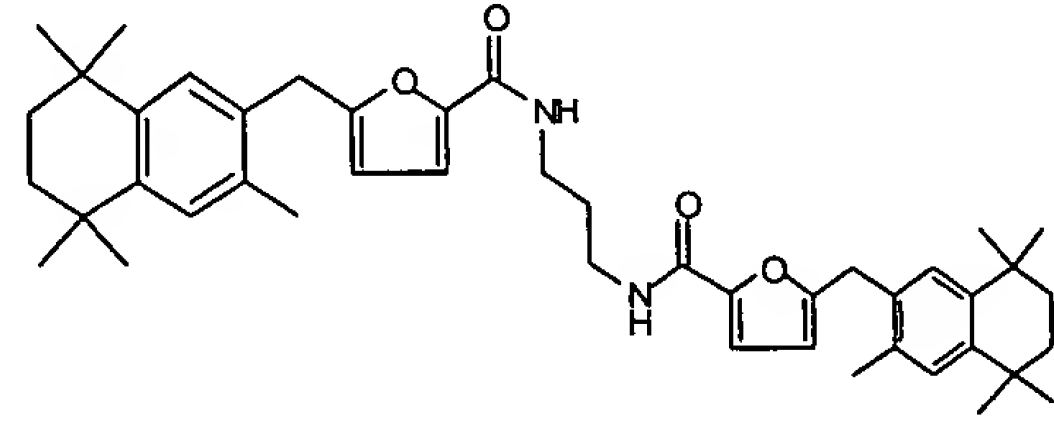
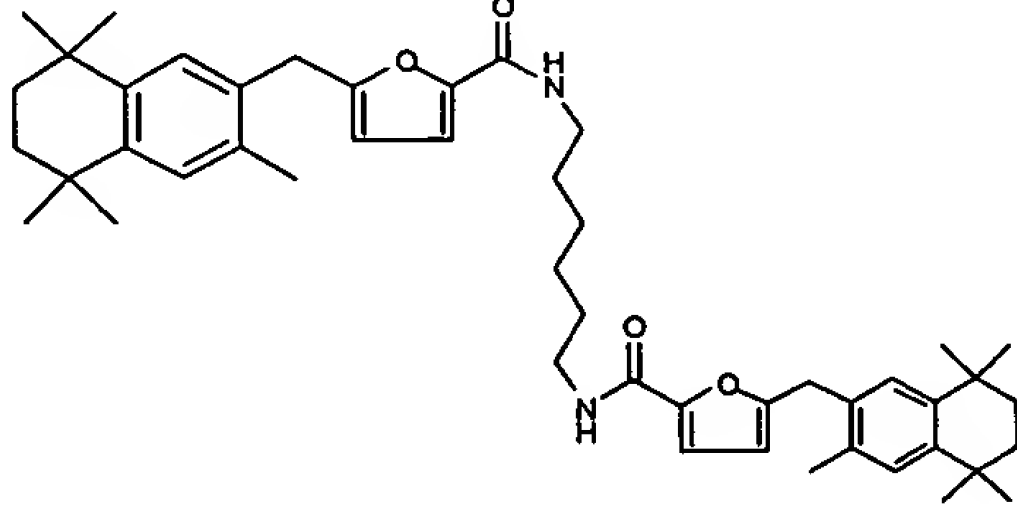
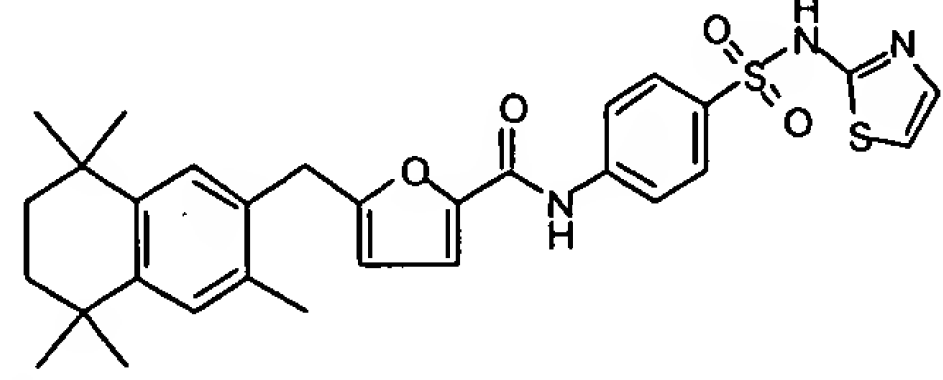
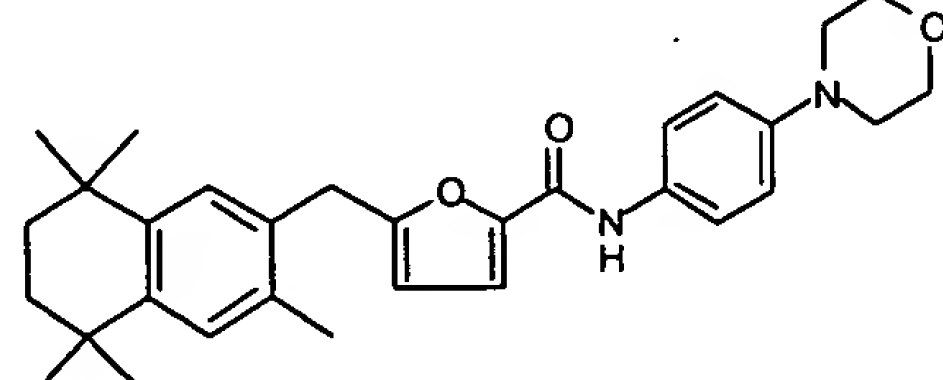
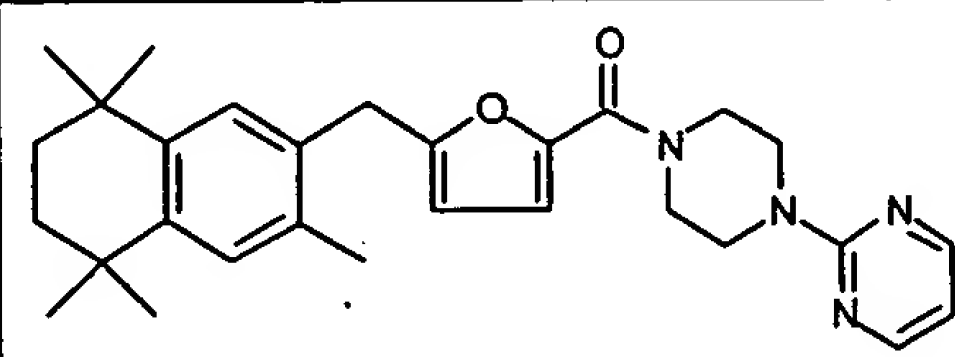
Cmpd. #	Structure	hGnRH IC50 (nM)	bGnRH IC50 (nM)	rGnRH IC50 (nM)	mGnRH IC50 (nM)	GnRH functional	GnRH Kb (nM)
221		15		11	9.2		
222		110		78	65		
223		1660					
224		>10000					
225		174		1510	850		
226		240		2130	720		
227		>10000					
228		27		57	14		

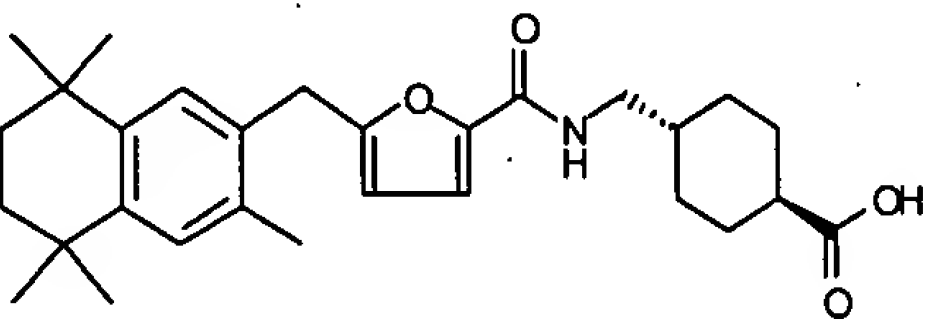
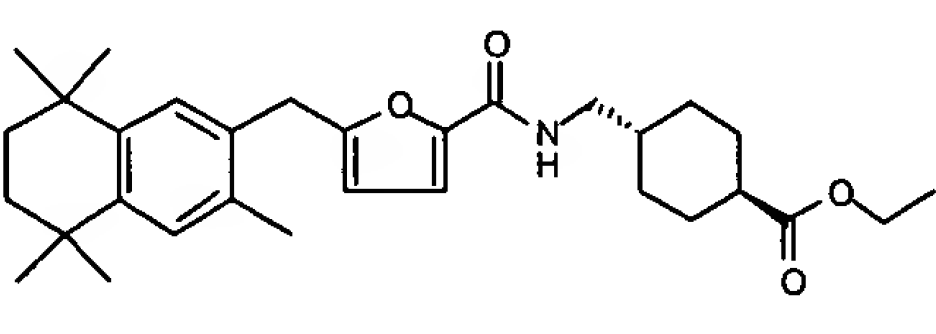
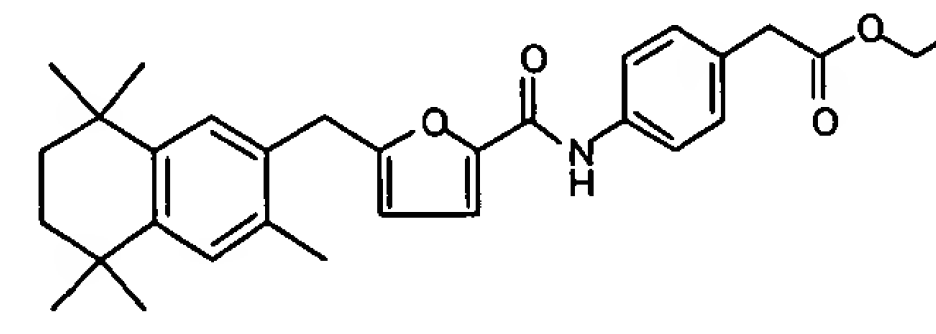
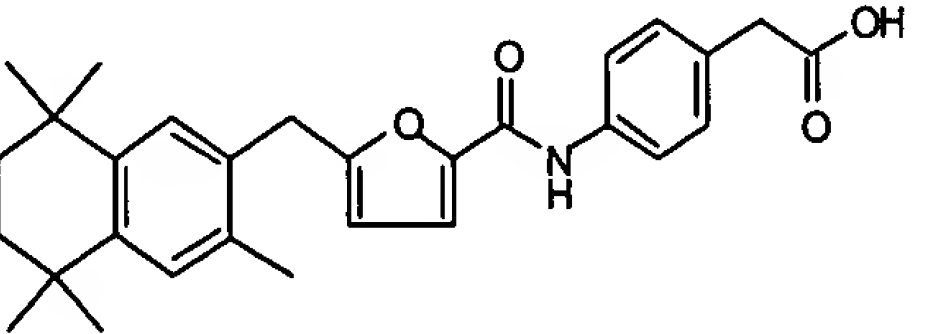
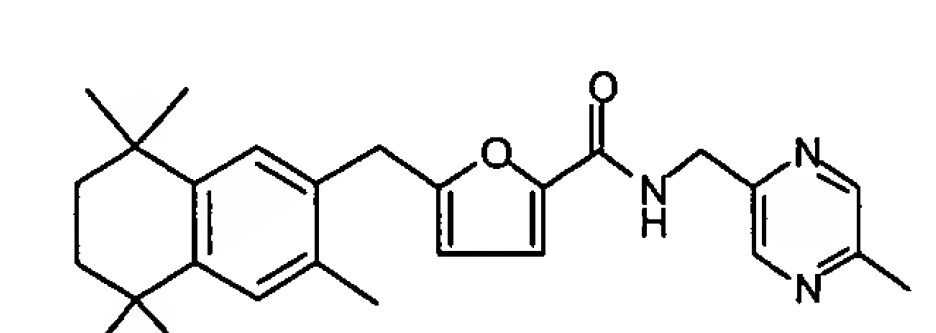
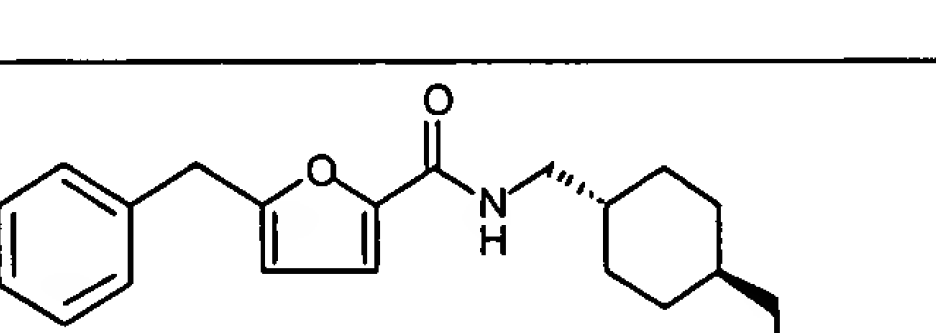
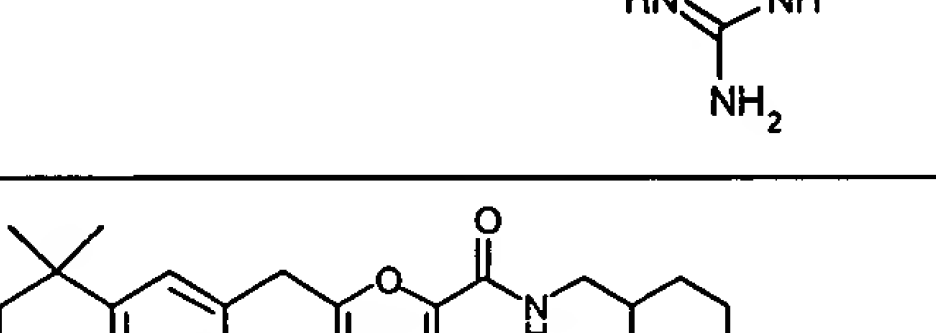
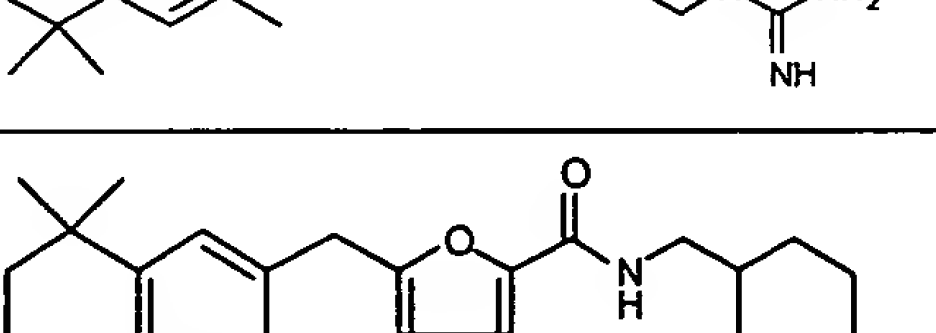
Cmpd. #	Structure	hGnRH IC50 (nM)	bGnRH IC50 (nM)	rGnRH IC50 (nM)	mGnRH IC50 (nM)	GnRH functional	GnRH Kb (nM)
229		250					
230		580					
231		5		51	24		
232		19		5.7	9		
233		60		51	68		
234		>10000					
235		3720					
236		903					
237		>10000					

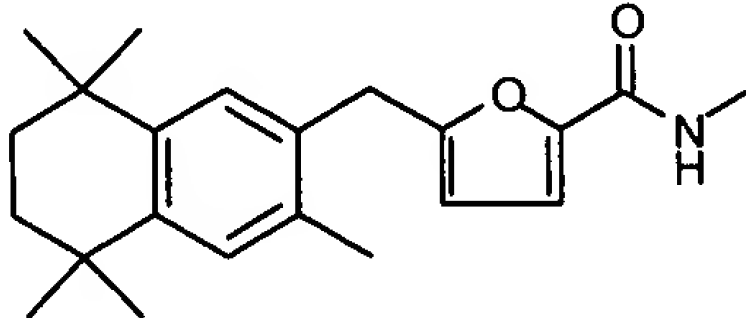
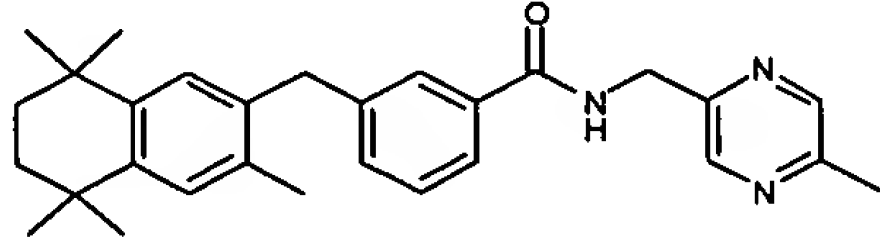
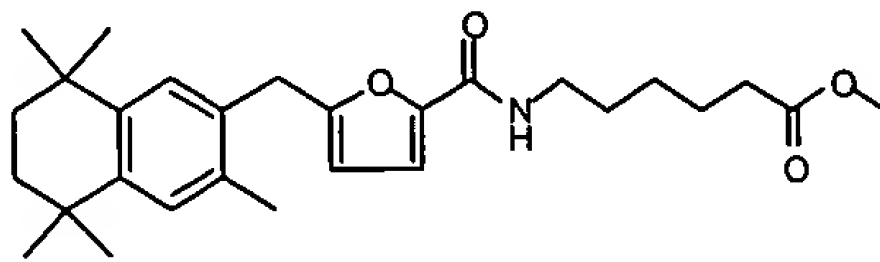
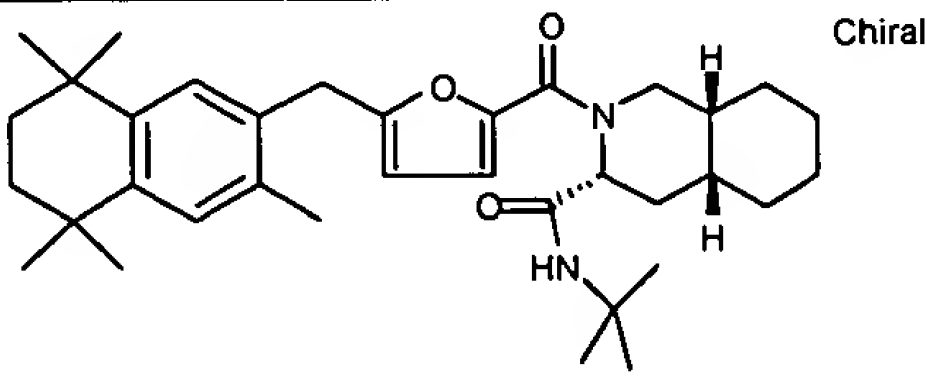
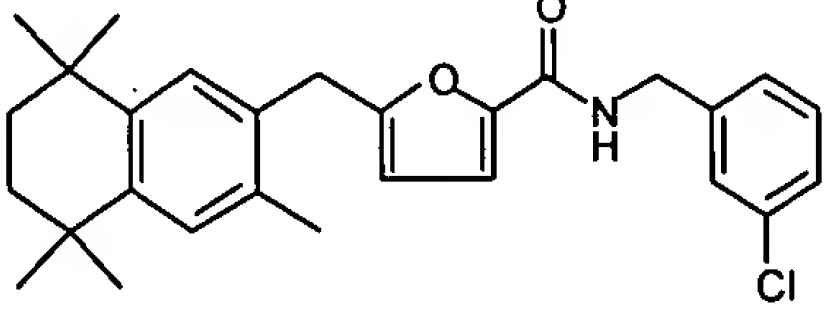
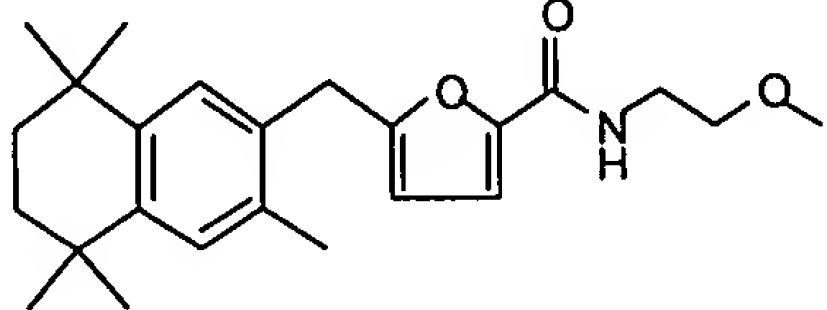
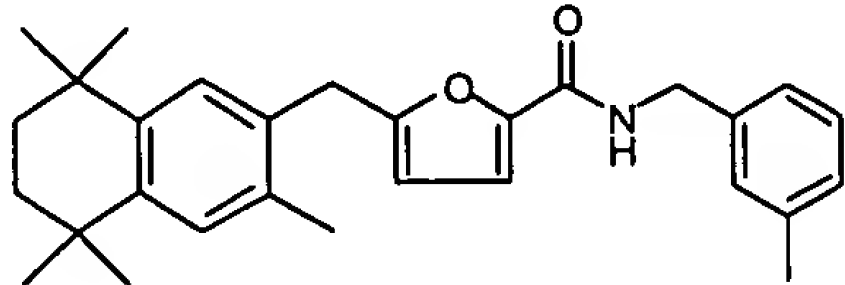
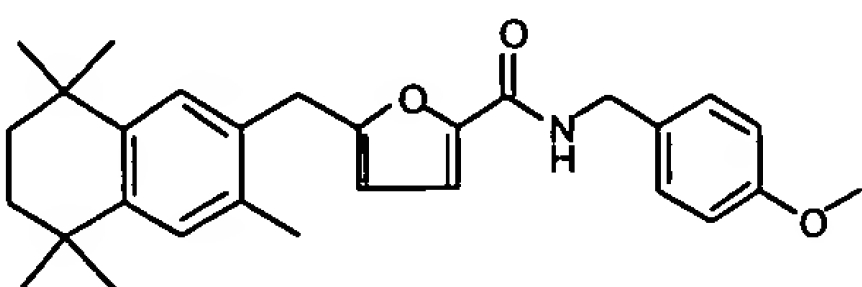
Cmpd. #	Structure	hGnRH IC50 (nM)	bGnRH IC50 (nM)	rGnRH IC50 (nM)	mGnRH IC50 (nM)	GnRH functional	GnRH Kb (nM)
238		7830					
299		80		290	105		
300		3.2			3.1		
301		45			38		
302		167		220	119		
303		320					
304		303					
AXC- 07047		1410					

Cmpd. #	Structure	hGnRH IC50 (nM)	bGnRH IC50 (nM)	rGnRH IC50 (nM)	mGnRH IC50 (nM)	GnRH functional	GnRH Kb (nM)
AXC- 06675		2000		2800			
AXC- 07450		>10000					
AXC- 07350		>10000					

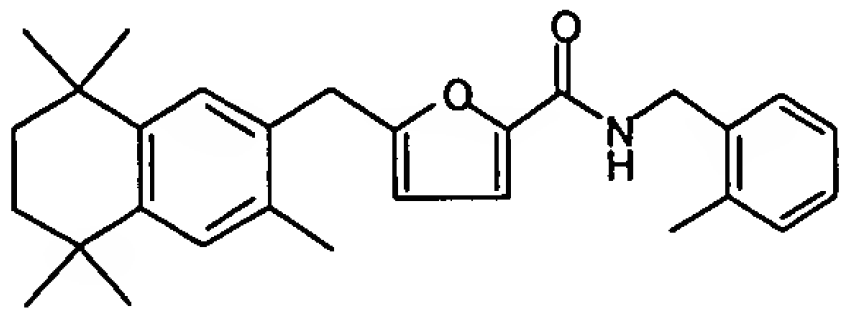
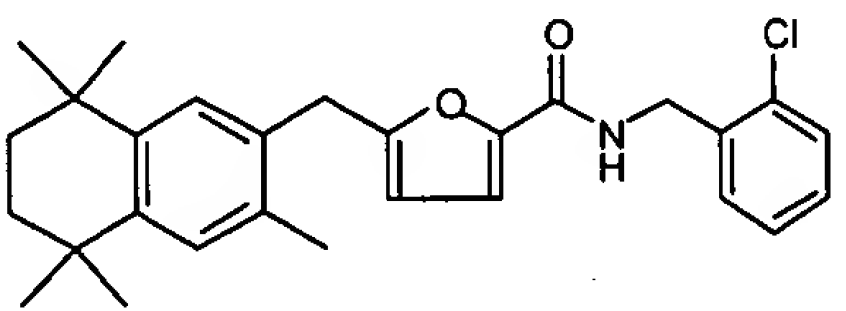
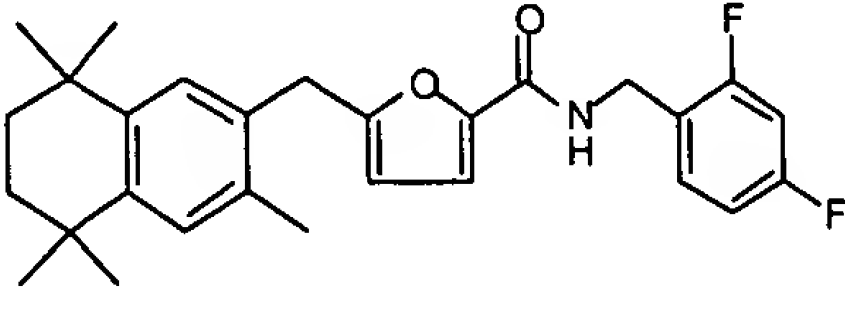
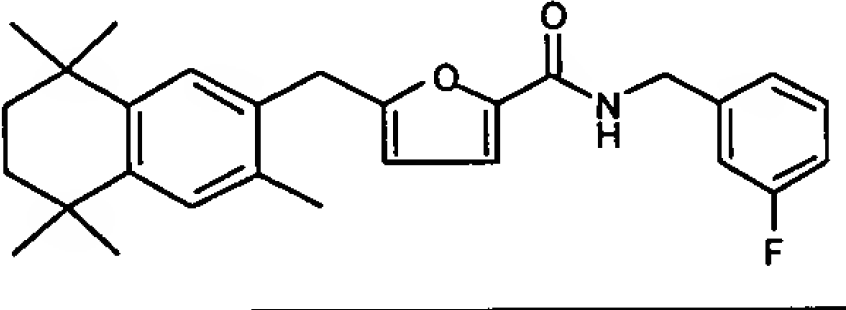
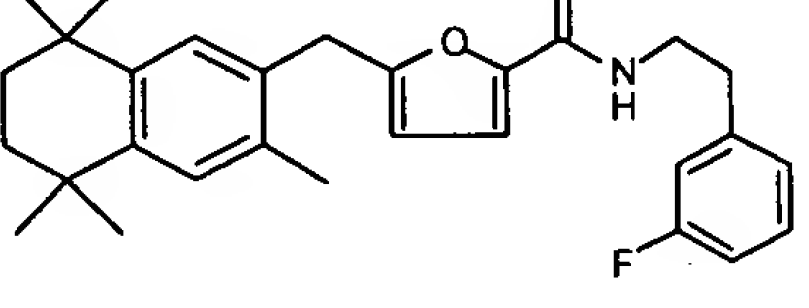
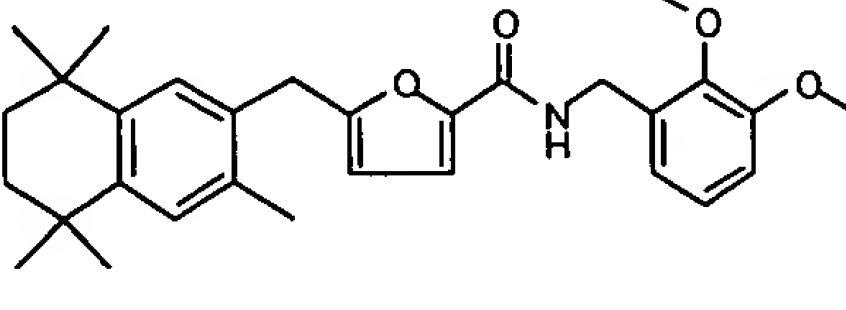
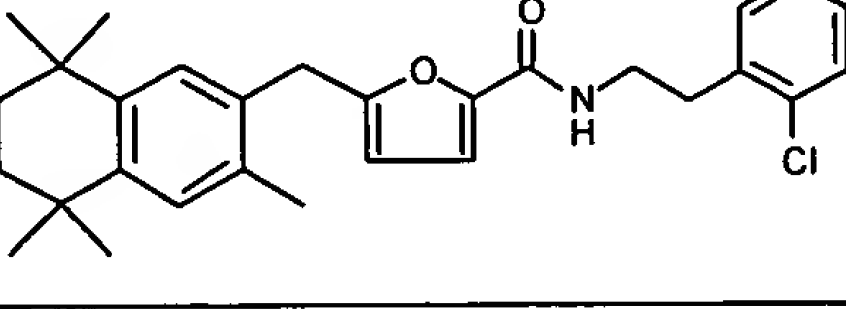
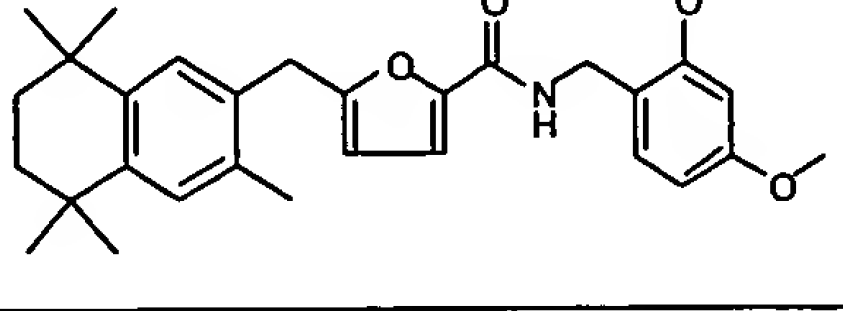
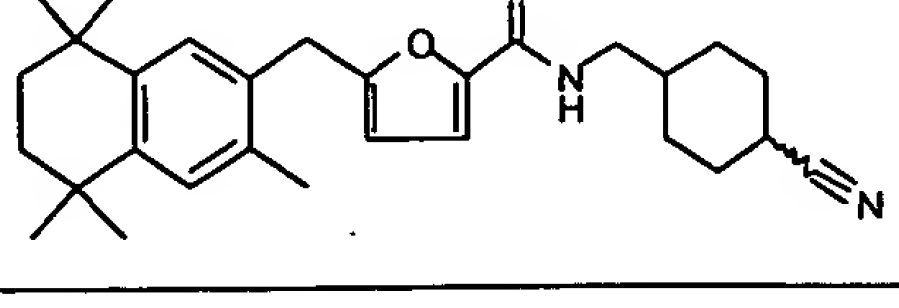
Cmpd. #	Structure	5-HT7 KI (nM)	5-HT2a KI (nM)	D2 DA KI (nM)	IL8 IC50 (nM)	NPY Y1 IC50 (nM)	Glucagon IC50 (nM)	GLP-1 IC50 (nM)
9								
10		1270	1200	550	6400	9700	6600	
21								
11		1800		150	>10000	2550	3200	>10000
22								
23								
24								

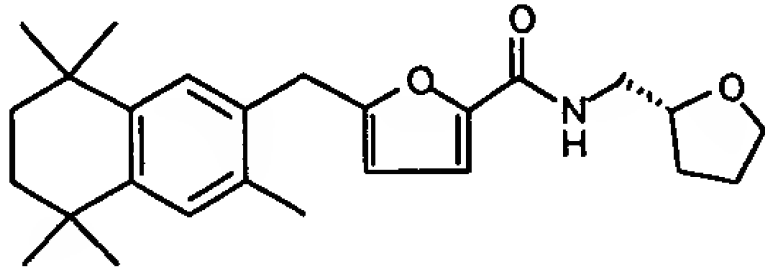
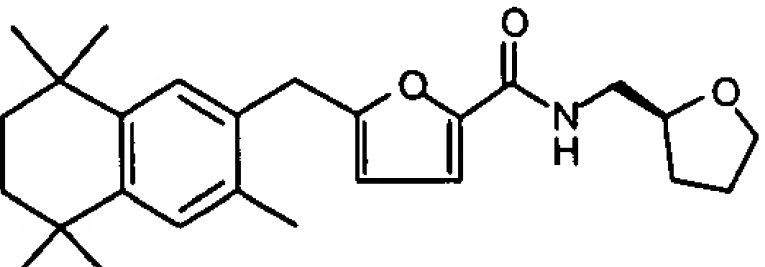
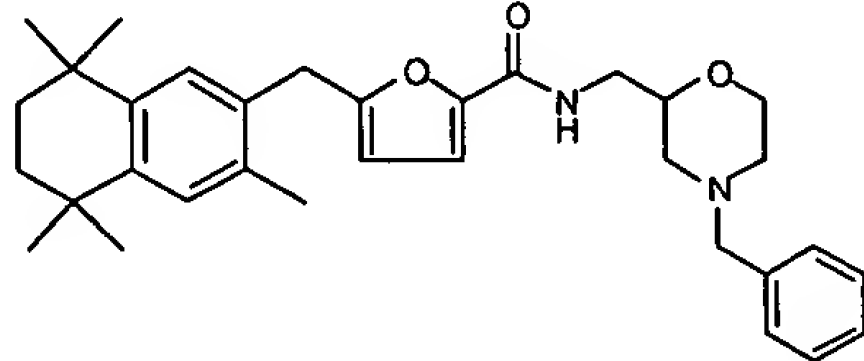
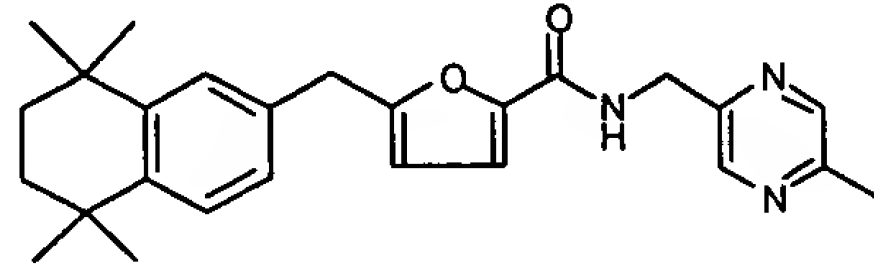
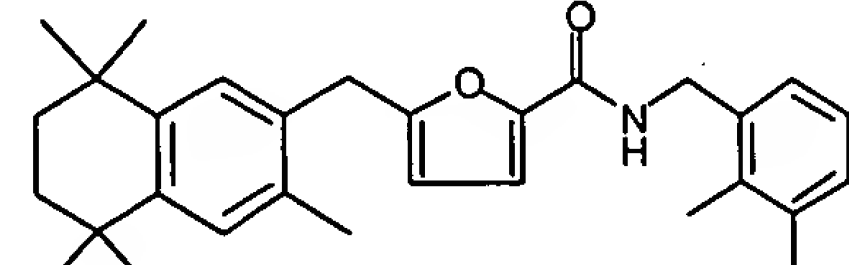
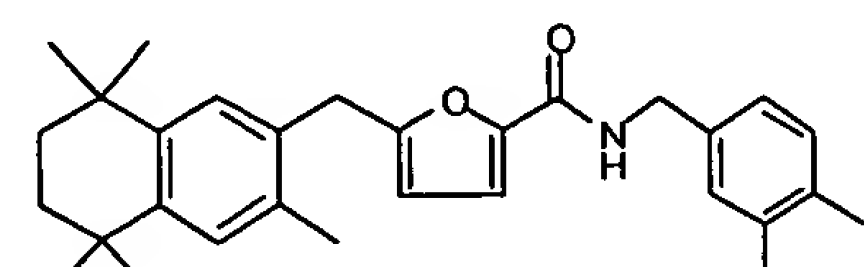
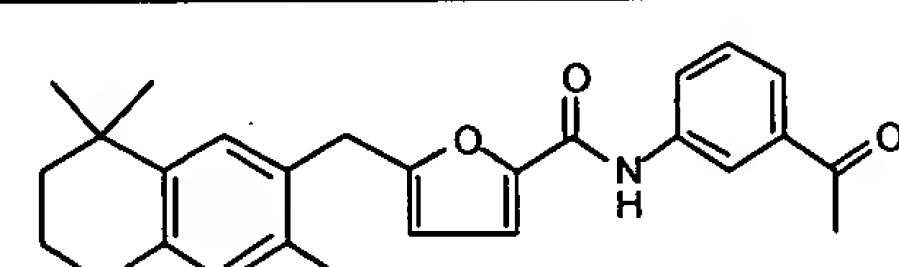
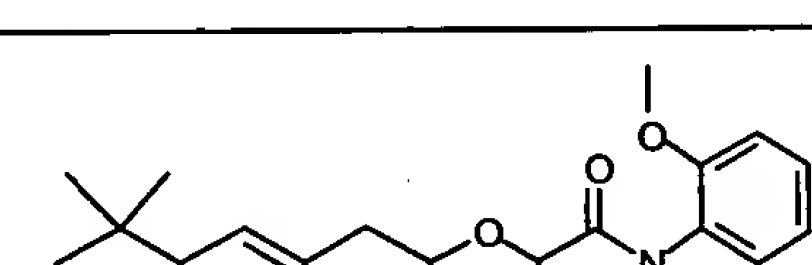
Cmpd. #	Structure	5-HT7 Ki (nM)	5-HT2a Ki (nM)	D2 DA Ki (nM)	IL8 IC50 (nM)	NPY Y1 IC50 (nM)	Glucagon IC50 (nM)	GLP-1 IC50 (nM)
25								
26								
27								
28								
29								
30								
30.5								

Cmpd. #	Structure	5-HT7 Ki (nM)	5-HT2a Ki (nM)	D2 DA Ki (nM)	IL8 IC50 (nM)	NPY Y1 IC50 (nM)	Glucagon IC50 (nM)	GLP-1 IC50 (nM)
12		>4500	>4000	1400	>10000	>10000	10000	
31								
32								
33								
13		>4400		>3200	>10000	>10000	7900	>10000
34								
35								
36								

Cmpd. #	Structure	5-HT7 Ki (nM)	5-HT2a Ki (nM)	D2 DA Ki (nM)	IL8 IC50 (nM)	NPY Y1 IC50 (nM)	Glucagon IC50 (nM)	GLP-1 IC50 (nM)
37								
37.5								
38								
38.5	 Chiral							
39								
40								
41								
42								

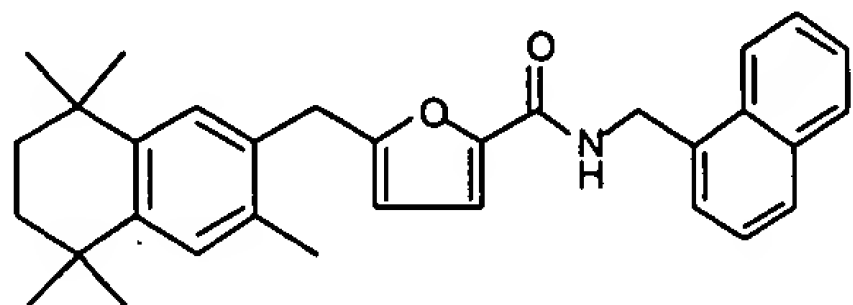
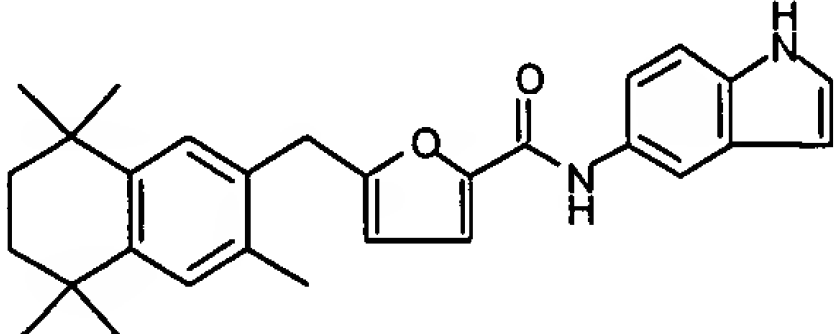
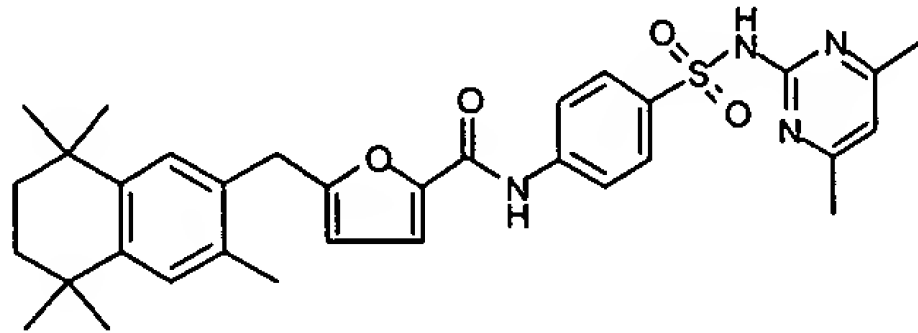
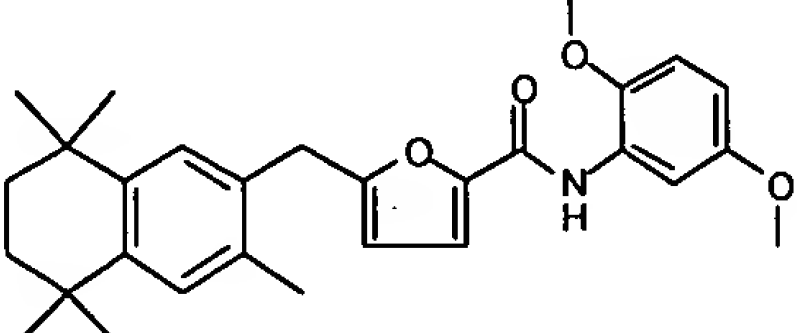
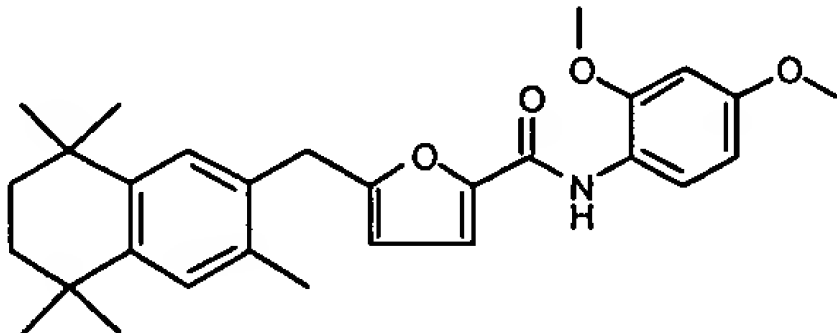
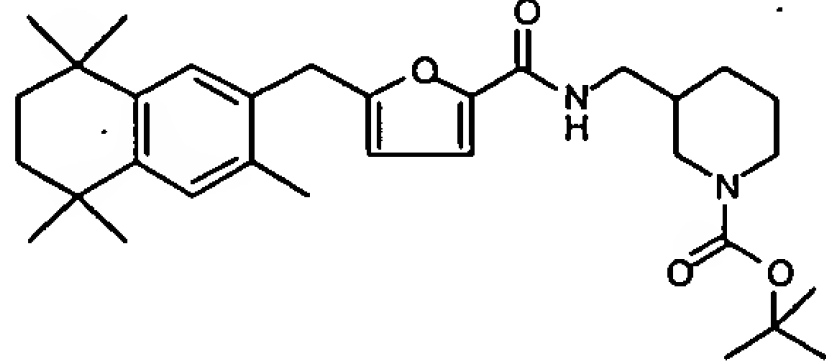
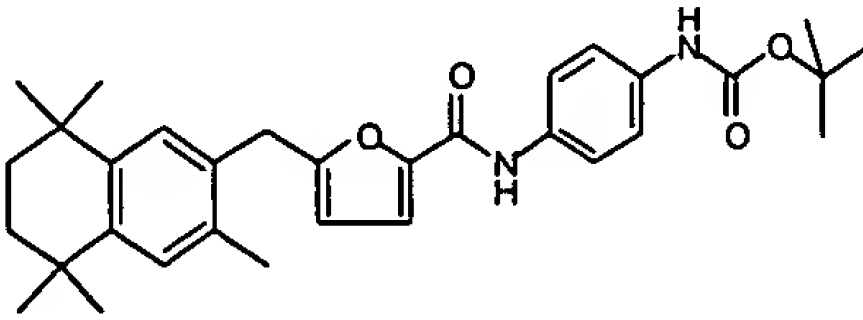
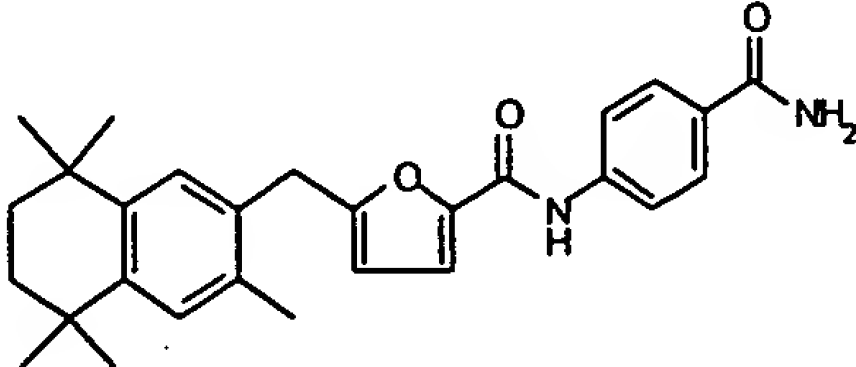
Cmpd. #	Structure	5-HT7 Ki (nM)	5-HT2a Ki (nM)	D2 DA Ki (nM)	IL8 IC50 (nM)	NPY Y1 IC50 (nM)	Glucagon IC50 (nM)	GLP-1 IC50 (nM)
43								
44								
45								
46								
47								
48		>4400		>3200	>10000	>10000	6300	>10000
14								
49								

Cmpd. #	Structure	5-HT7 Ki (nM)	5-HT2a Ki (nM)	D2 DA Ki (nM)	IL8 IC50 (nM)	NPY Y1 IC50 (nM)	Glucagon IC50 (nM)	GLP-1 IC50 (nM)
50								
51								
52								
53								
54								
15								
55								
56								
16								

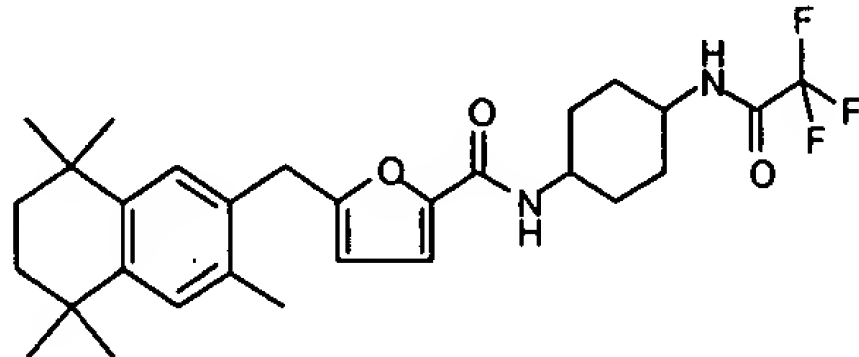
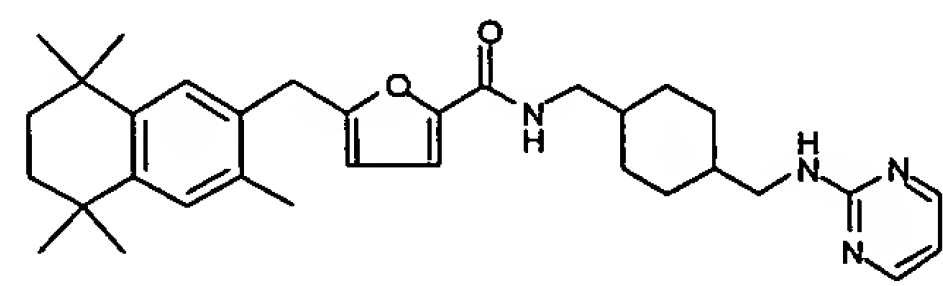
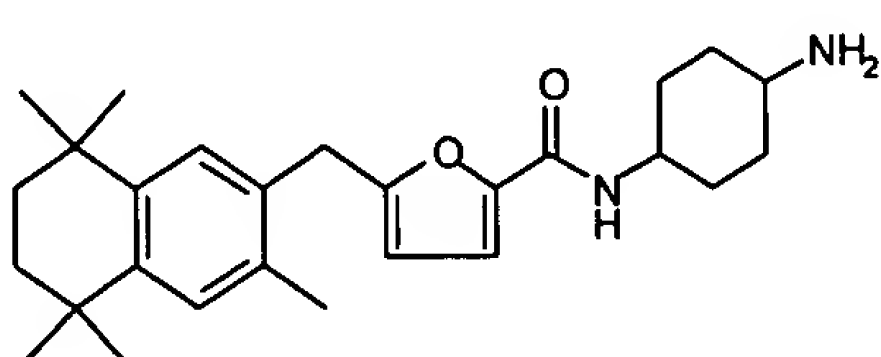
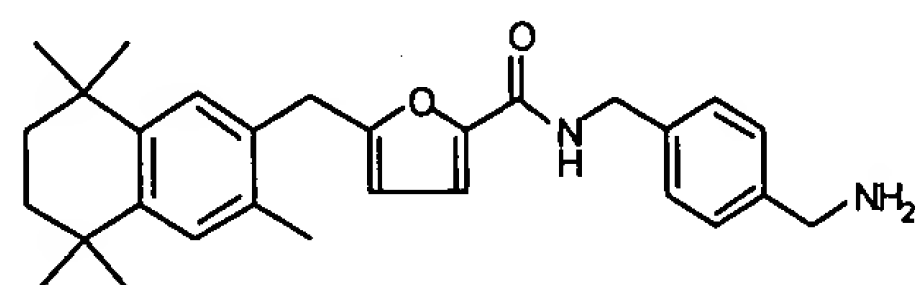
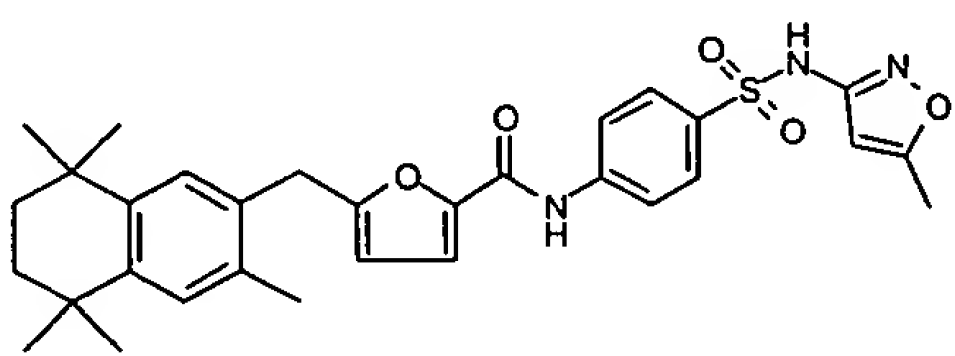
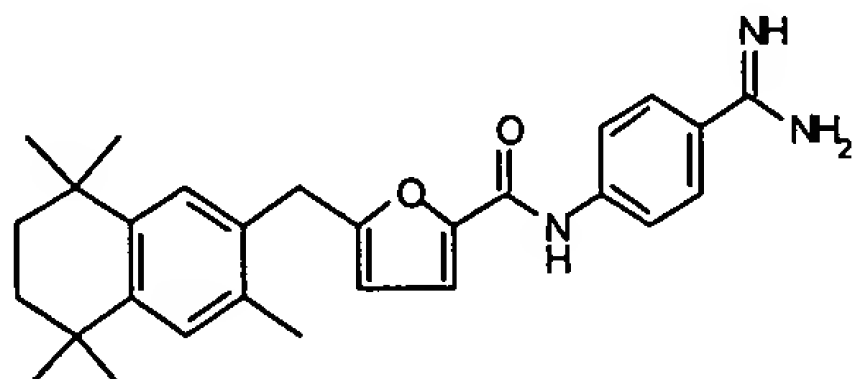
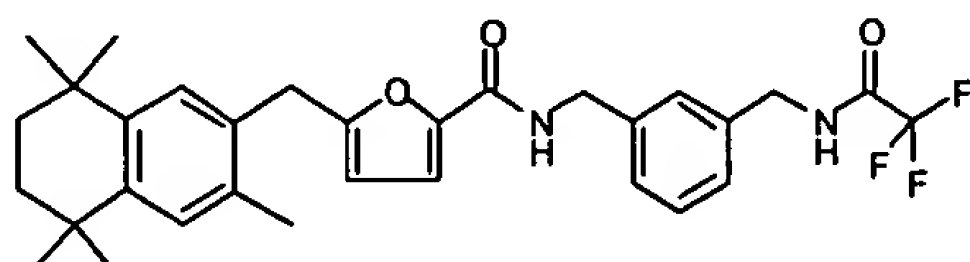
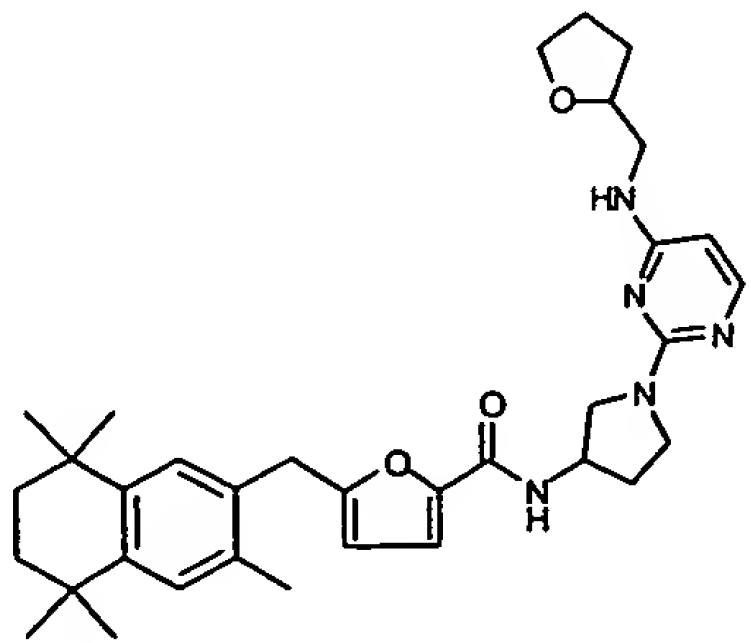
Cmpd. #	Structure	5-HT7 KI (nM)	5-HT2a KI (nM)	D2 DA KI (nM)	IL8 IC50 (nM)	NPY Y1 IC50 (nM)	Glucagon IC50 (nM)	GLP-1 IC50 (nM)
57	 Chiral							
58	 Chiral							
59								
60								
17								
18								
19								
61								

Cmpd. #	Structure	5-HT7 KI (nM)	5-HT2a KI (nM)	D2 DA KI (nM)	IL8 IC50 (nM)	NPY Y1 IC50 (nM)	Glucagon IC50 (nM)	GLP-1 IC50 (nM)
62								
20		>4400		>3200	>10000	>10000	6700	>10000
63								
64								
65								
66								
67								

Cmpd. #	Structure	5-HT7 Ki (nM)	5-HT2a Ki (nM)	D2 DA Ki (nM)	IL8 IC50 (nM)	NPY Y1 IC50 (nM)	Glucagon IC50 (nM)	GLP-1 IC50 (nM)
68								
69								
70								
71								
72								
73								
73								
75								

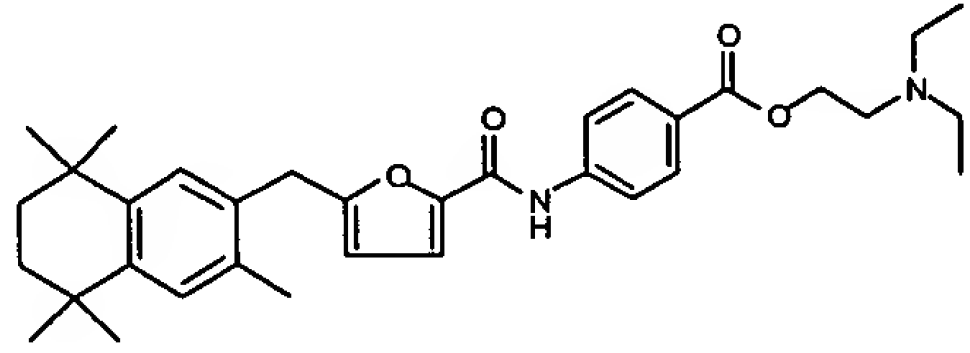
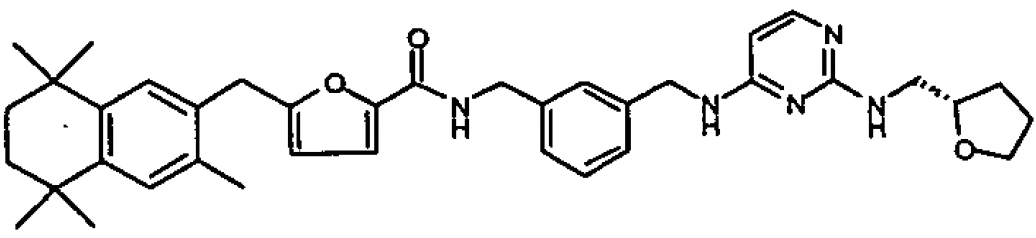
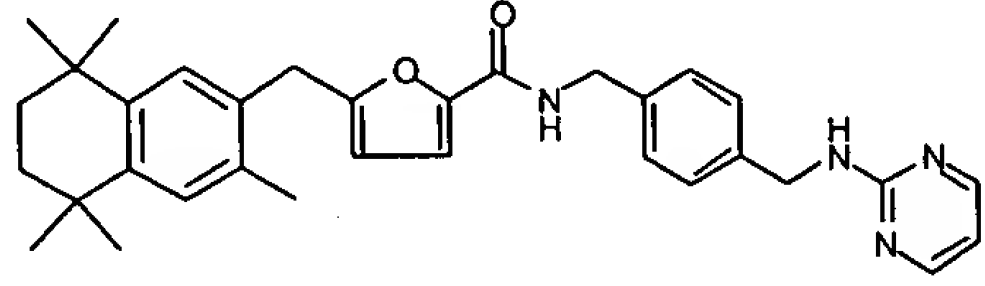
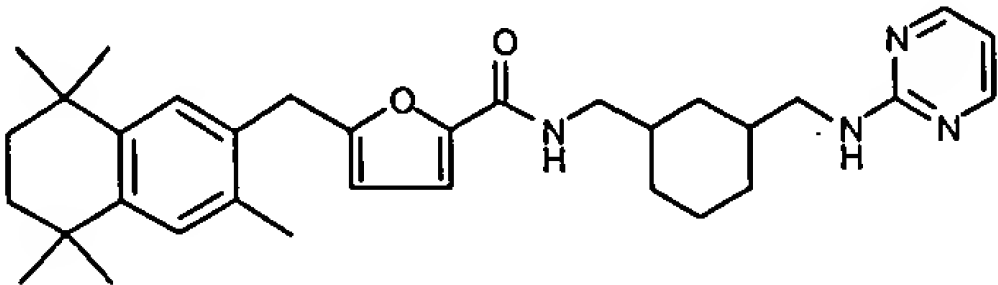
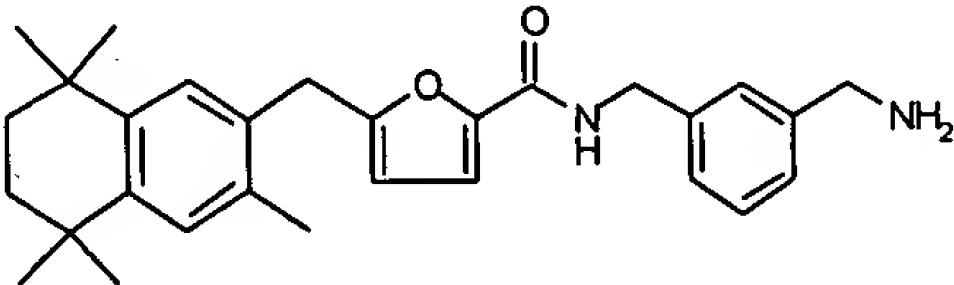
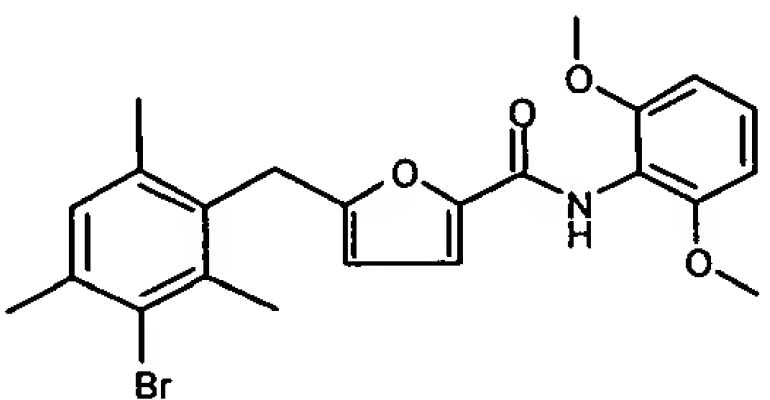
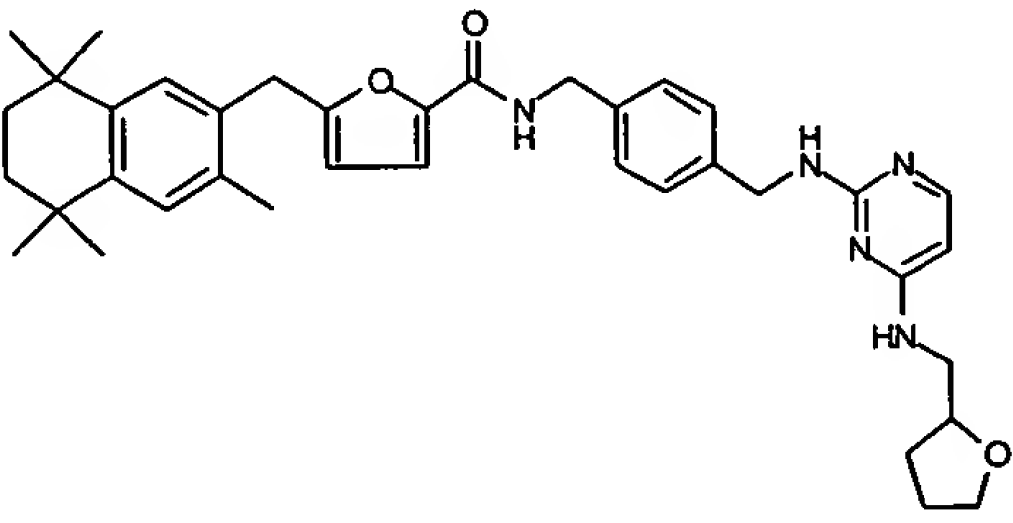
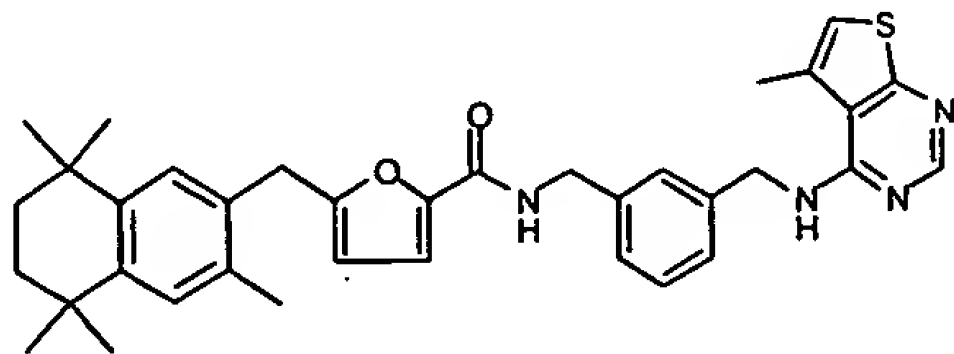
Cmpd. #	Structure	5-HT7 KI (nM)	5-HT2a KI (nM)	D2 DA KI (nM)	IL8 IC50 (nM)	NPY Y1 IC50 (nM)	Glucagon IC50 (nM)	GLP-1 IC50 (nM)
95								
96								
97								
98								
99								
100								
101								
102								

Cmpd. #	Structure	5-HT7 Ki (nM)	5-HT2a Ki (nM)	D2 DA Ki (nM)	IL8 IC50 (nM)	NPY Y1 IC50 (nM)	Glucagon IC50 (nM)	GLP-1 IC50 (nM)
103								
104								
105								
106								
107								
108								
109								

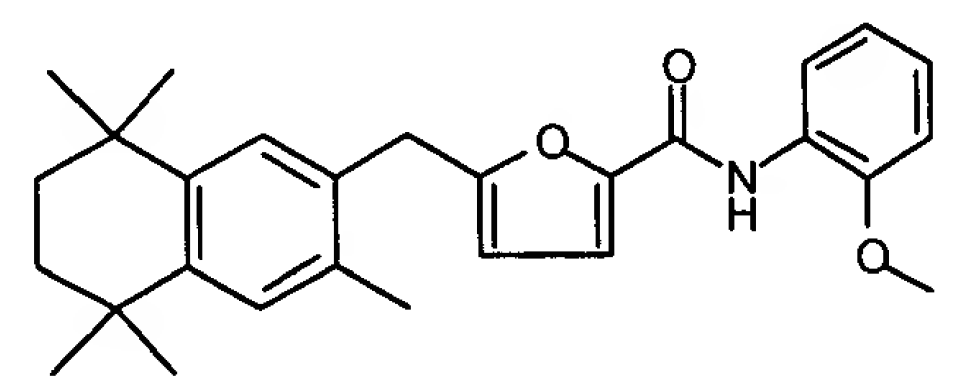
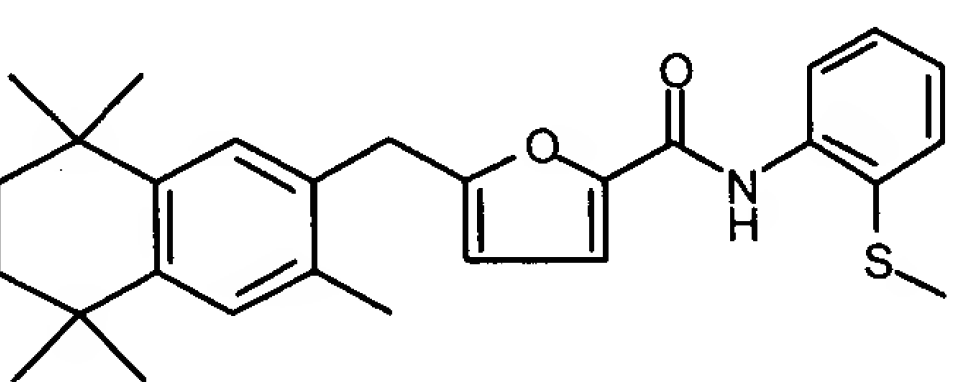
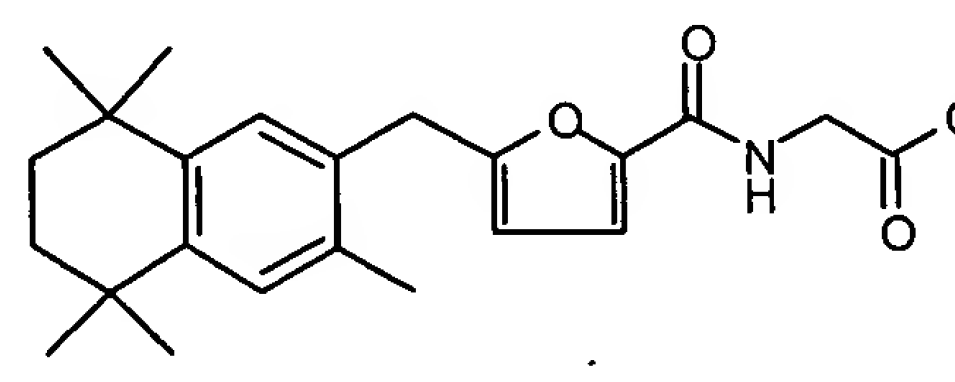
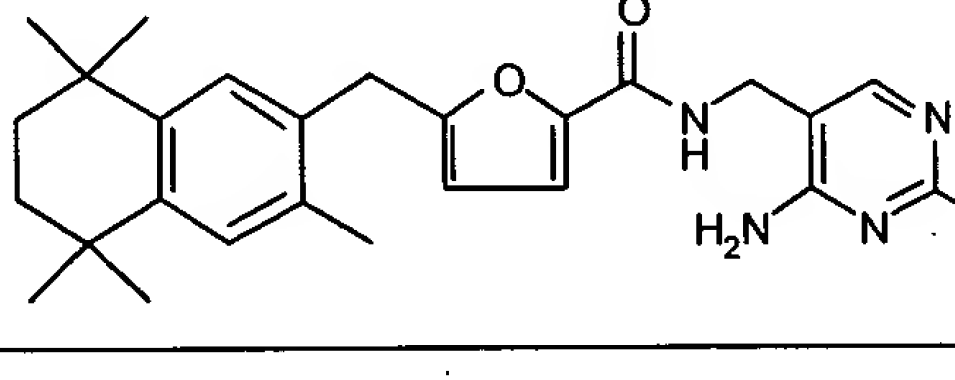
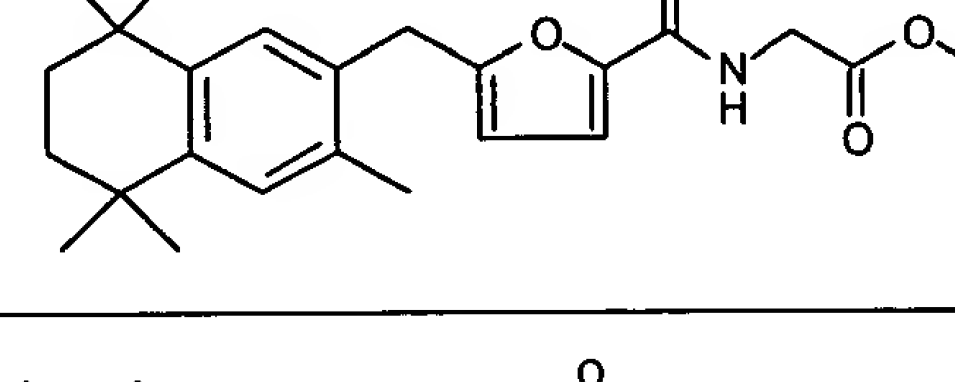
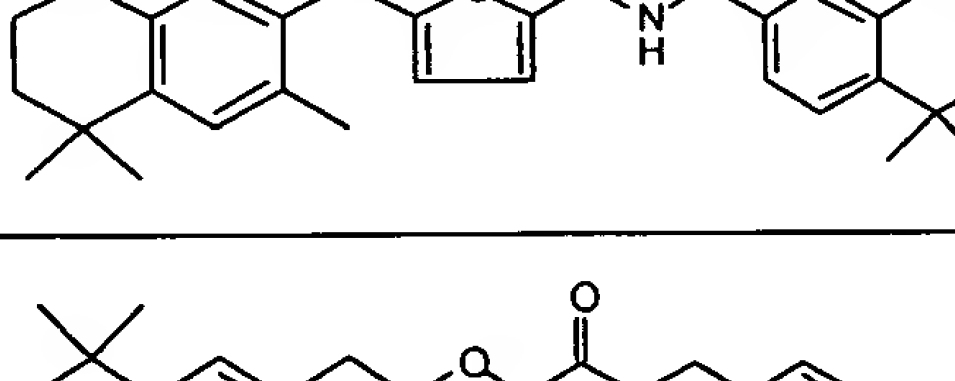
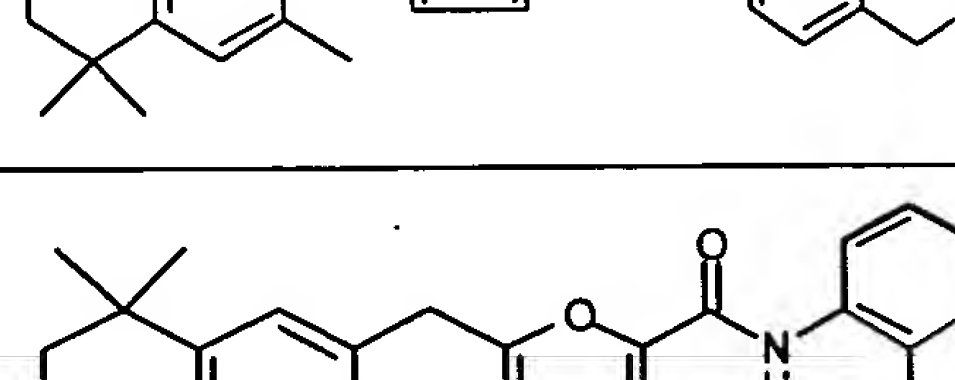

Cmpd. #	Structure	5-HT7 Ki (nM)	5-HT2a Ki (nM)	D2 DA Ki (nM)	IL8 IC50 (nM)	NPY Y1 IC50 (nM)	Glucagon IC50 (nM)	GLP-1 IC50 (nM)
110								
111								
112								
113								
114								
115								
116								
117								

Cmpd. #	Structure	5-HT7 Ki (nM)	5-HT2a Ki (nM)	D2 DA Ki (nM)	IL8 IC50 (nM)	NPY Y1 IC50 (nM)	Glucagon IC50 (nM)	GLP-1 IC50 (nM)
118								
119								
120								
121								
122								
123								
124								
125								

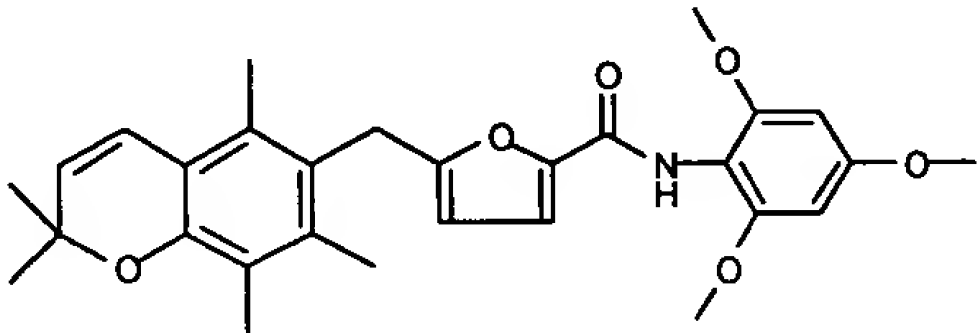
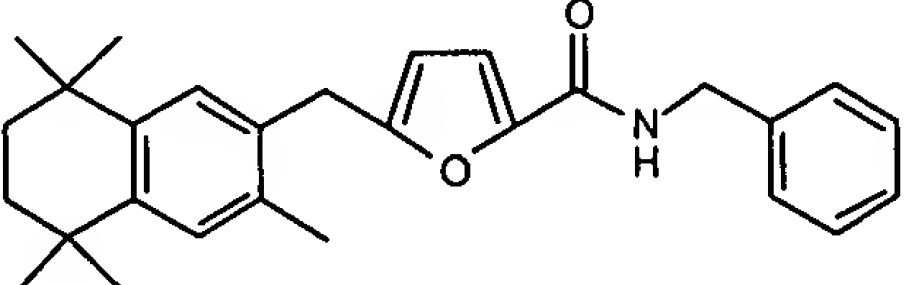
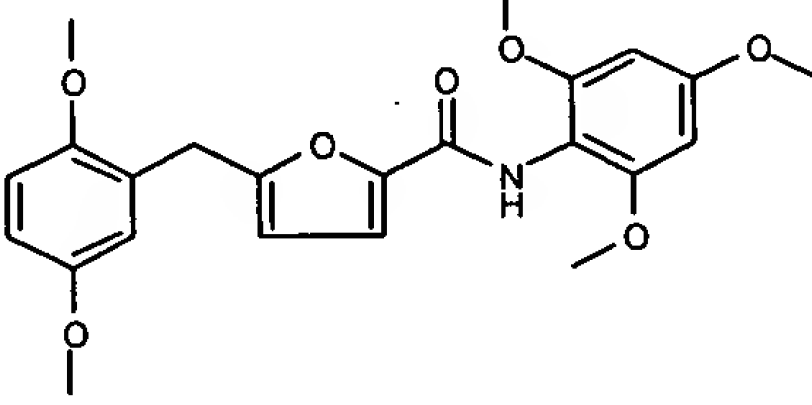
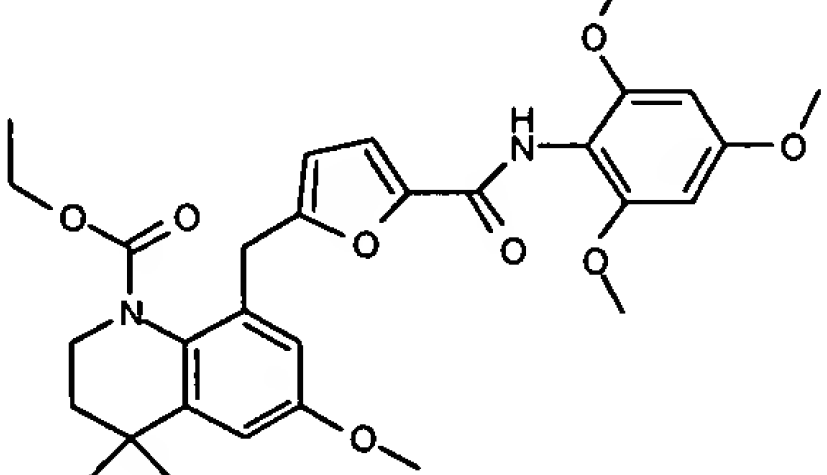
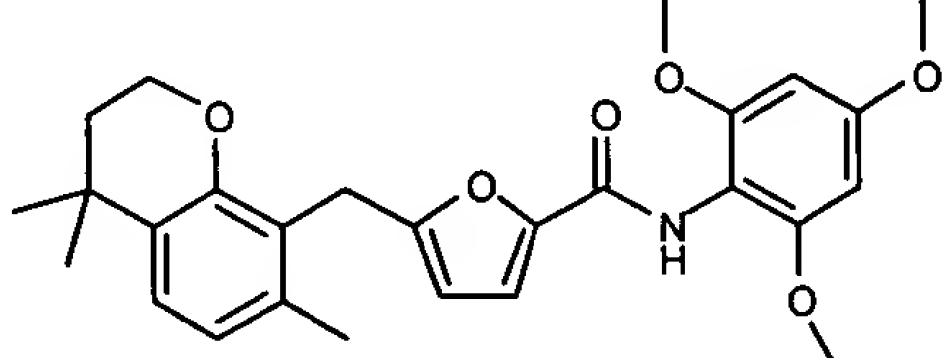
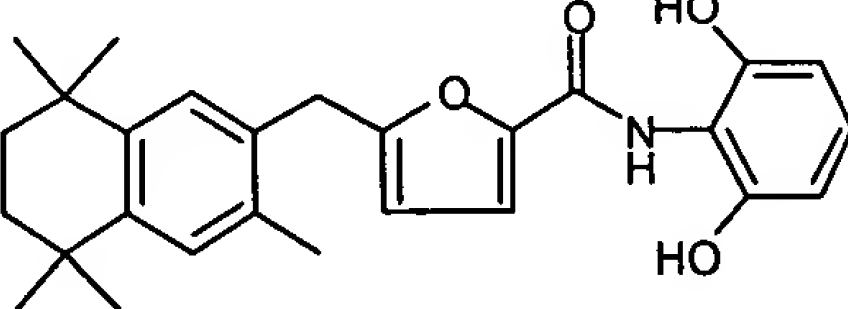
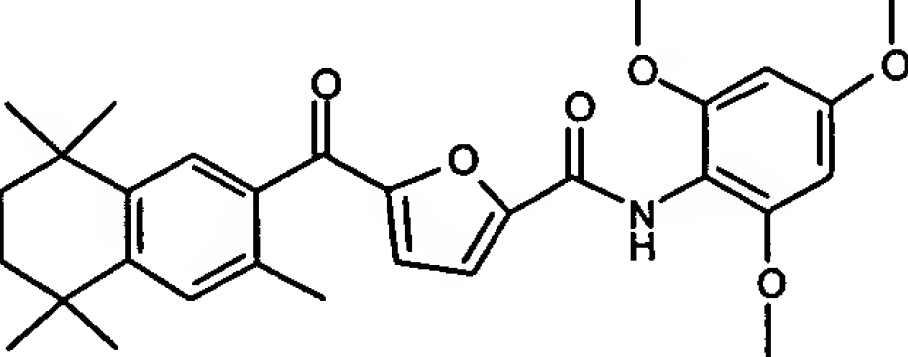
Cmpd. #	Structure	5-HT7 Ki (nM)	5-HT2a Ki (nM)	D2 DA Ki (nM)	IL8 IC50 (nM)	NPY Y1 IC50 (nM)	Glucagon IC50 (nM)	GLP-1 IC50 (nM)
126								
127								
128								
129								
130								
131								
132								
133								
134		>4400	3720	2400				

Cmpd. #	Structure	5-HT7 KI (nM)	5-HT2a KI (nM)	D2 DA KI (nM)	IL8 IC50 (nM)	NPY Y1 IC50 (nM)	Glucagon IC50 (nM)	GLP-1 IC50 (nM)
135								
136	 Chiral	2400		160				
137								
138								
139								
140	 Br	>4400		2300				
141								
142								

Cmpd. #	Structure	5-HT7 Ki (nM)	5-HT2a Ki (nM)	D2 DA Ki (nM)	IL8 IC50 (nM)	NPY Y1 IC50 (nM)	Glucagon IC50 (nM)	GLP-1 IC50 (nM)
143								
144								
145								
146								
147								
148								
149								
150								

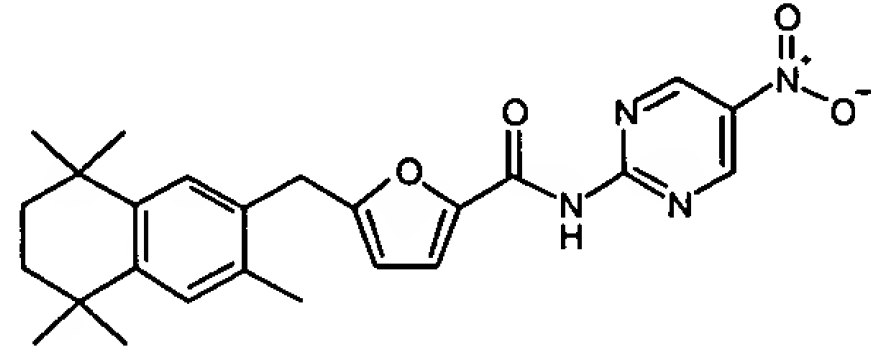
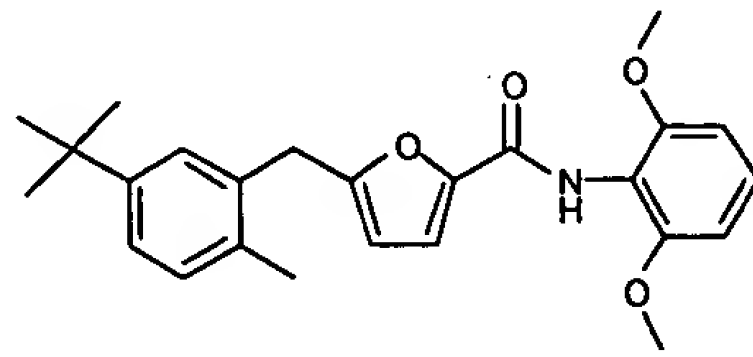
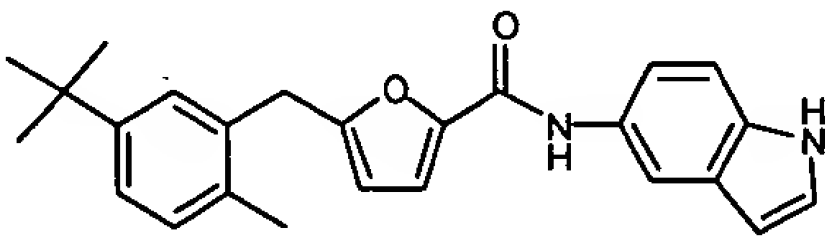
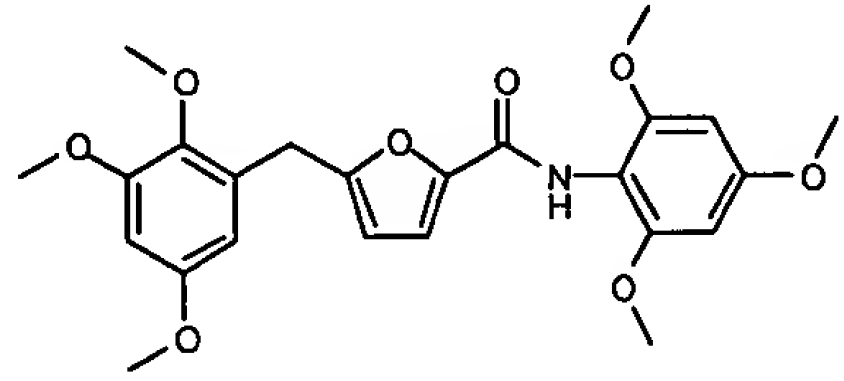
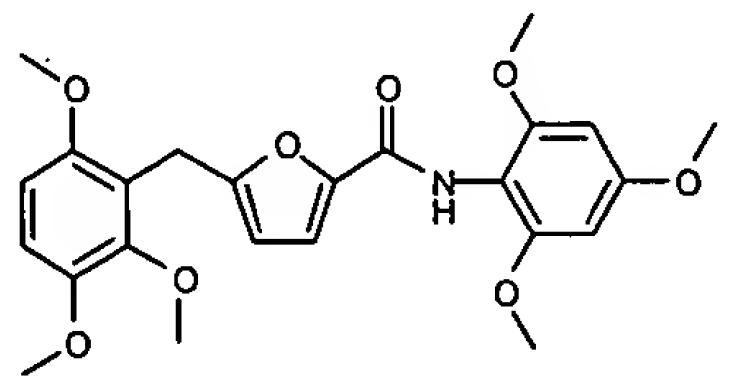
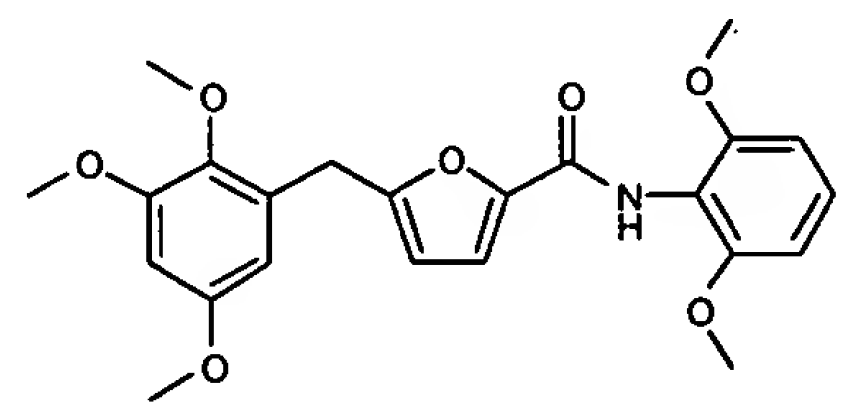
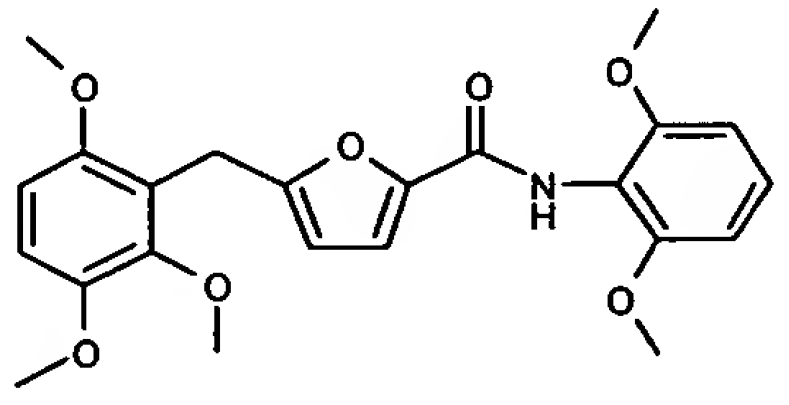
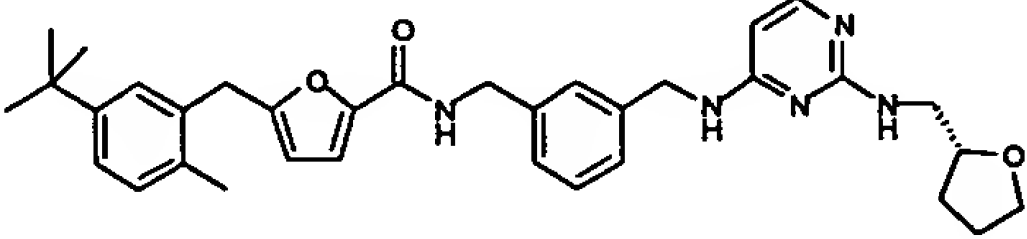
Cmpd. #	Structure	5-HT7 Ki (nM)	5-HT2a Ki (nM)	D2 DA Ki (nM)	IL8 IC50 (nM)	NPY Y1 IC50 (nM)	Glucagon IC50 (nM)	GLP-1 IC50 (nM)
160								
161								
162								
163								
164								
165								
166								
167								

Cmpd. #	Structure	5-HT7 KI (nM)	5-HT2a KI (nM)	D2 DA KI (nM)	IL8 IC50 (nM)	NPY Y1 IC50 (nM)	Glucagon IC50 (nM)	GLP-1 IC50 (nM)
168								
169			2710	3400				
170								
171								
172								
173								
174								
175								

Cmpd. #	Structure	5-HT7 KI (nM)	5-HT2a KI (nM)	D2 DA KI (nM)	IL8 IC50 (nM)	NPY Y1 IC50 (nM)	Glucagon IC50 (nM)	GLP-1 IC50 (nM)
184								
185								
186								
187								
188								
189								
190								

Cmpd. #	Structure	5-HT7 KI (nM)	5-HT2a KI (nM)	D2 DA KI (nM)	IL8 IC50 (nM)	NPY Y1 IC50 (nM)	Glucagon IC50 (nM)	GLP-1 IC50 (nM)
190.5								
191								
192								
193								
194								
195								
196								
197								

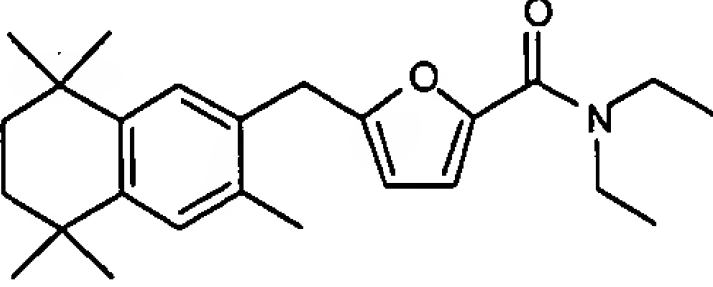
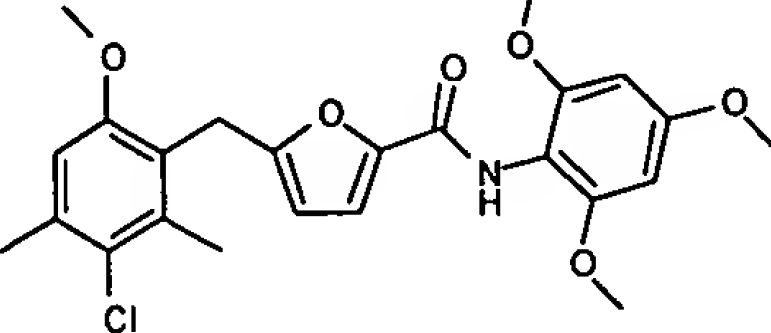
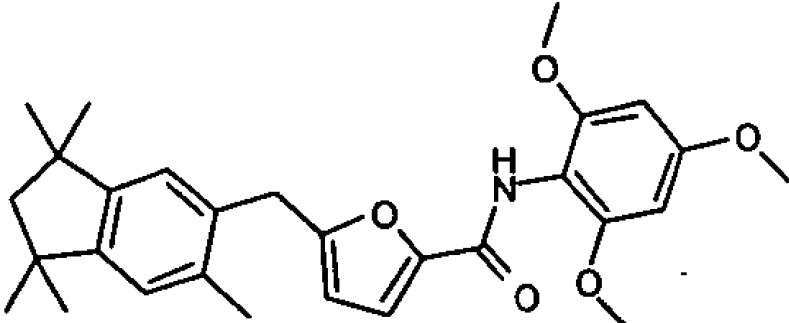
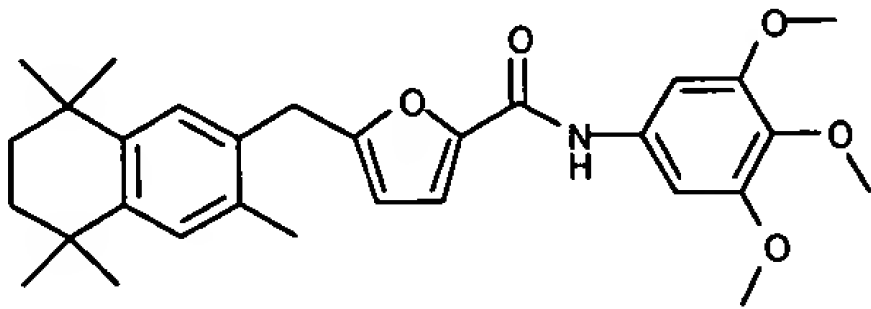
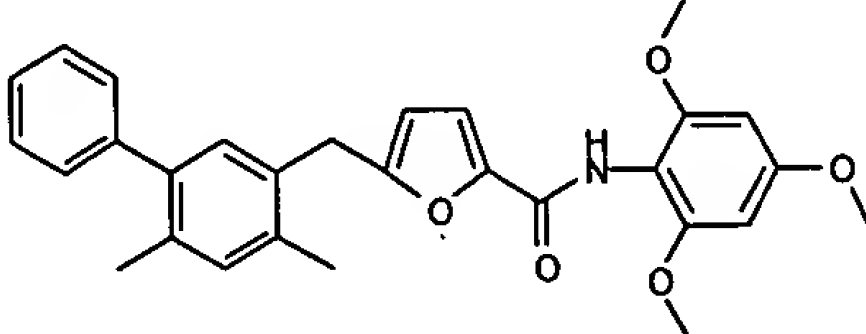
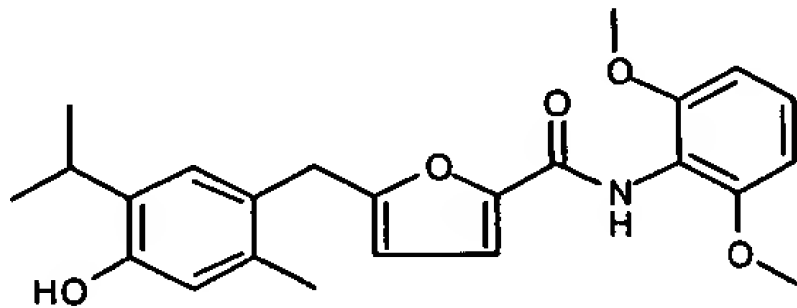
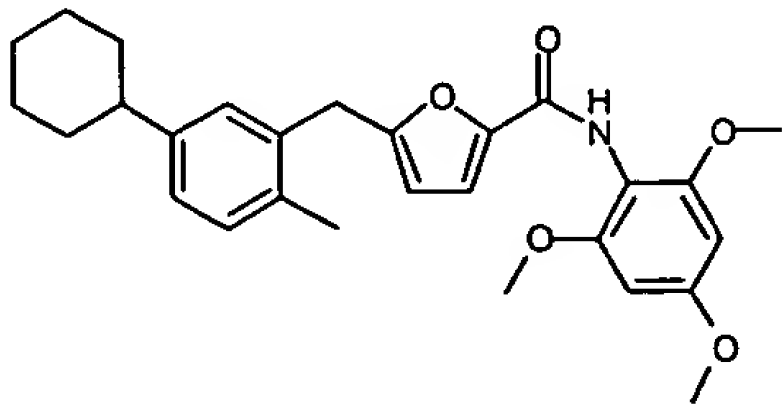
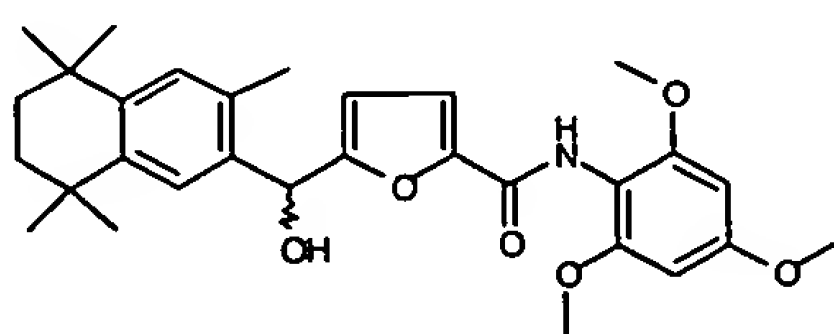
Cmpd. #	Structure	5-HT7 Ki (nM)	5-HT2a Ki (nM)	D2 DA Ki (nM)	IL8 IC50 (nM)	NPY Y1 IC50 (nM)	Glucagon IC50 (nM)	GLP-1 IC50 (nM)
198								
199								
200								
200.5								
201								
202								
203								
204								

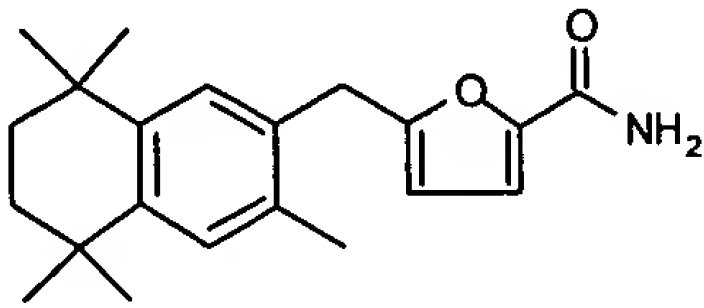
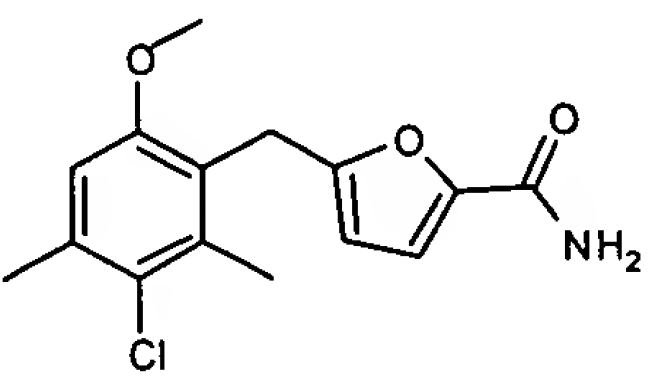
Cmpd. #	Structure	5-HT7 KI (nM)	5-HT2a KI (nM)	D2 DA KI (nM)	IL8 IC50 (nM)	NPY Y1 IC50 (nM)	Glucagon IC50 (nM)	GLP-1 IC50 (nM)
205								
206								
207								
208								
209								
210								
211								
212	 Chiral							

Cmpd. #	Structure	5-HT7 Ki (nM)	5-HT2a Ki (nM)	D2 DA Ki (nM)	IL8 IC50 (nM)	NPY Y1 IC50 (nM)	Glucagon IC50 (nM)	GLP-1 IC50 (nM)
213								
214								
215								
216								
217								
218								
219								
220								

Cmpd. #	Structure	5-HT7 KI (nM)	5-HT2a KI (nM)	D2 DA KI (nM)	IL8 IC50 (nM)	NPY Y1 IC50 (nM)	Glucagon IC50 (nM)	GLP-1 IC50 (nM)
221								
222								
223								
224								
225								
226								
227								
228								

Cmpd. #	Structure	5-HT7 KI (nM)	5-HT2a KI (nM)	D2 DA KI (nM)	IL8 IC50 (nM)	NPY Y1 IC50 (nM)	Glucagon IC50 (nM)	GLP-1 IC50 (nM)
229								
230								
231								
232								
233								
234								
235								
236								
237								

Cmpd. #	Structure	5-HT7 KI (nM)	5-HT2a KI (nM)	D2 DA KI (nM)	IL8 IC50 (nM)	NPY Y1 IC50 (nM)	Glucagon IC50 (nM)	GLP-1 IC50 (nM)
238								
299								
300								
301								
302								
303								
304								
AXC-07047								

Cmpd. #	Structure	5-HT7 Ki (nM)	5-HT2a Ki (nM)	D2 DA Ki (nM)	IL8 IC50 (nM)	NPY Y1 IC50 (nM)	Glucagon IC50 (nM)	GLP-1 IC50 (nM)
AXC-06675								
AXC-07450								
AXC-07350	